

Computational Structural Methods for Ligand (Drug) Discovery

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NCI RAS Initiative

Guest lecture for MSB 530 Bioinformatics

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The hook: Why you should care

- Drugs impact human health and quality of life!
- Drug discovery is a long and expensive process.
- Almost all recent drugs have been touched by computational methods: molecular graphics, molecular docking, free energy calculations ...

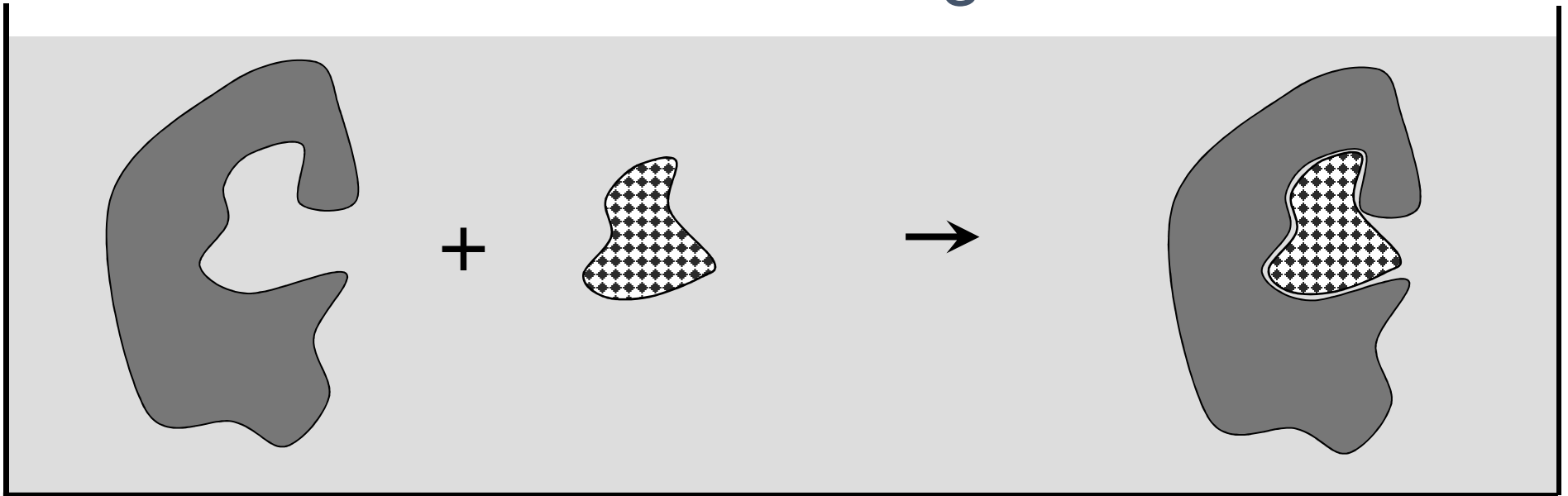
Outline

- The binding event using a thermodynamic cycle
- Introduction to molecular mechanics
- Introduction to molecular dynamics
- Introduction to molecular docking
- Applications from my work
 - Large-scale docking
 - Receptor flexibility
 - Receptor desolvation

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Molecular Recognition



- Understanding the binding event -- important for drug discovery
- Structure-based, Targeted drug discovery
- Computational methods

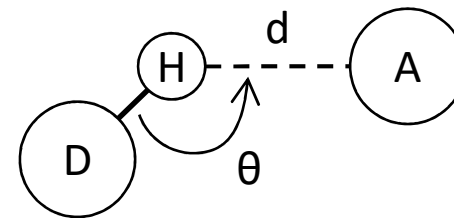
Important molecular forces for ligand binding

- Electrostatics
- Van der Waals
- Hydrogen bond
- Dipolar interactions
- Quadrupole interactions

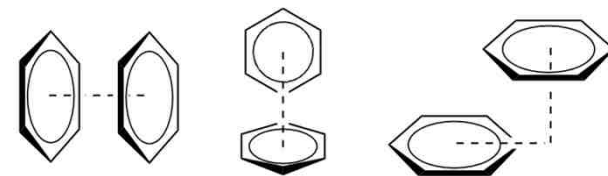
- Interactions with water (solvent)
 - Hydrophobic effect

- Entropy

Hydrogen bond



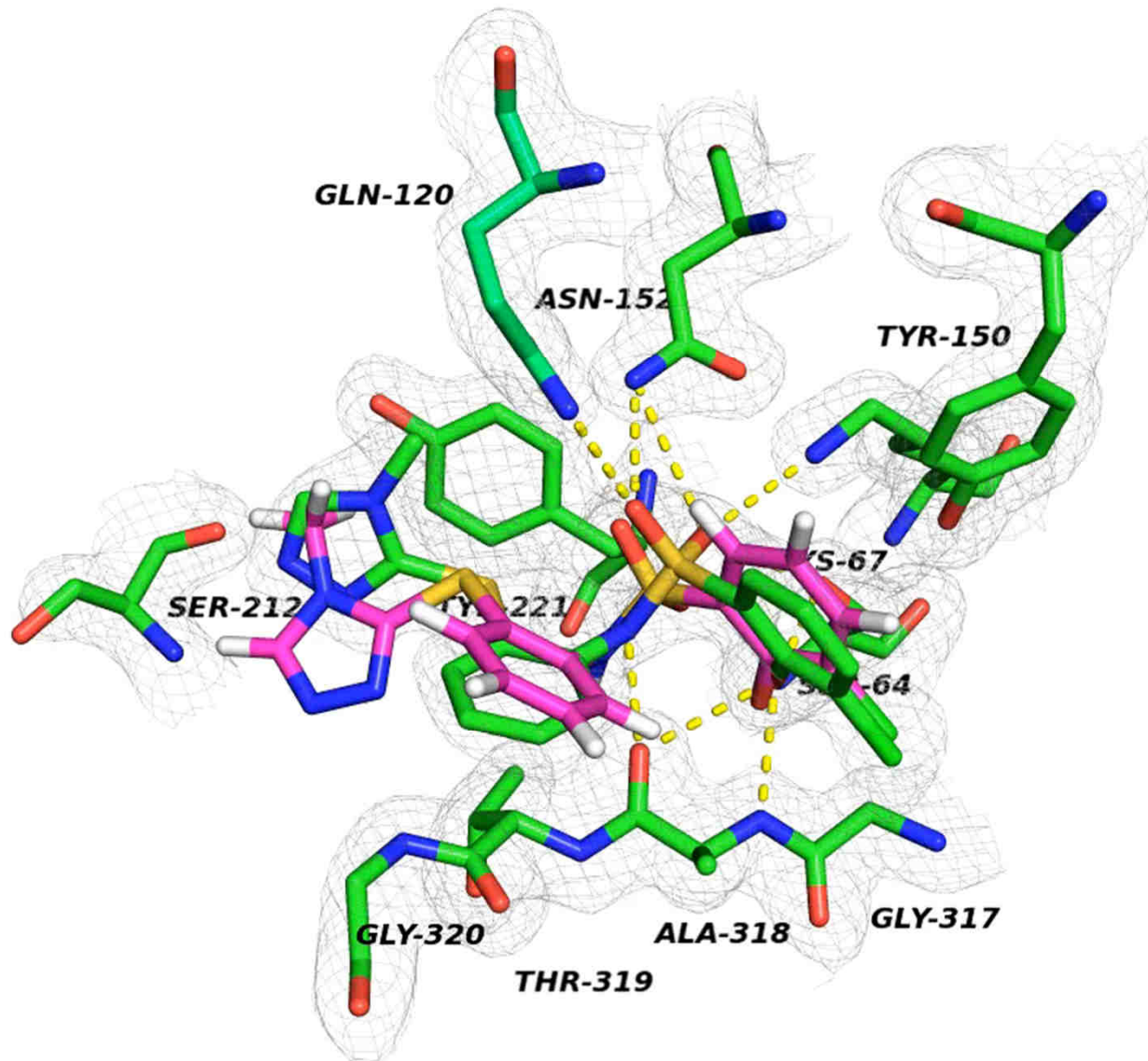
Pi-stacking



Sandwich T-shaped Parallel-displaced

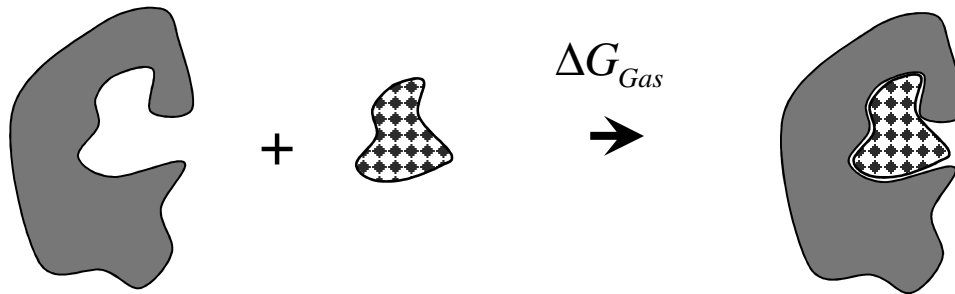
By Emily ricq - Own work, CC BY-SA 3.0,
<https://commons.wikimedia.org/w/index.php?curid=17057524>

Molecular Recognition



Critical role of water in receptor-ligand binding

Thermodynamic cycle of the binding event

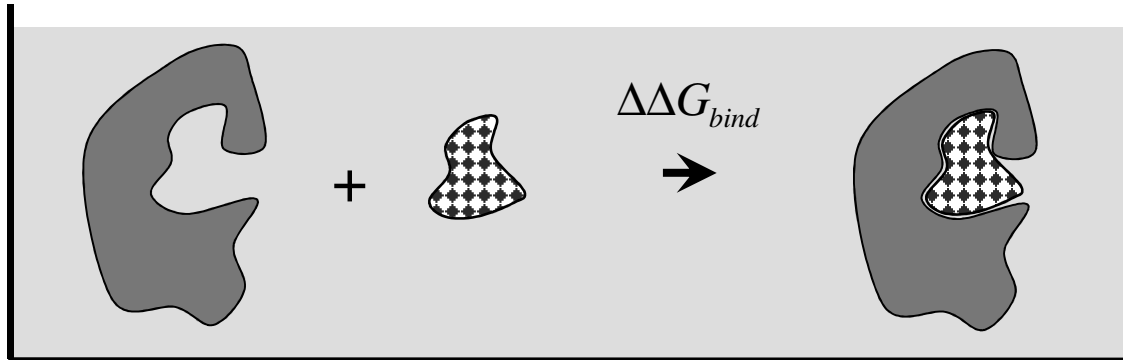


$$\Delta G_{gas} = \Delta G_{VDW} + \Delta G_{ES}$$

$$\downarrow \Delta G_{solv}^{Rec}$$

$$\downarrow \Delta G_{solv}^{Lig}$$

$$\downarrow \Delta G_{solv}^{Com}$$



$$\Delta G_{desolv} = \Delta G_{solv}^{Com} - (\Delta G_{solv}^{Lig} + \Delta G_{solv}^{Rec})$$

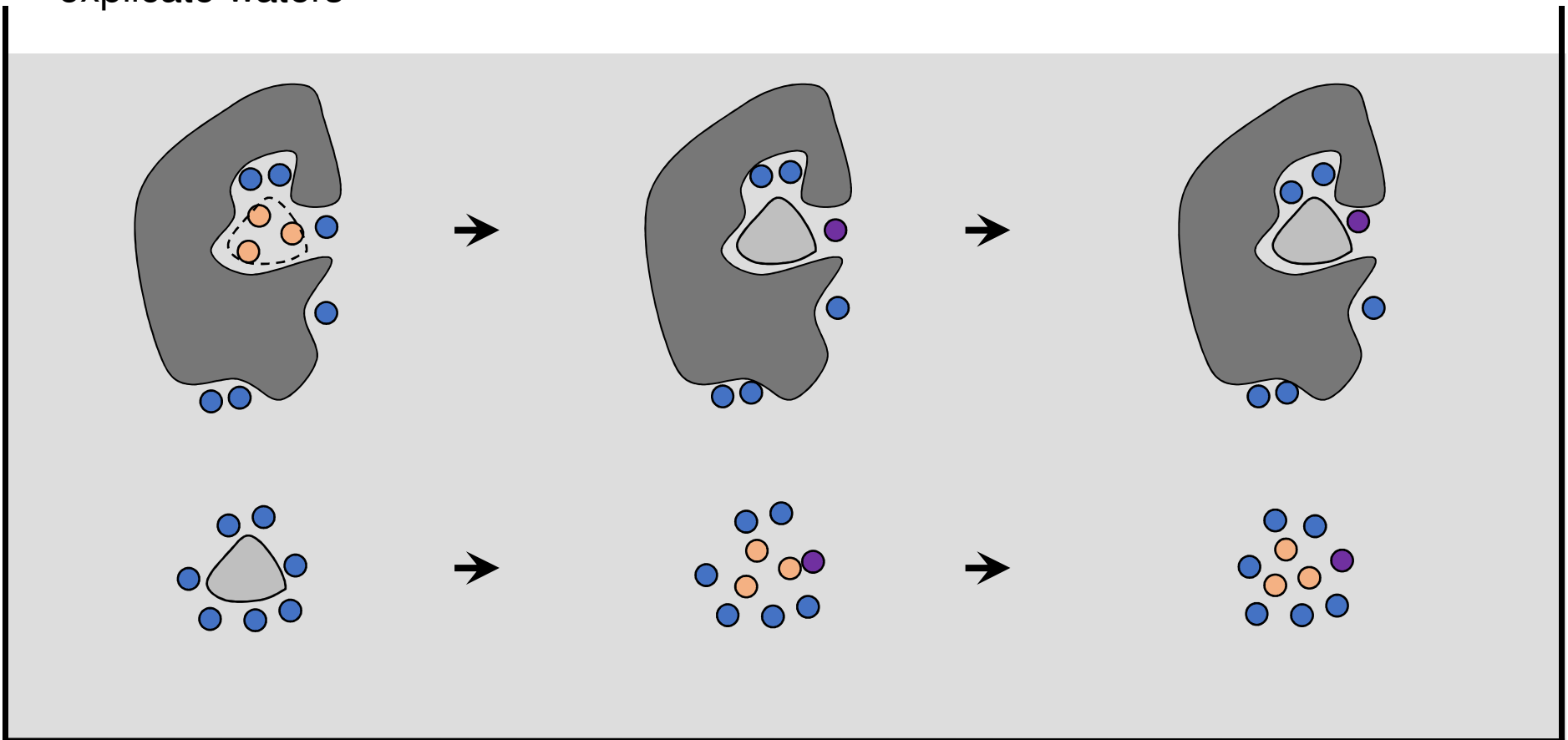
$$\Delta \Delta G_{bind} = \Delta G_{Gas} + \Delta G_{desolv}$$

Water displacement and medicated interactions

Receptor and ligand hydrated with explicit waters

Ligands and receptor Displaces waters.

Waters adjust to new environment



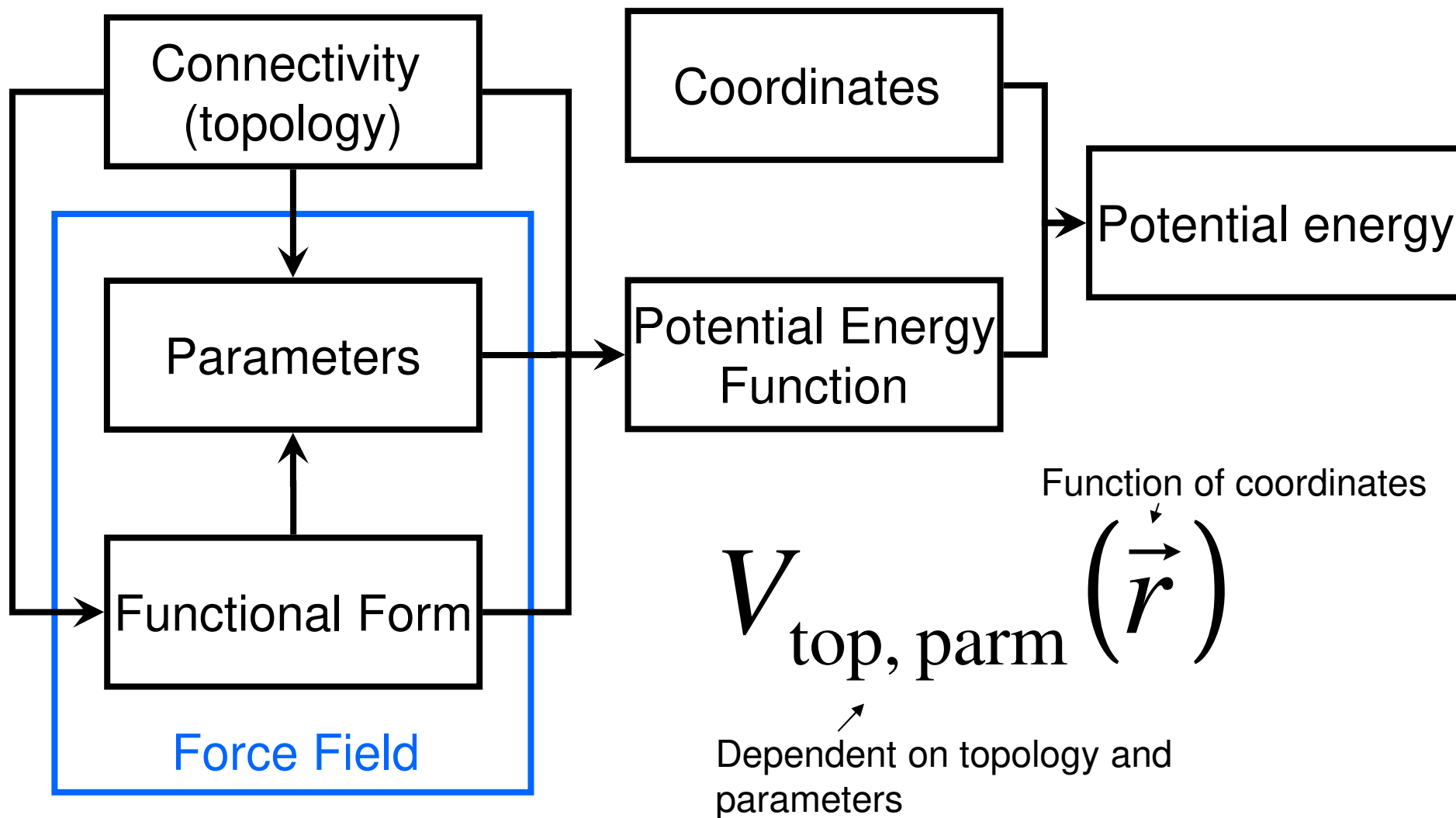
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Molecular Models

- Quantum Mechanics
 - physical, but expensive
 - Schrödinger equation: $H \Psi = E \Psi$
 - wave functions defines electron density
- Molecular Mechanics
 - less physical --> empirical parameterization
 - cheap and accurate

Molecular Mechanics Force Field



Molecular Mechanics

- Every atom is a sphere with a radius (Lennard Jones)
- Point charge is located at each atomic center (Coulomb's law)
- Bonds and angles are held by springs to ideal lengths
 - e.g. $V_{bond} = k_b (r - r_0)^2$
 - Hooke's Law, K_b : spring constant, r_0 : ideal length
- Dihedrals are represented by sigmoidal function which has energy wells at favorable angles.
- Improper torsions force atoms to be a defined angle to plane.

The "Tinker-toy Model"

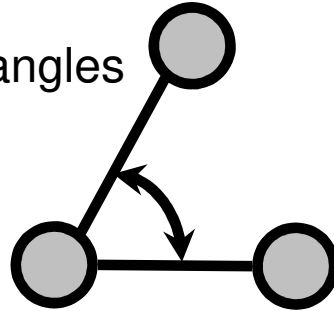
Bonded terms

bonds



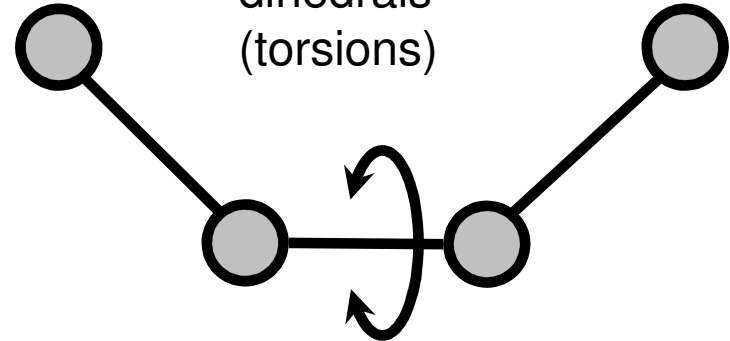
$$V_{bond} = k_b (r - r_0)^2$$

angles



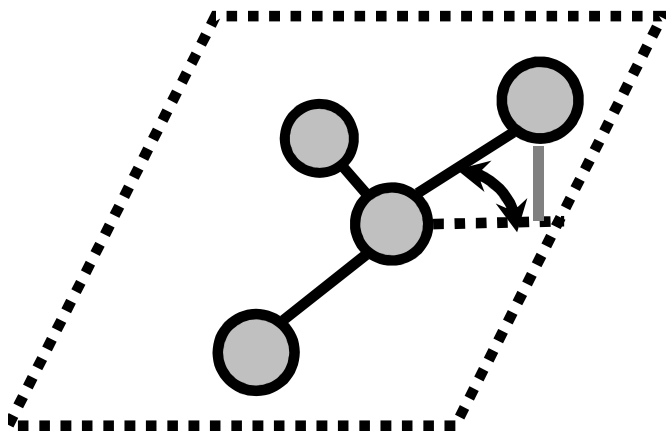
$$V_{angle} = k_\theta (\theta - \theta_0)^2$$

dihedrals
(torsions)

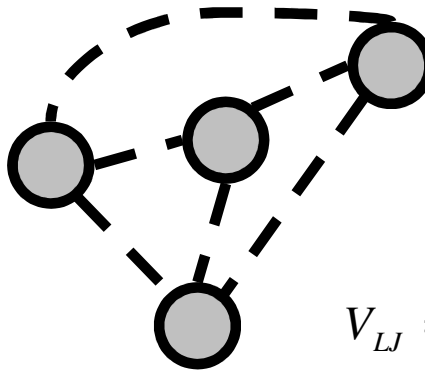


$$V_{dihedral} = K_\chi (1 + \cos(n\chi - \delta))$$

improper



$$V_{improper} = k_\varphi (\varphi - \varphi_0)^2$$



Through space
interactions

$$V_{coul} = \frac{q_i q_j}{\epsilon r_{ij}}$$

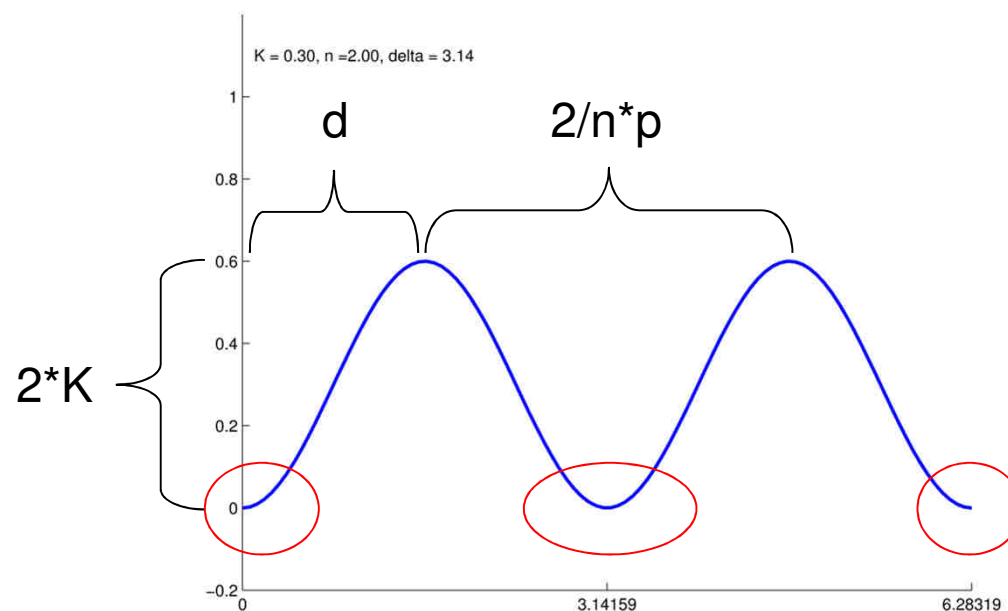
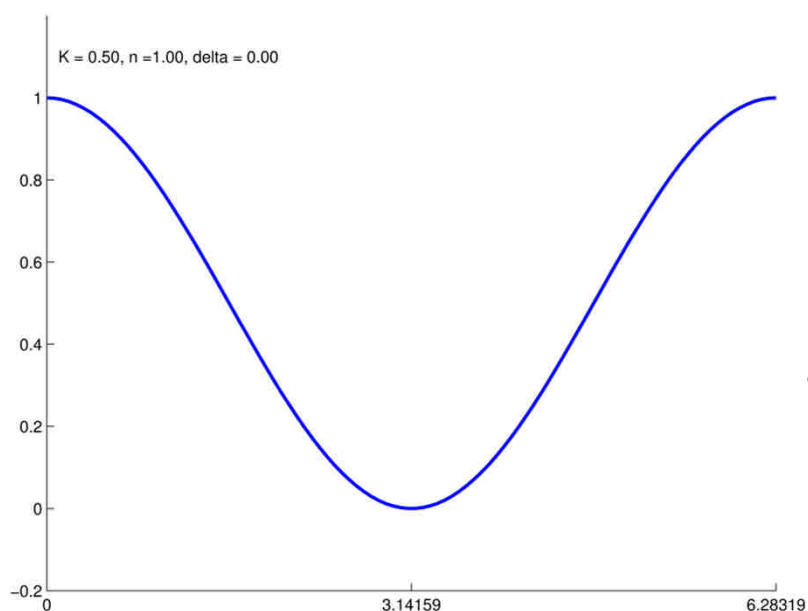
$$V_{LJ} = \epsilon_{i,j} \left[\left(\frac{R \min_{i,j}}{r_{i,j}} \right)^{12} - 2 \left(\frac{R \min_{i,j}}{r_{i,j}} \right)^6 \right]$$

$$R \min_{i,j} = \frac{R \min_i + R \min_j}{2}$$

$$\epsilon_{i,j} = \sqrt{\epsilon_i * \epsilon_j}$$

Dihedral Term is a Sigmodal Function

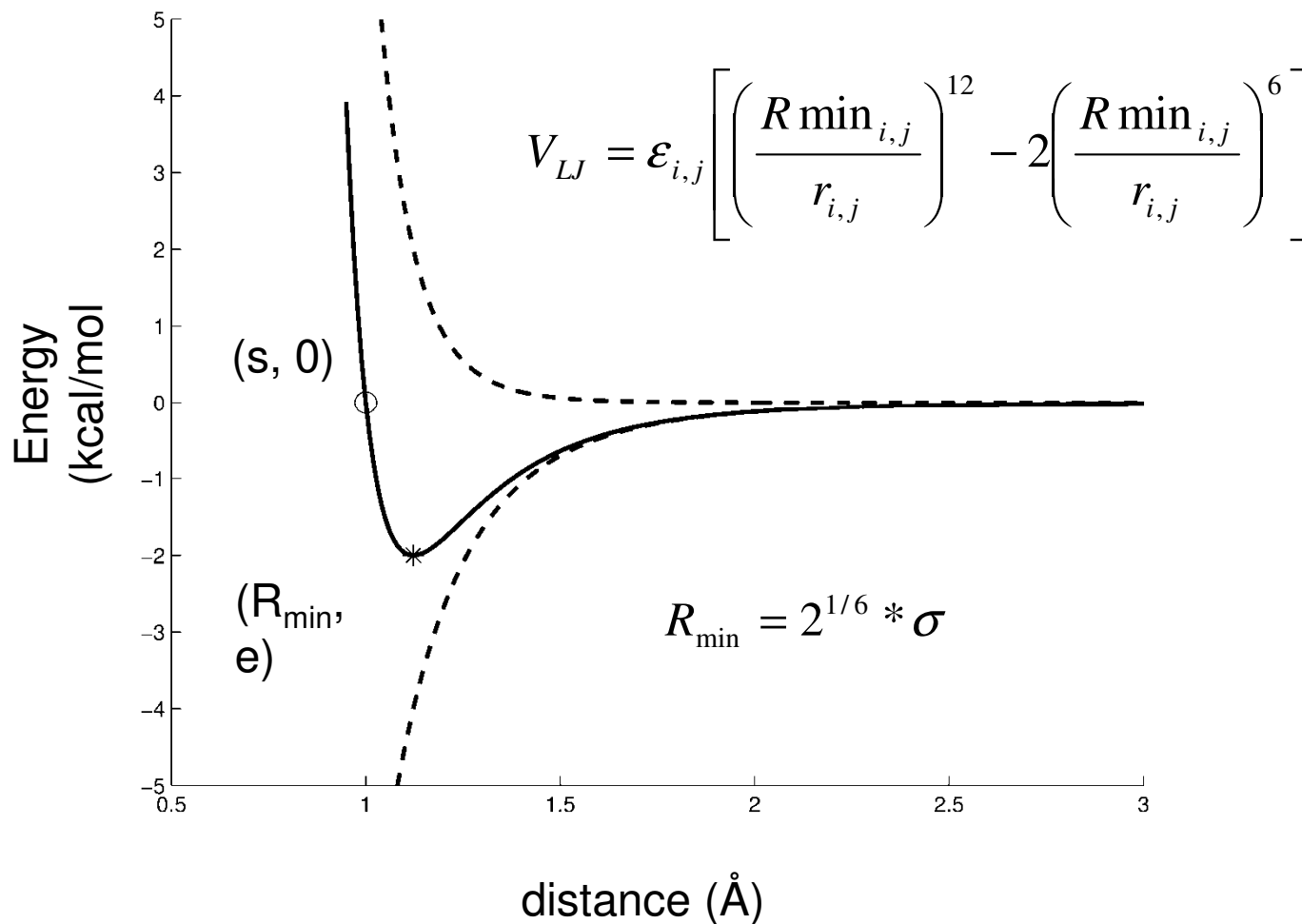
$$V_{dihedral} = K_{\chi} (1 + \cos(n\chi - \delta))$$



p (3.14) radians = 180 degrees

Molecular Modelling Principles and applications,
Leach Pearson Prentice hall second edition (chapter 4)

Lennard-Jones Equation



Molecular Modelling Principles and applications,
Leach Pearson Prentice hall second edition (chapter 4)

Potential Energy Function

$$V = \sum_{\text{bonds}} k_b (b - b_0)^2 + \sum_{\text{angles}} k_\theta (\theta - \theta_0)^2 + \sum_{\text{impropers}} k_\phi (\phi - \phi_0)^2 \\ + \sum_{\text{dihedrals}} K_\chi (1 + \cos(n\chi - \delta)) + \sum_{i=1}^N \sum_{j=i+1}^N \epsilon_{i,j} \left[\left(\frac{R \min_{i,j}}{r_{i,j}} \right)^{12} - 2 \left(\frac{R \min_{i,j}}{r_{i,j}} \right)^6 \right] + \frac{q_i q_j}{\epsilon r_{ij}}$$

Different Force-Field

- CHARMM
- AMBER
- GROMOS
- OPLS

Parameterization

- Experimental observables
- Quantum Mechanical calculations

Interdependences among parameters

Molecular Modelling Principles and applications,
Leach Pearson Prentice hall second edition (chapter 4)

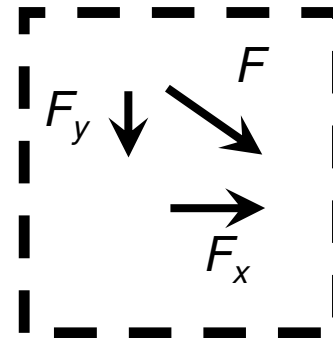
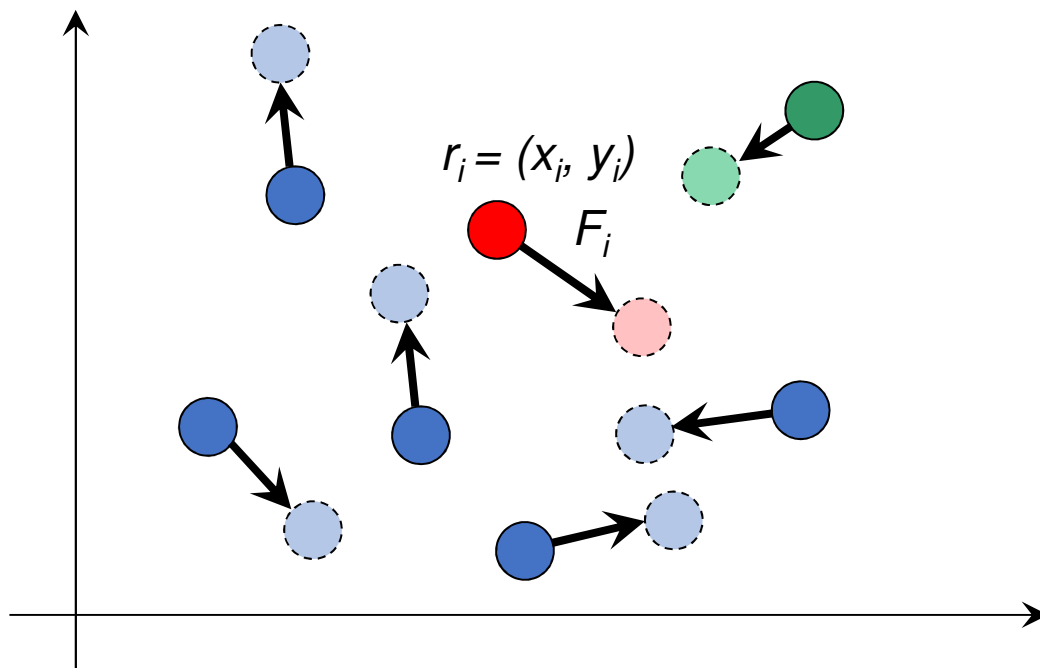
Mackerell, Vol. 25, No. 13, Journal of Computational Chemistry

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Molecular Dynamics

$$\begin{aligned} E(\vec{r}) \\ \vec{F}_i &= -\nabla_i E \\ \vec{a}_i &= \vec{F}_i / m_i \\ \vec{r}_{n,i} &= \vec{r}_i + \iint \vec{a}_i dt \end{aligned} \quad \left[\vec{F}_i = -\nabla_i E_{top,param}(\vec{r}) = - \begin{bmatrix} \frac{\partial}{\partial x_i} E_{top,param}(\vec{r}) \\ \frac{\partial}{\partial y_i} E_{top,param}(\vec{r}) \end{bmatrix} \right]$$



Molecular Dynamics

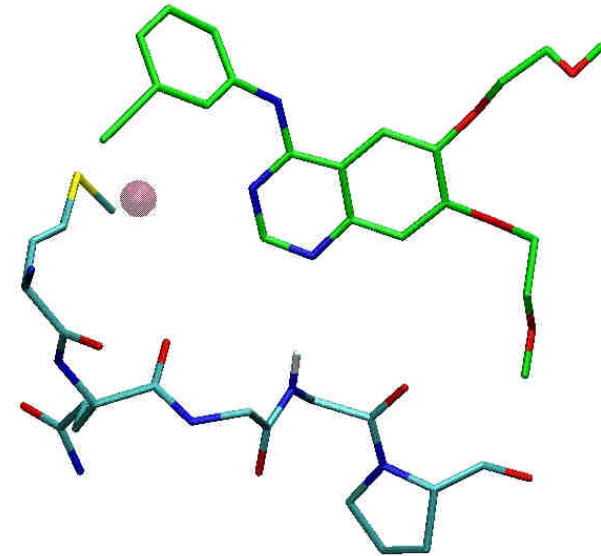
- Newton Equations

$$E(\mathbf{X}_{\text{position}})$$

$$\mathbf{F} = -\nabla E$$

$$\mathbf{X}_{\text{position}}^{\text{new}} = \iint \frac{\mathbf{F}}{m} \partial t^2$$

- Differential Eq.
(velocity verlet algorithm)
 - propagate to get motion



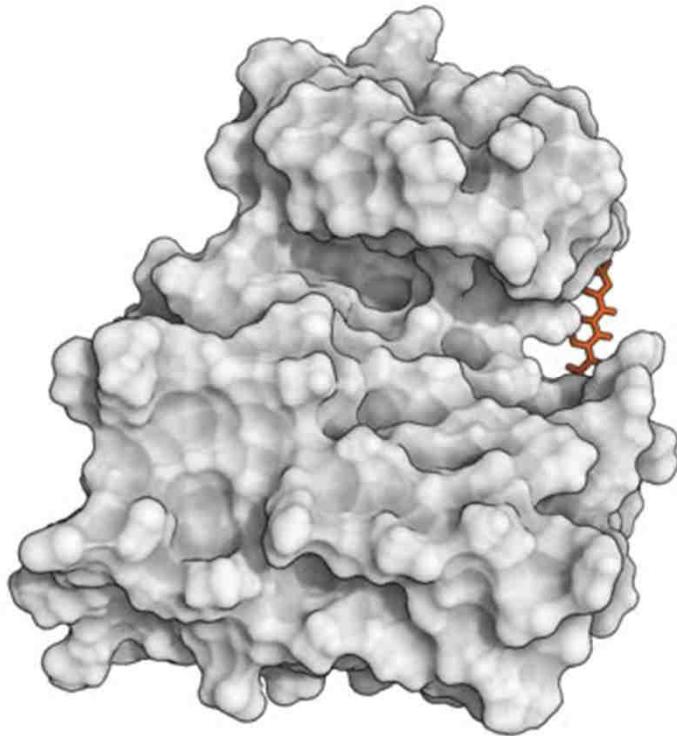
2 fs time step

- Energy functions:

$$E = E_{\text{bonded}} + E_{\text{steric}} + E_{\text{elect}}$$

$$E_{\text{bonded}} = E_{\text{bond}} + E_{\text{angle}} + E_{\text{dihedral}}$$

Simulate Binding of Ligand to Protein



4- μ s simulation of dasatinib binding to Src kinase;

Binding occurs 2.5 μ s into the simulation (in the 7th second of the movie)

2 fs = 1 time step

1s = 1,000,000 μ s

4 μ s = 2 billion time steps

Some Applications for Modular Dynamics

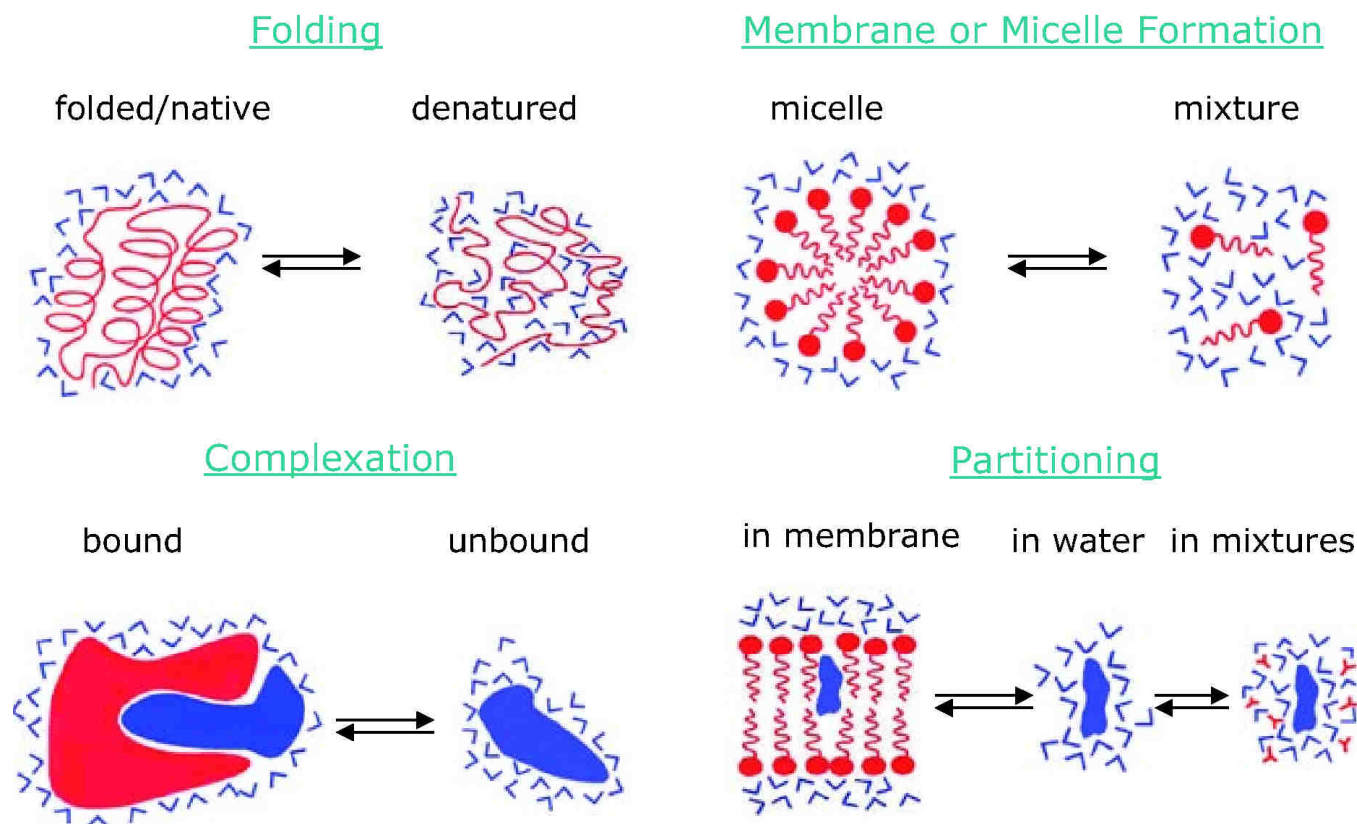


Figure 1. Four biomolecular processes that are governed by thermodynamic equilibria.

Things to Consider for Molecular Simulations

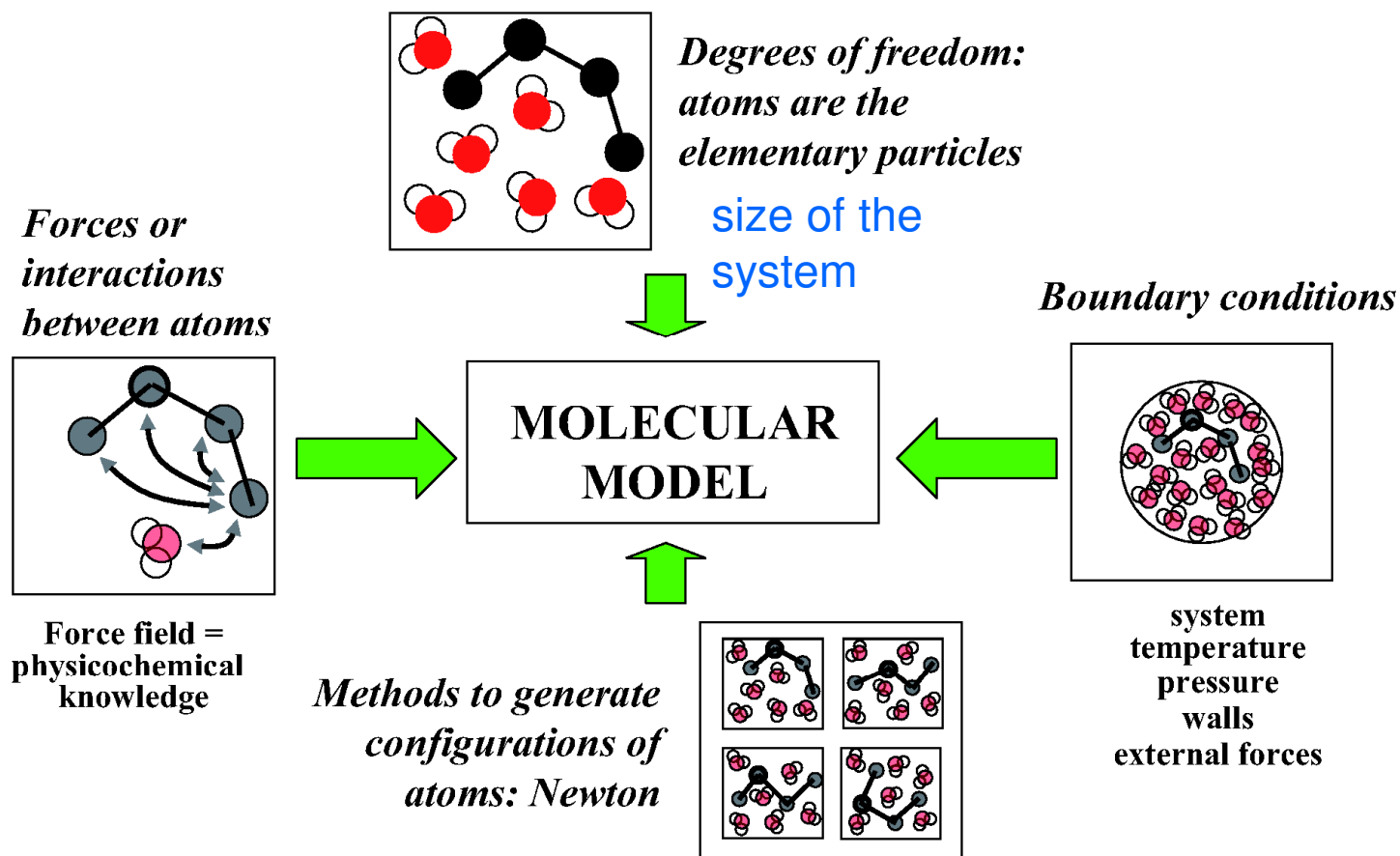
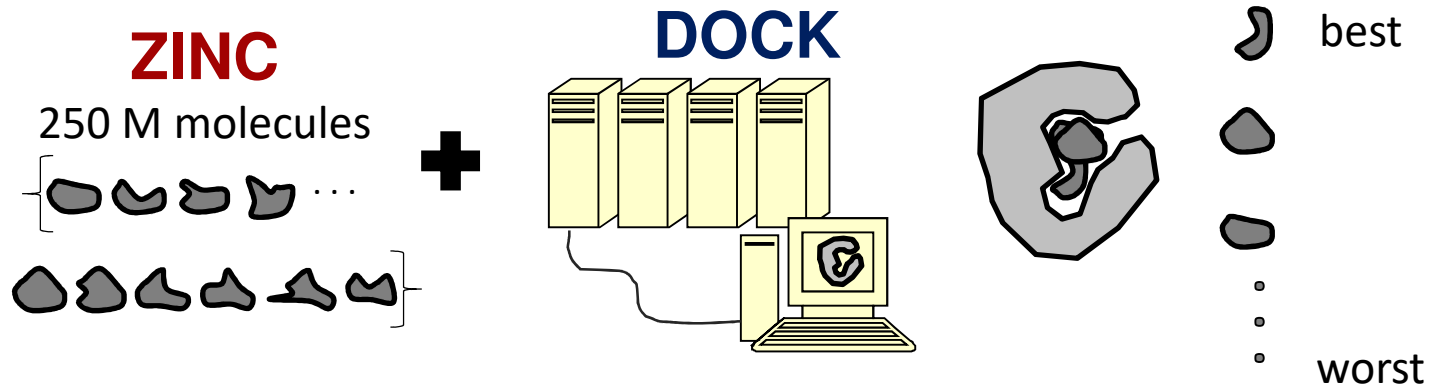


Figure 2. Four basic choices in the definition of a model for molecular simulation.

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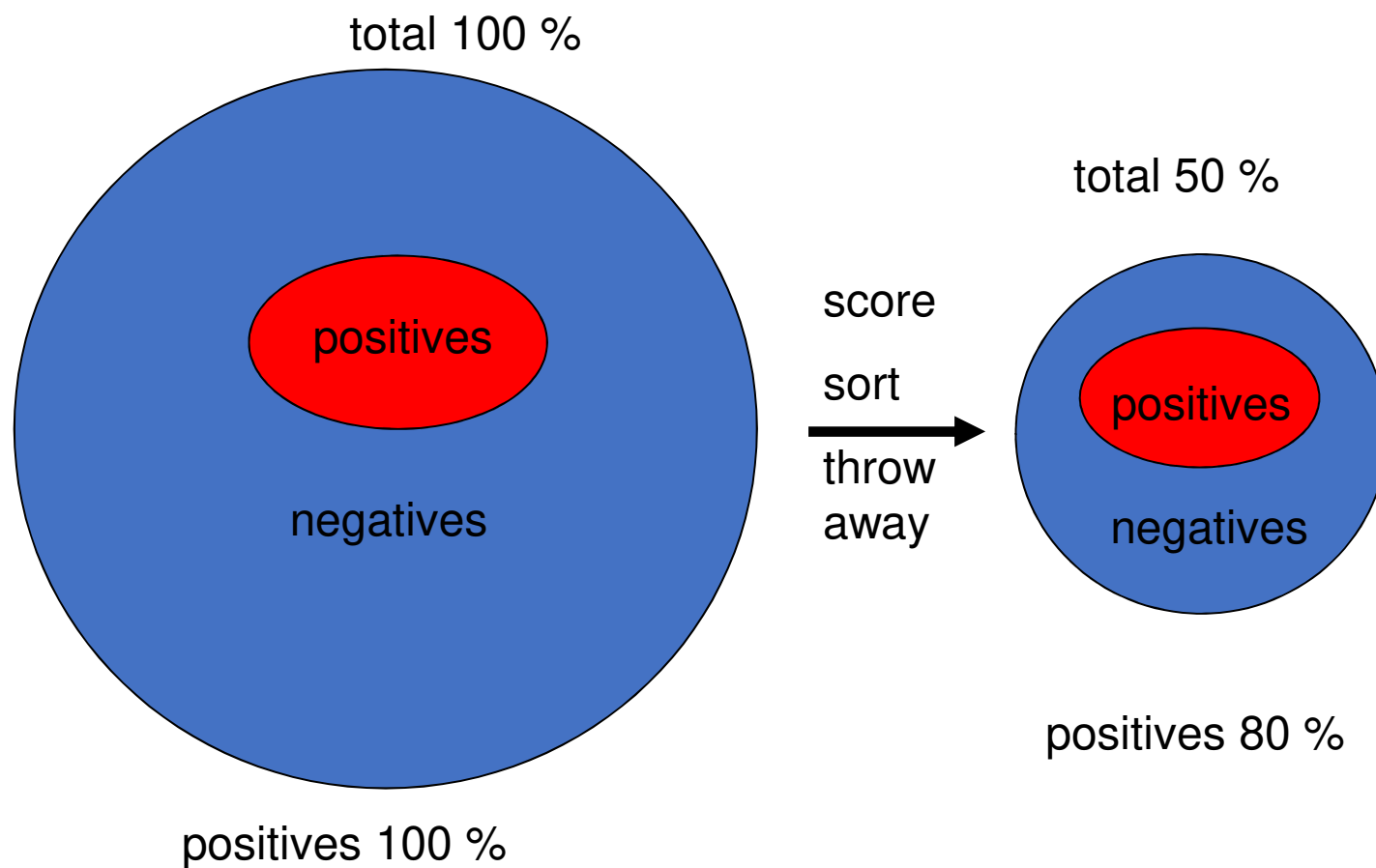
Docking is ligand discovery tool



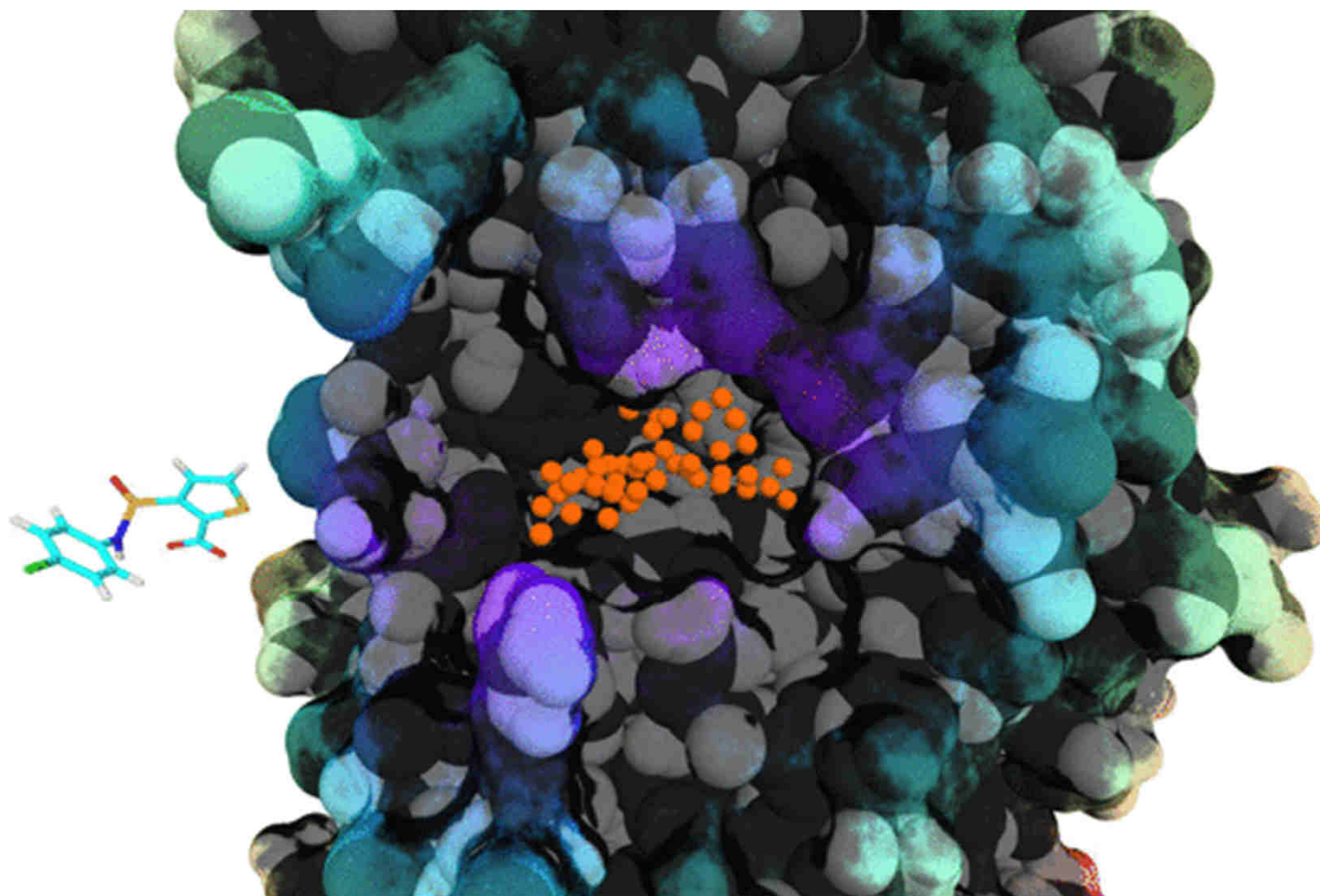
Docking Tasks

- Sampling
- Scoring
- Balance of speed and accuracy

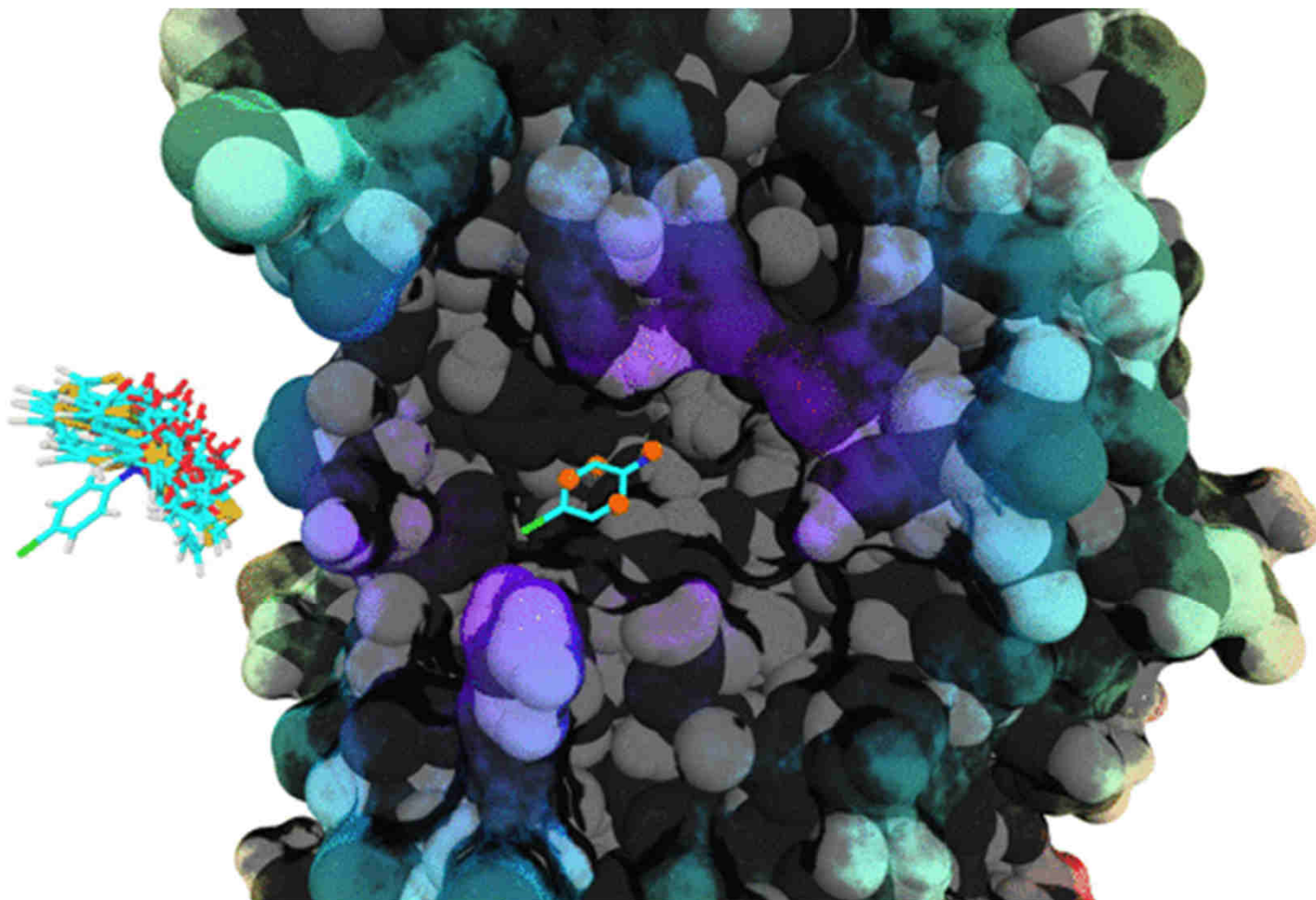
Enrichment for positives through docking



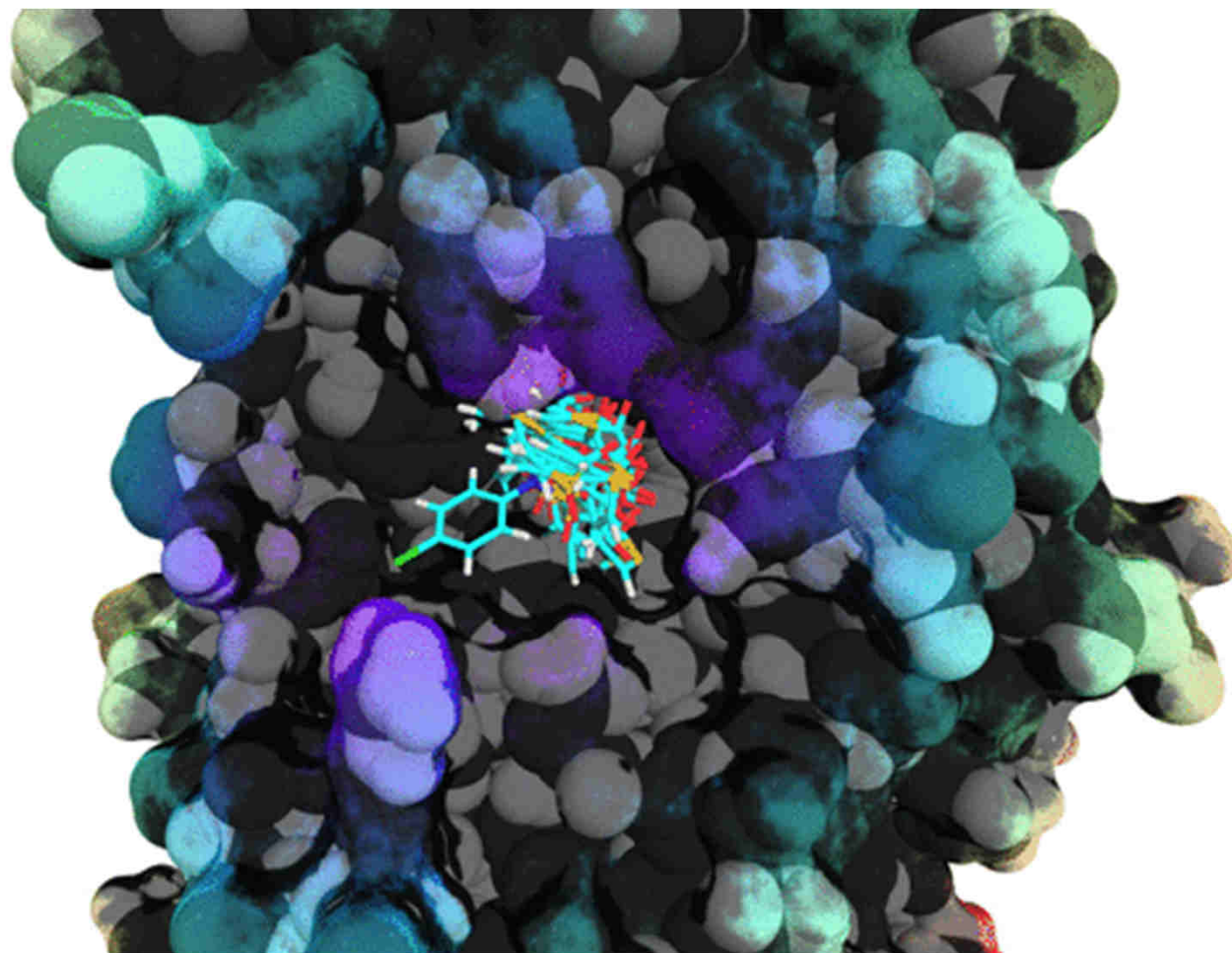
How it works in DOCK3.7—the movie



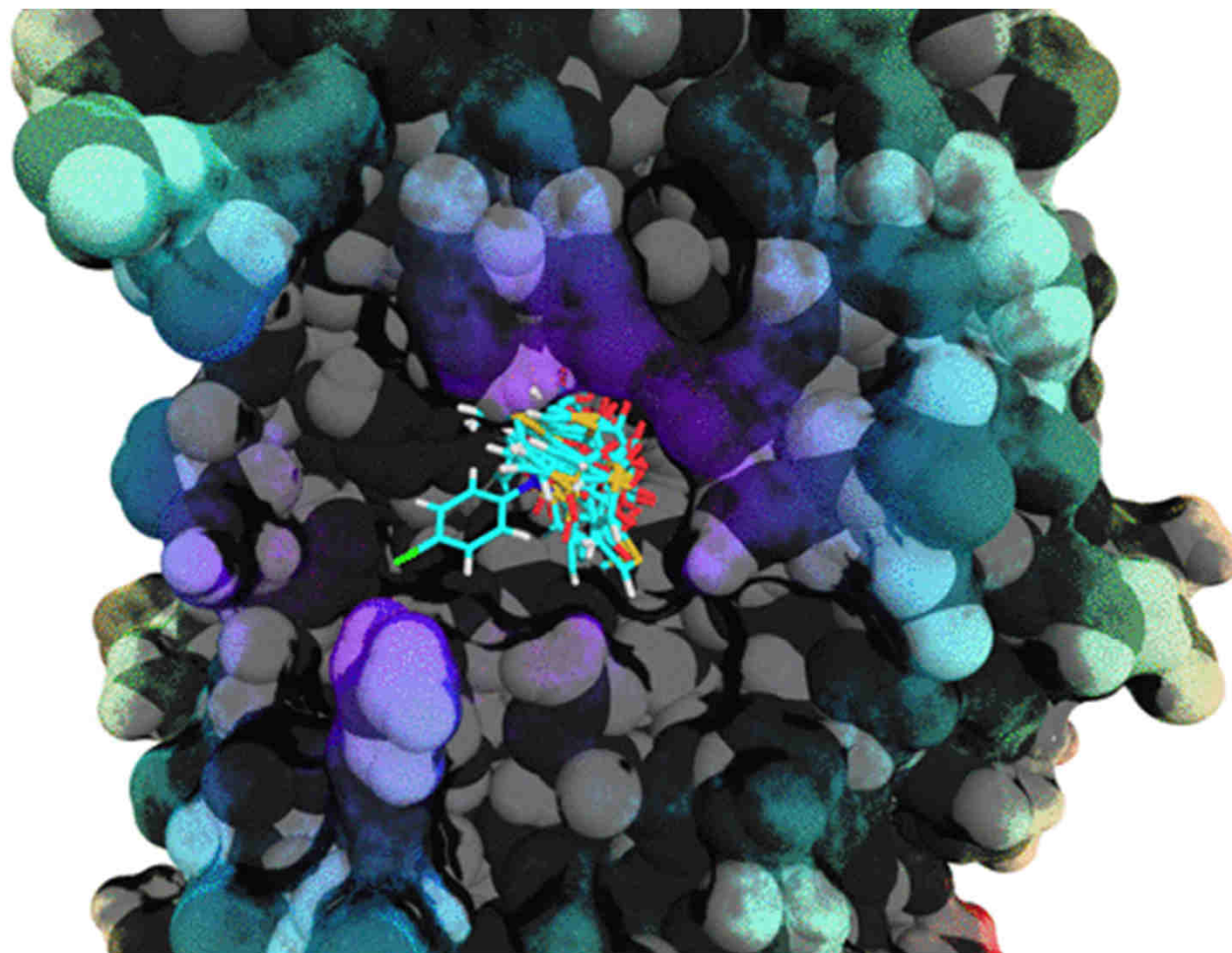
How it works in DOCK3.7—the movie



How it works in DOCK3.7—the movie

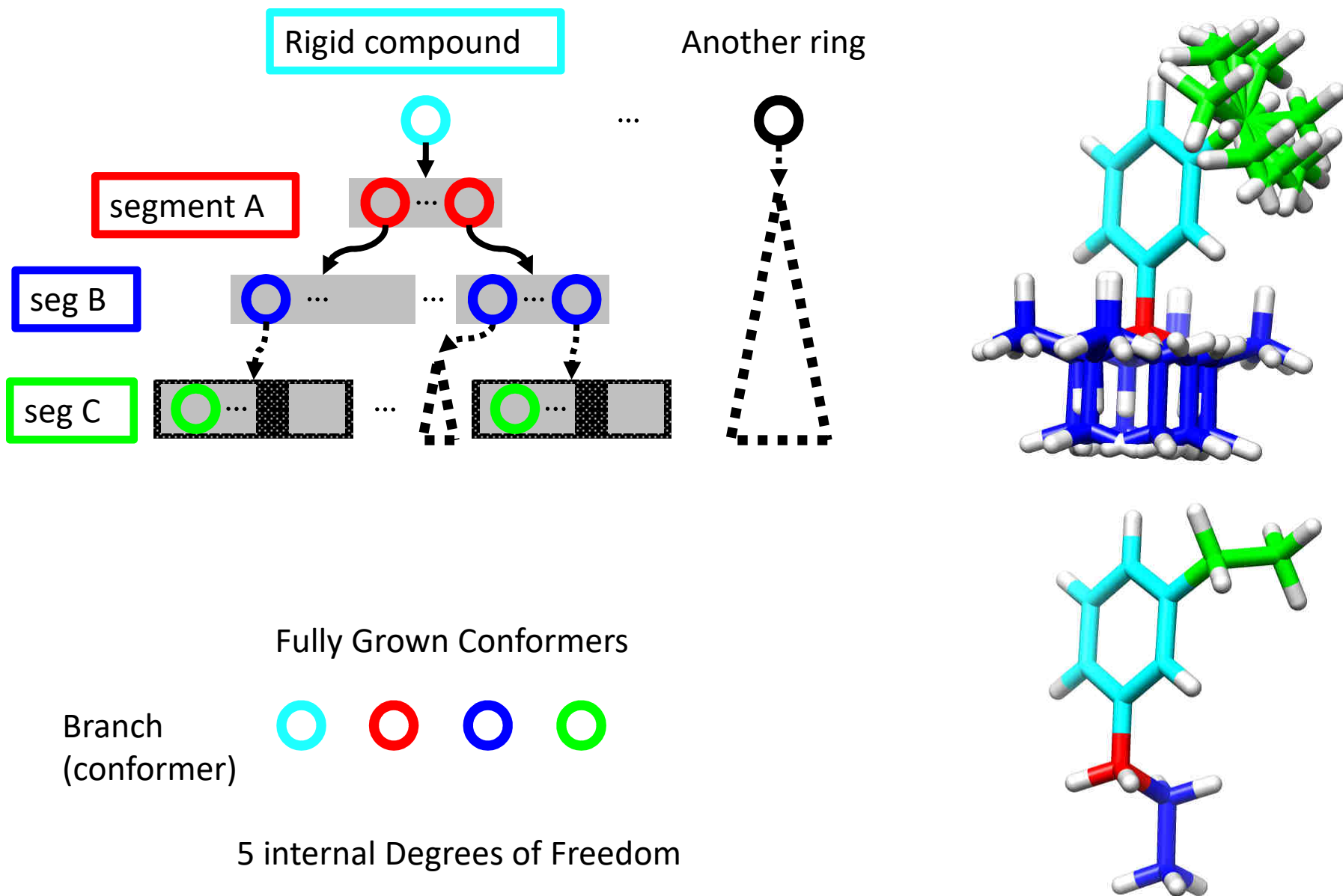


How it works in DOCK3.7—the movie

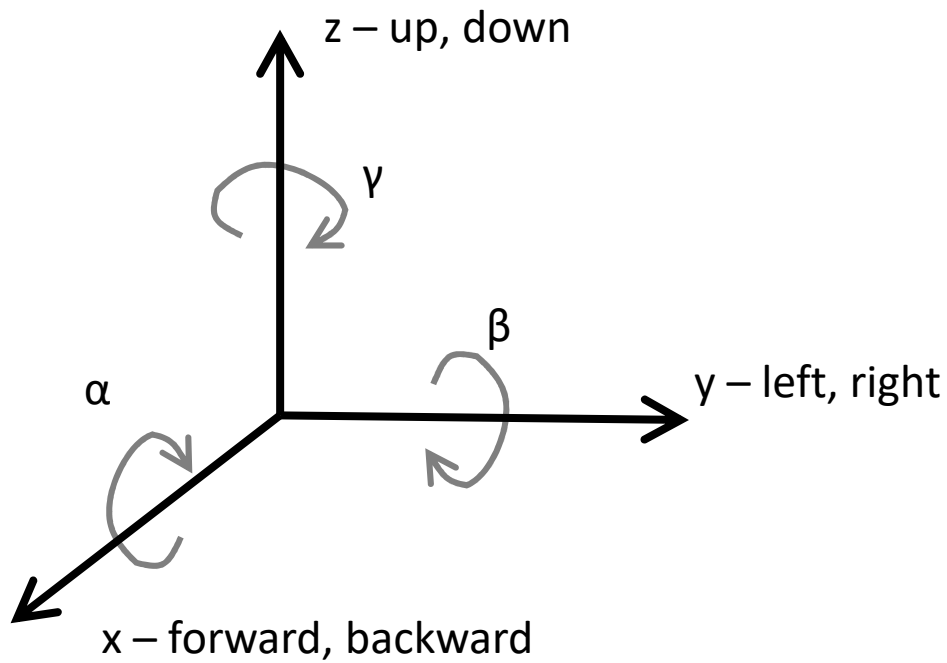


Ligand Sampling, Degrees of Freedom

Database Construction



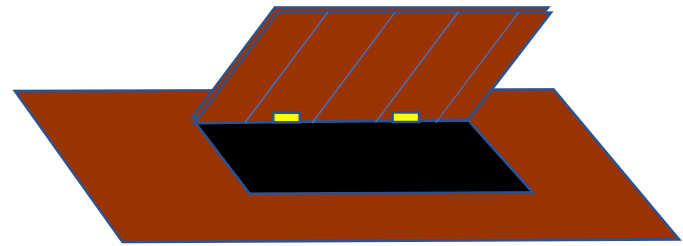
Six degrees for freedom



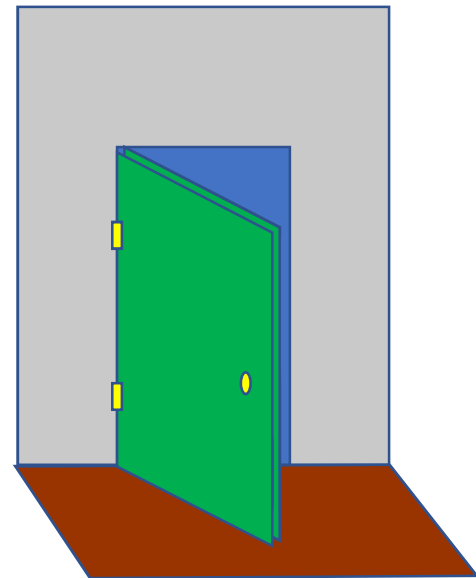
doorknob (x axis)



Trap door (hinge on y axis)

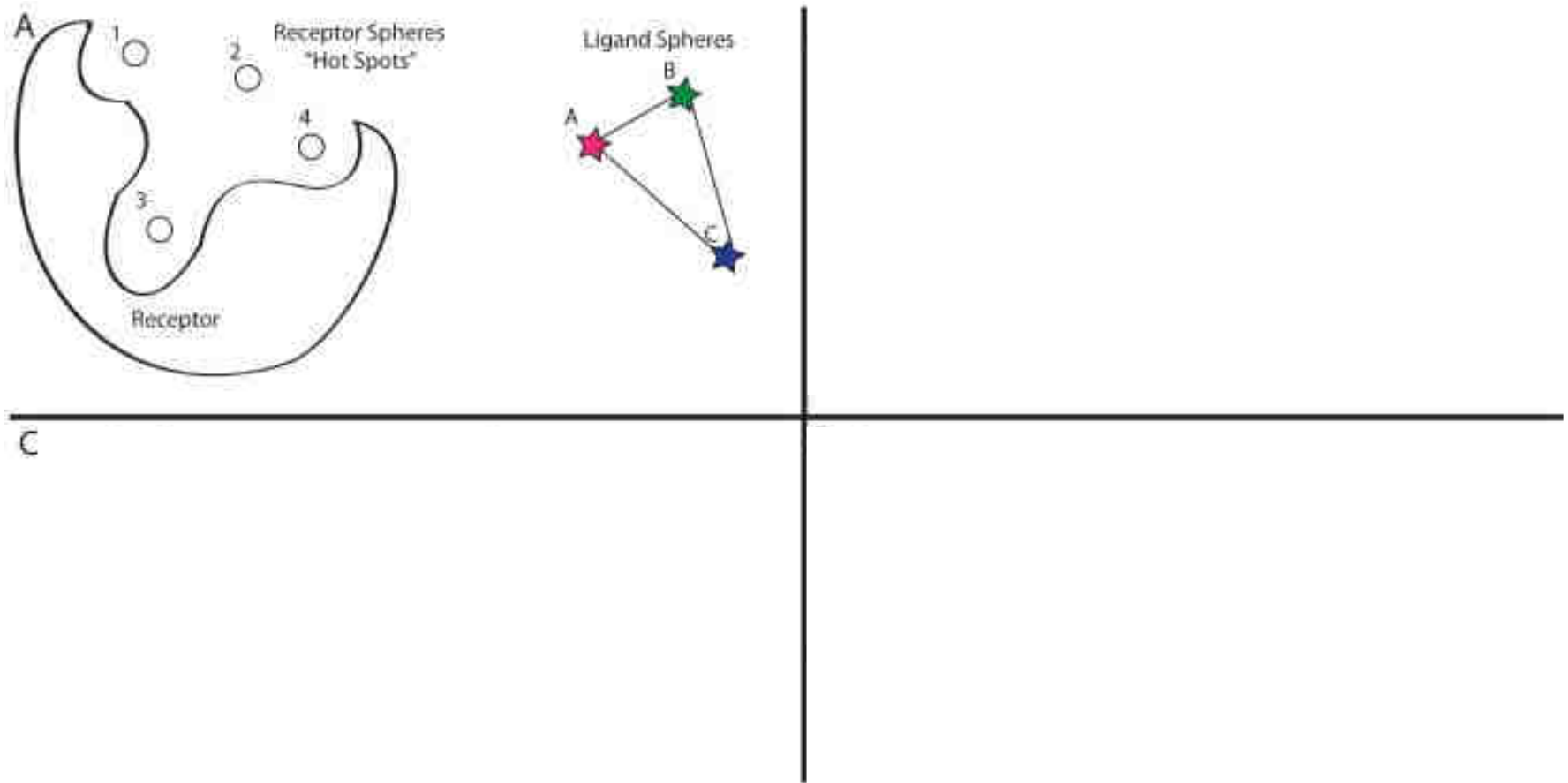


front door (hinge on z axis)



How Sampling works (orientational)

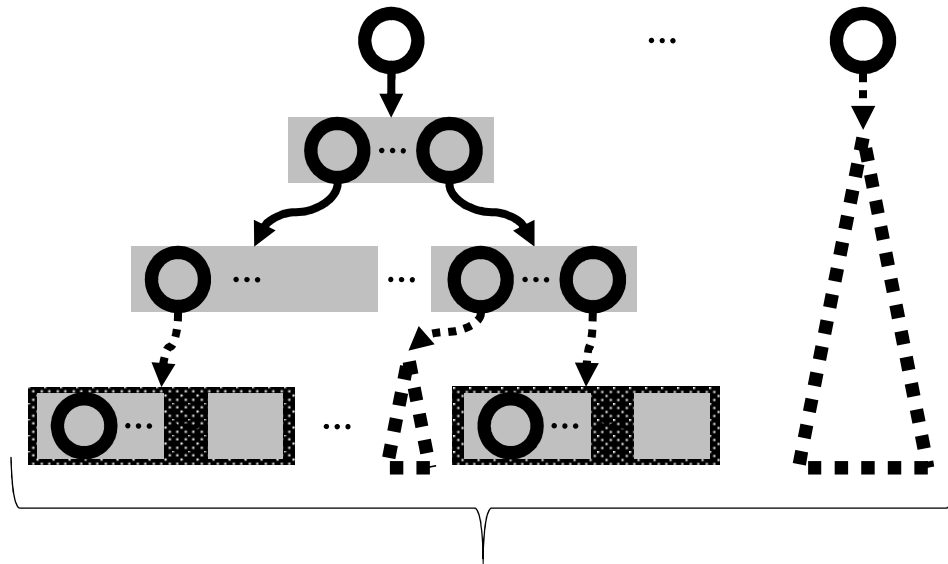
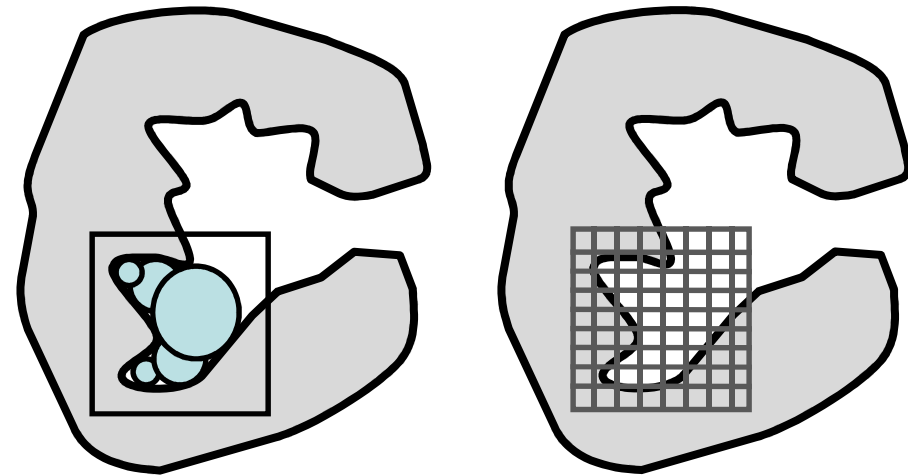
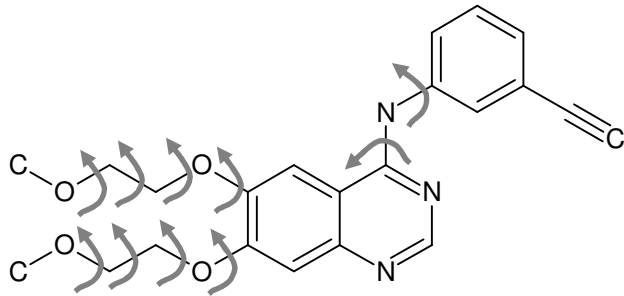
A toy example illustrating the matching sphere orientational matching algorithm



How DOCK 3.7 works

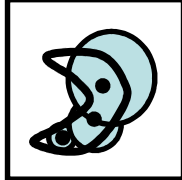
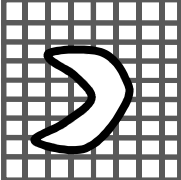
Preparation, Sampling, and Scoring

Ligand Sampling –
database construction



Dockable database file

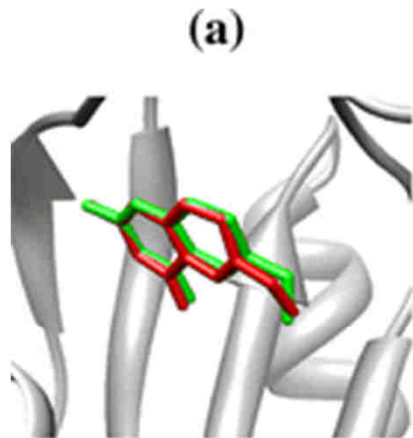
DOCKING

orient	score
	
Scoring using a grid to speed up the calculations	

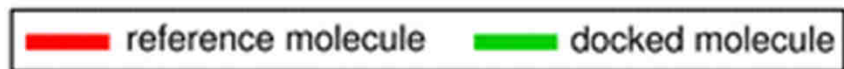
How to evaluate docking methods

- Pose reproduction, reproduce the crystallographic poses
- Enrichment calculations, make sure ligand found in the top of the rank orders lists.
- Prospective testing on model cavities, make a predication, and test it!

Pose Reproduction: RMSD Calculations



0.33 Å
success

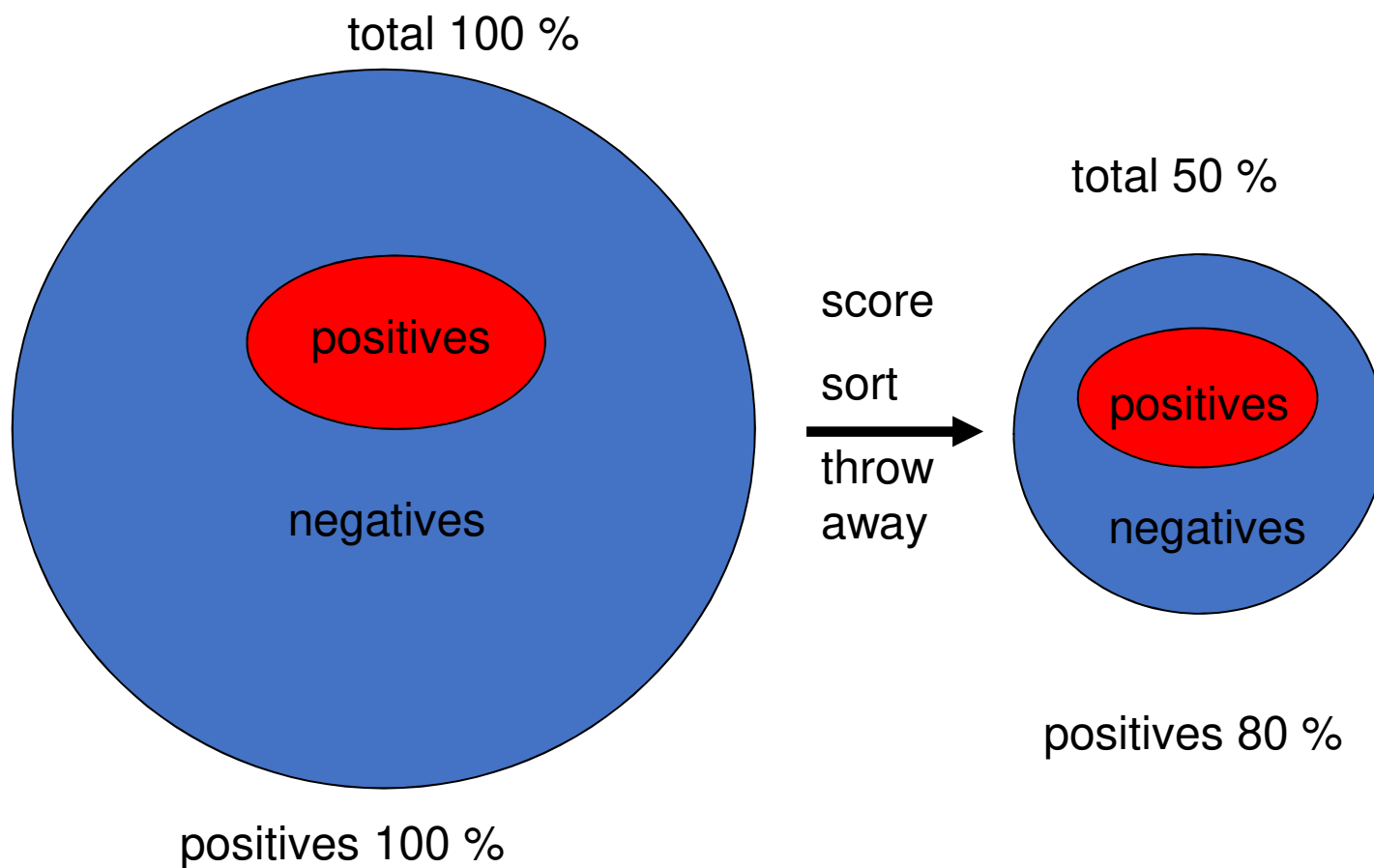


We can correct for molecular symmetry using the Hungarian algorithm

It is important that we are obtaining the correct binding mode.

Right for the right reasons

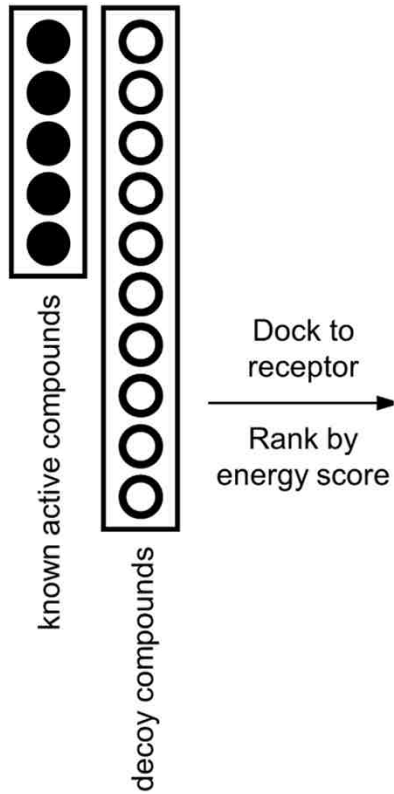
Enrichment Background



Retrospective testing

Enrich knowns over Decoys

(a)



Computational Prediction vs. Experimental Evidenced

	activity	inactivity
predicted activity	True Positive (selected actives)	False Positive (selected decoys)
predicted inactivity	False Negative (removed actives)	True Negative (removed decoys)

Computational Prediction vs. Experimental Evidenced

		activity	inactivity
Dock Score	predicted activity	True Positive (selected actives)	False Positive (selected decoys)
	predicted inactivity	False Negative (removed actives)	True Negative (removed decoys)

Computational Prediction vs. Experimental Evidenced

wet lab experiment
activity inactivity

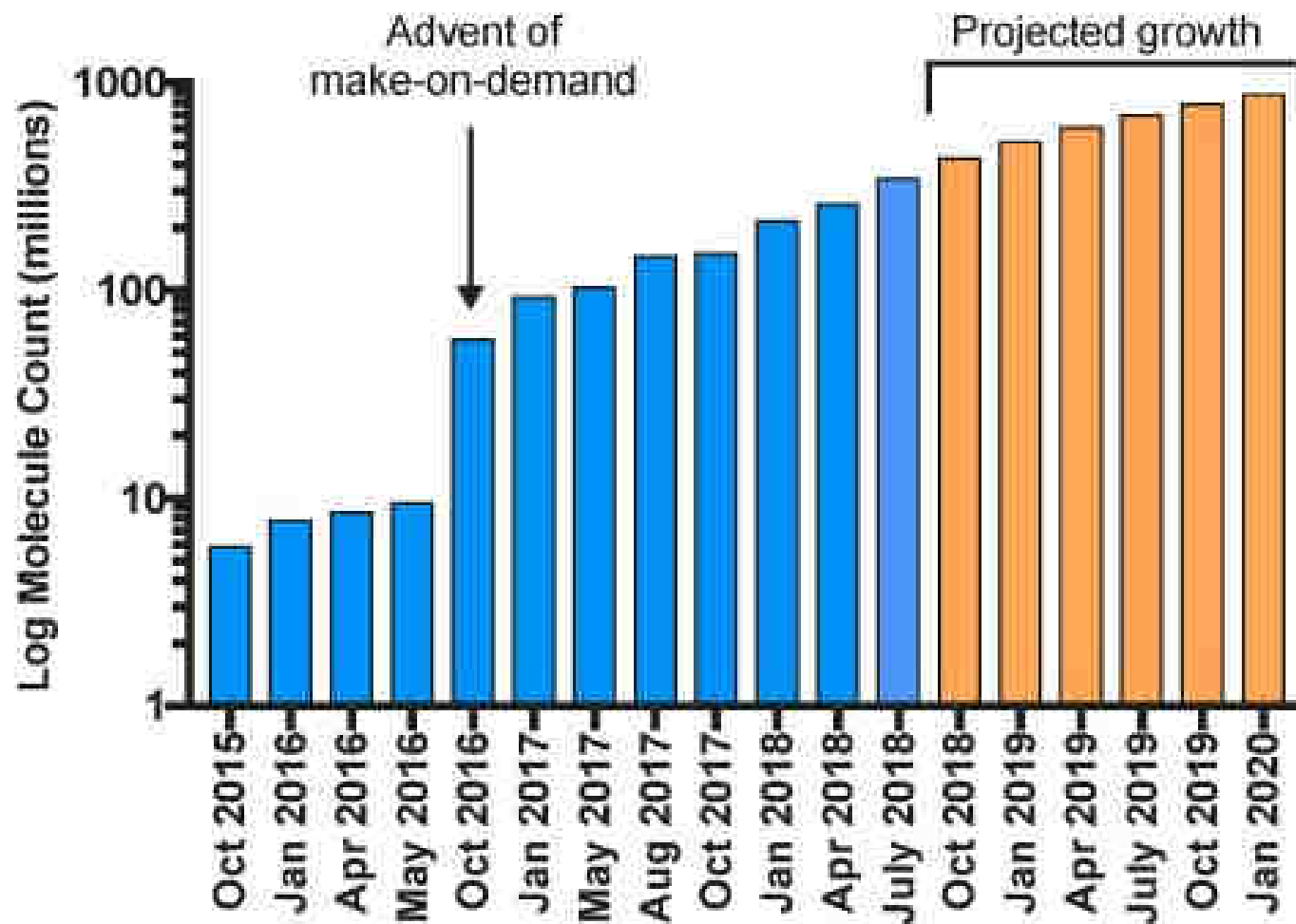
Dock Score	predicted activity	True Positive (selected actives)	False Positive (selected decoys)
	predicted inactivity	False Negative (removed actives)	True Negative (removed decoys)

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Large-scale docking:
Screen all of purchasable space

Irwin's law: Docking libraries, crucial for chemical discovery, doubling every 2.5 years

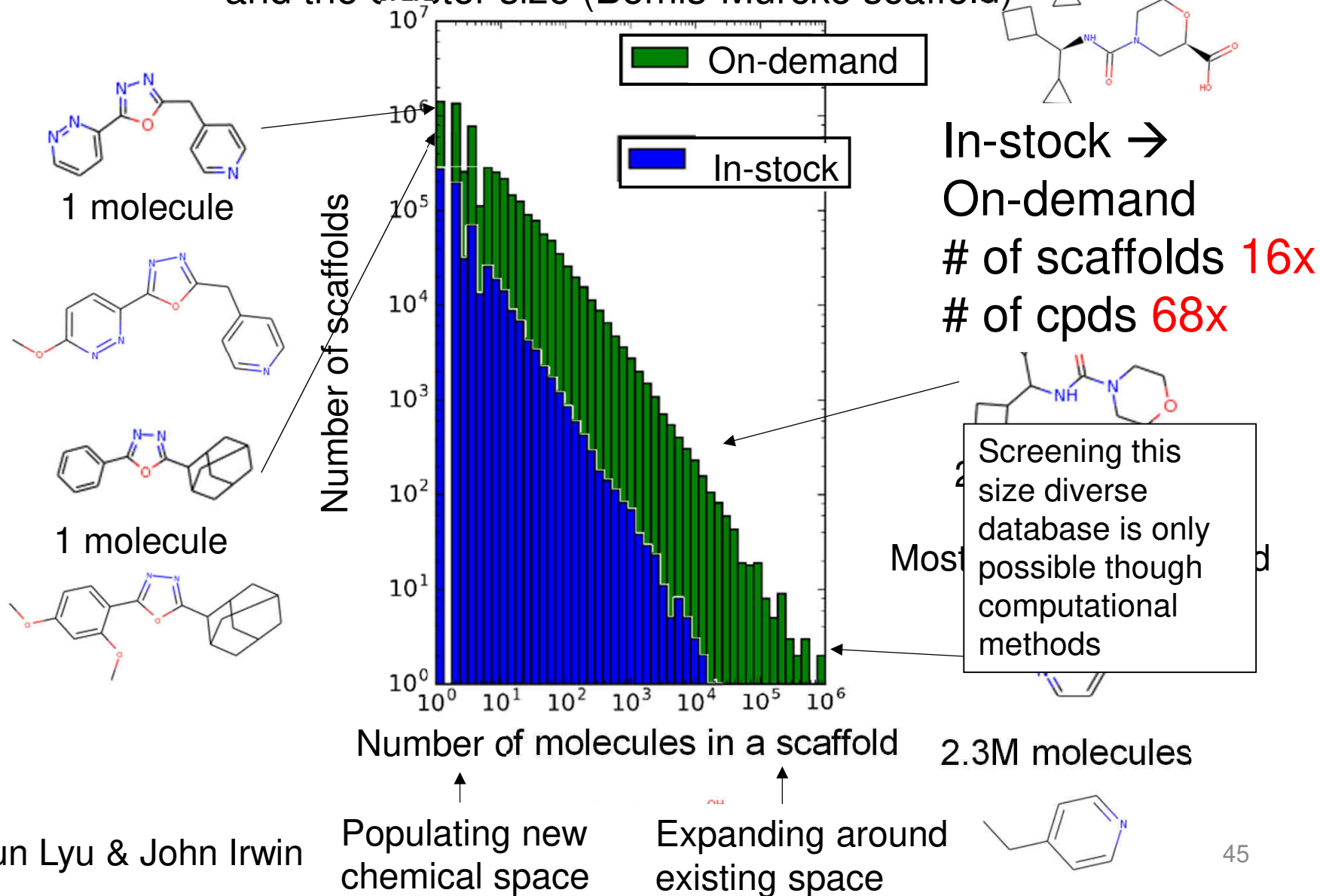


250 Million (now) -> 1B (next year) -> 10 Billion (next few years)!

How can we keep up in docking?

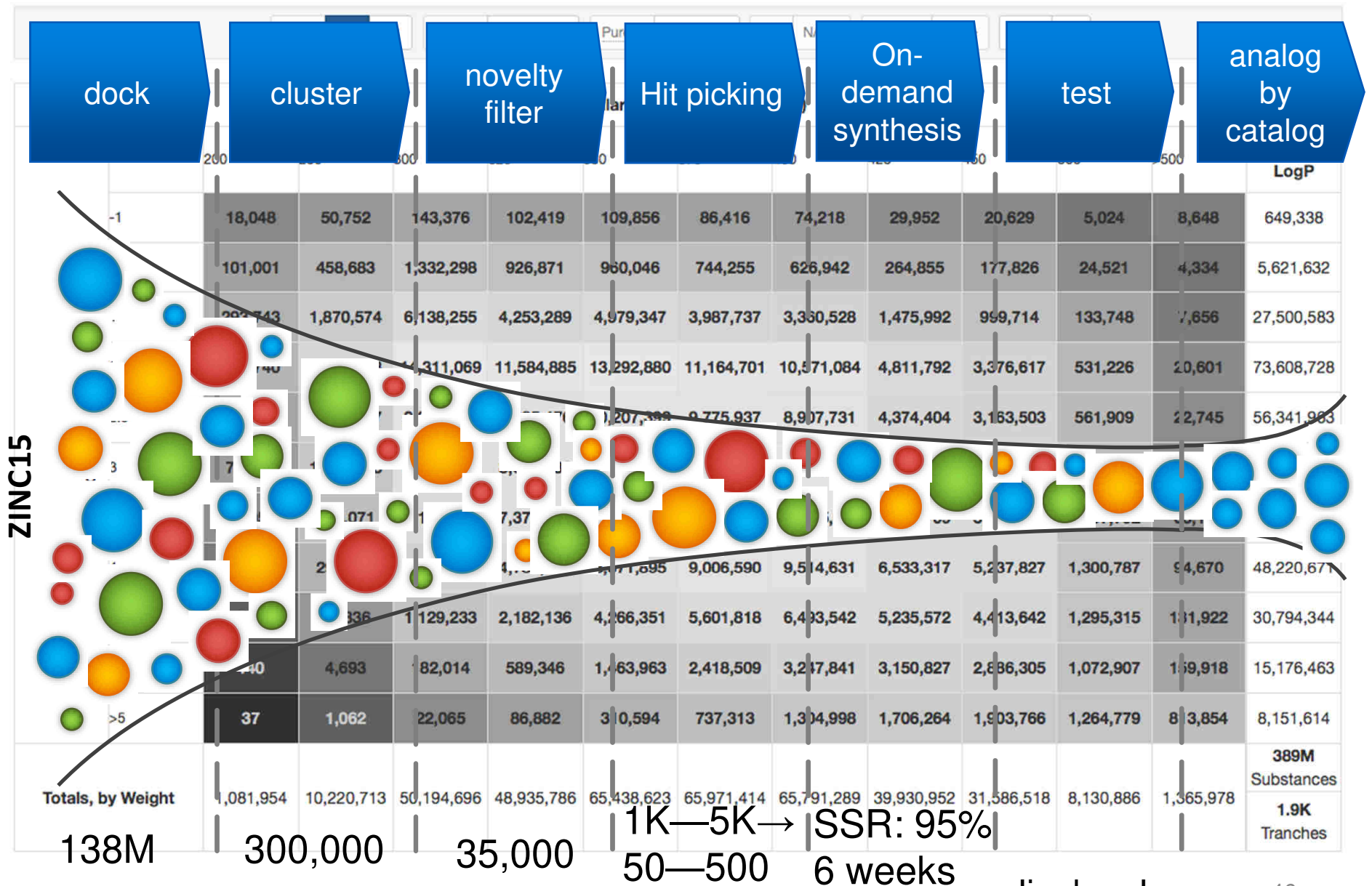
Database is larger and more diverse

The relationship between number of clusters and the cluster size (Bemis-Murcko scaffold)



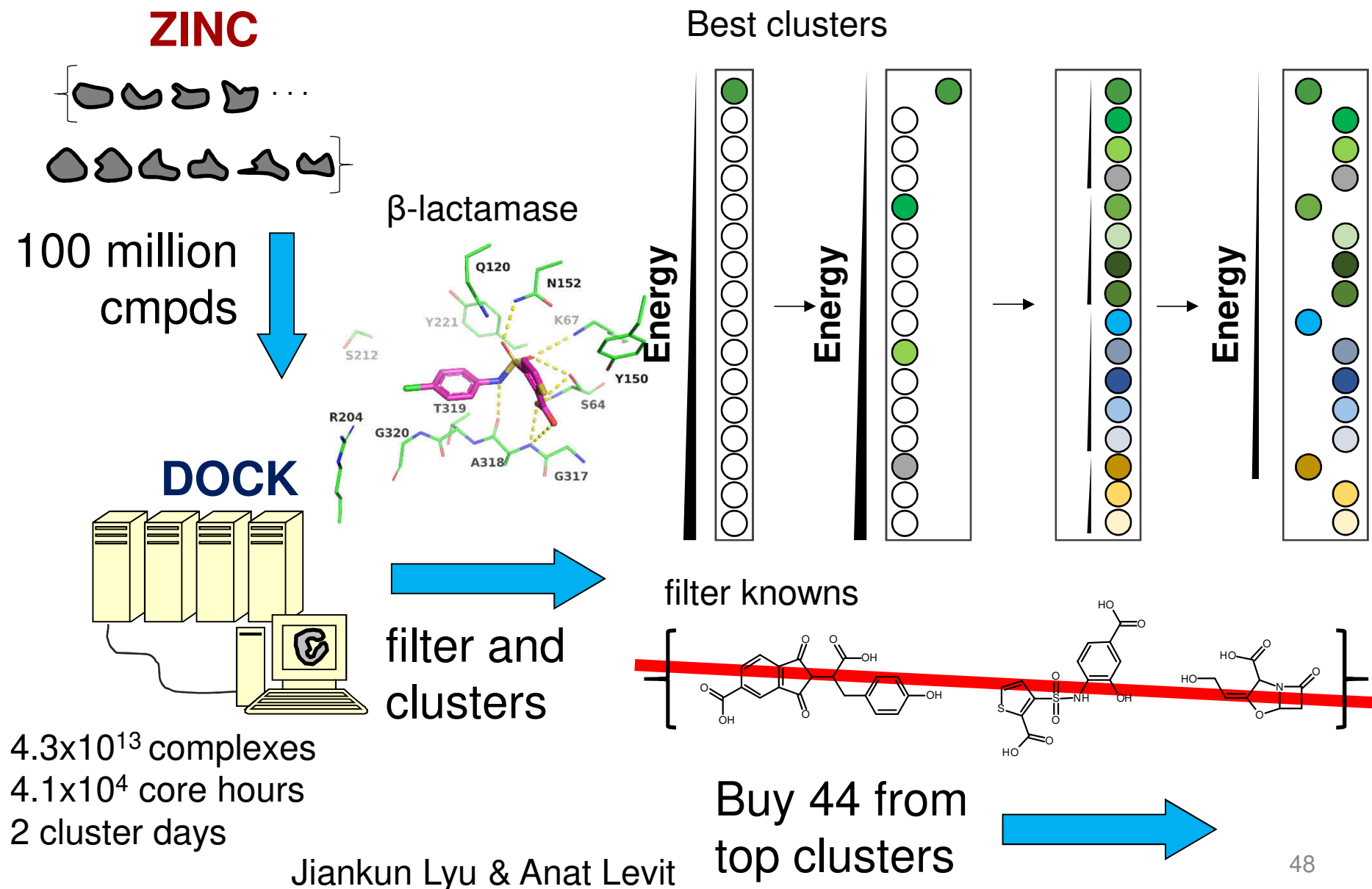
In-stock →
On-demand
of scaffolds **16x**
of cpds **68x**

Large scale docking flow

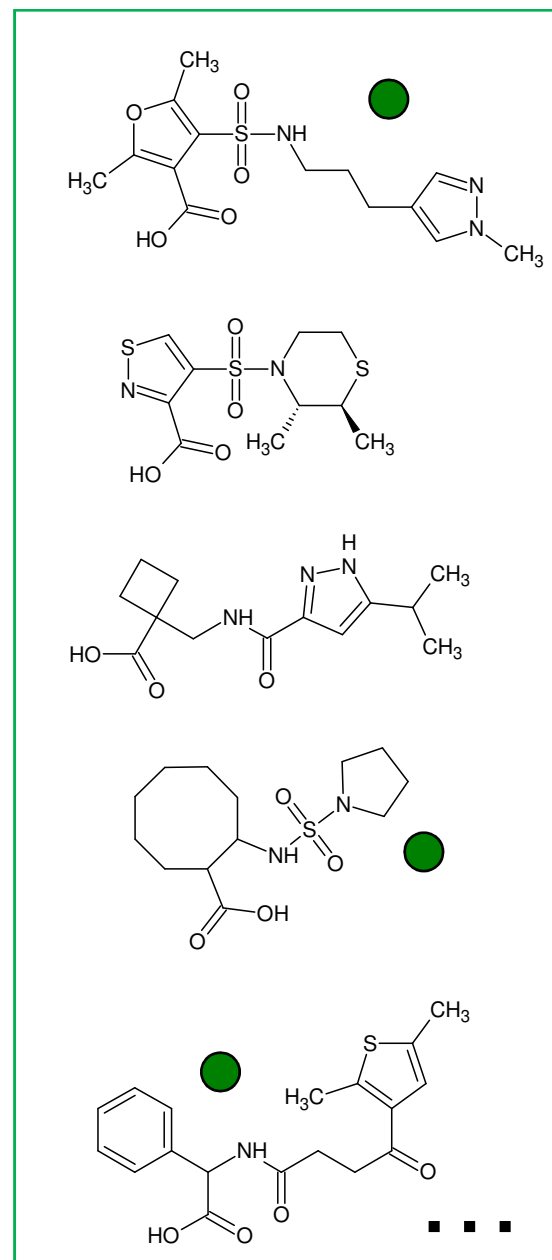
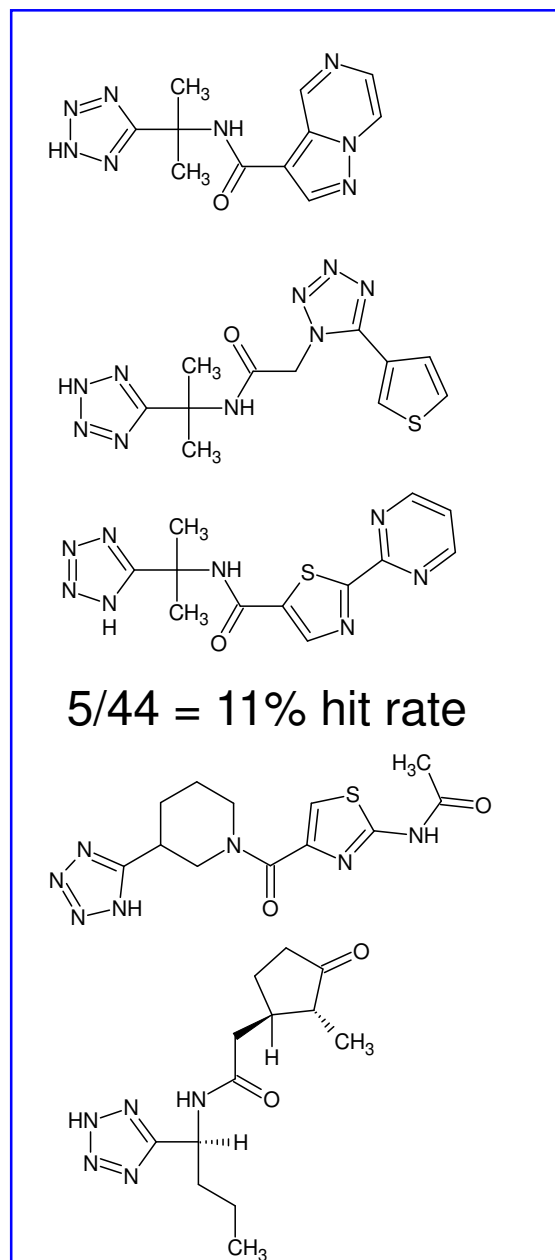
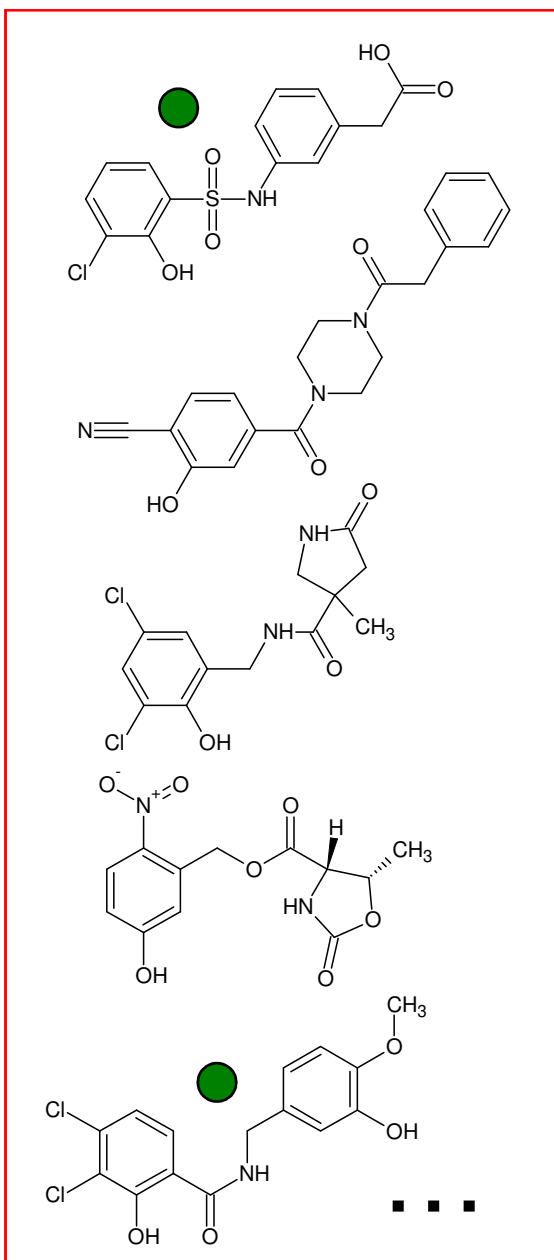


AmpC β -lactamase

We docked 100M molecules to **AmpC** β -lactamase

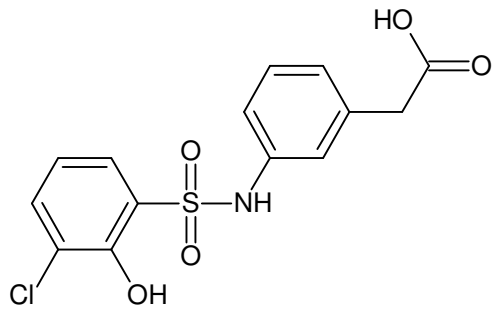


We bought new chemotypes

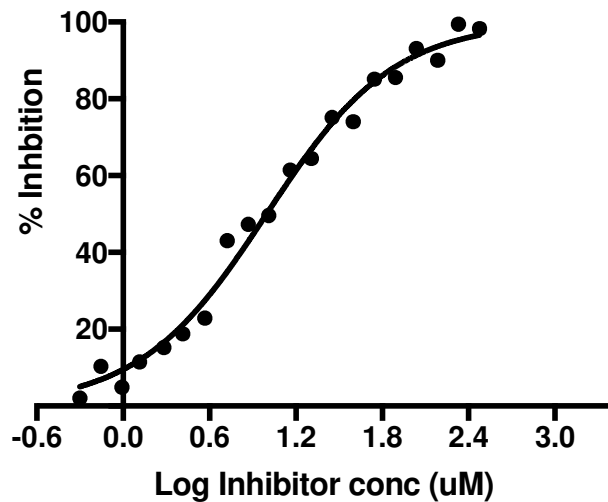
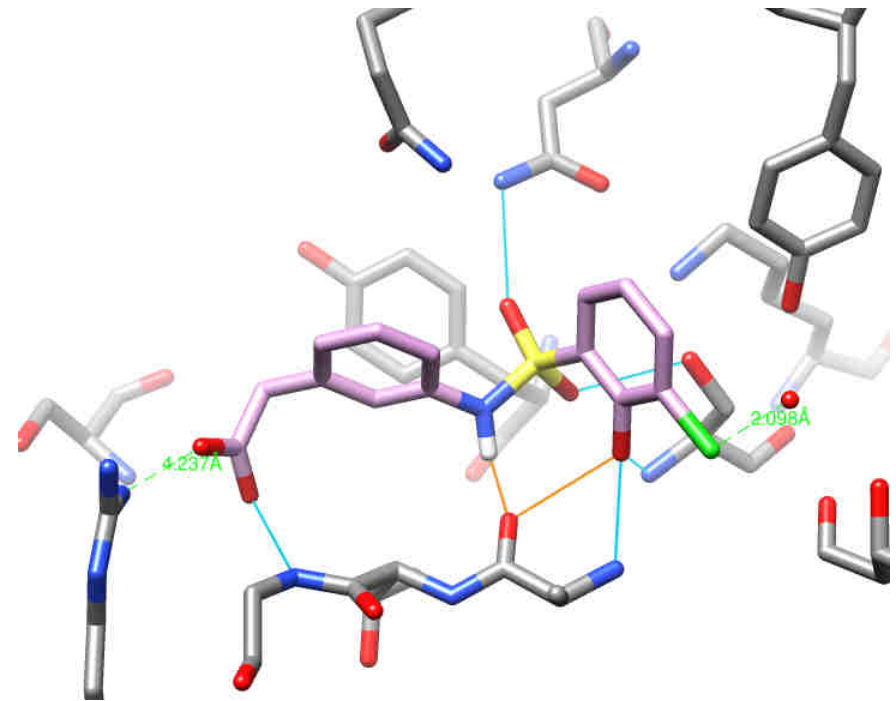


Best AmpC inhibitor found right out of docking (1 μM)

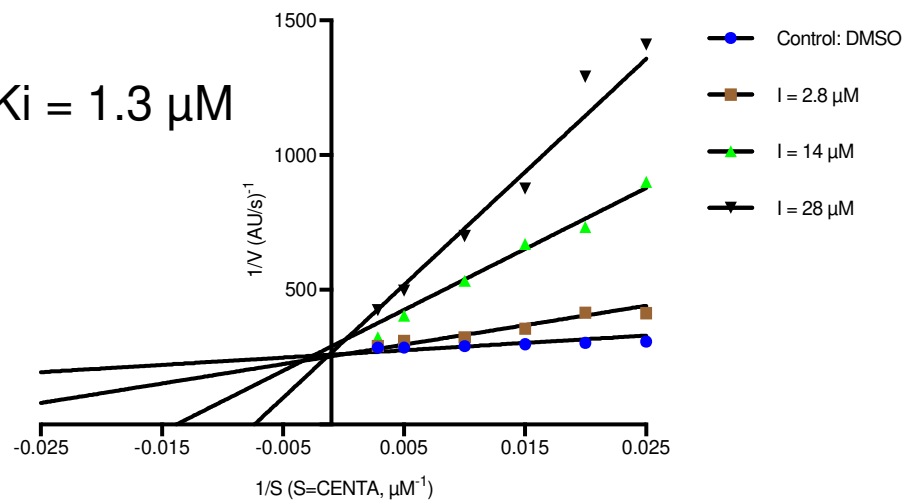
Compound 10 ZINC000339204163



Best AmpC inhibitor found straight out of docking or any screening method

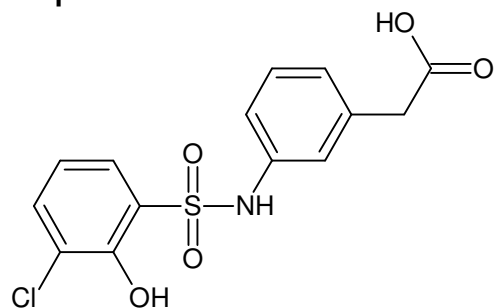


$K_i = 1.3 \mu\text{M}$



Crystal structure confirms docked pose and phenolate

Cmpd 10

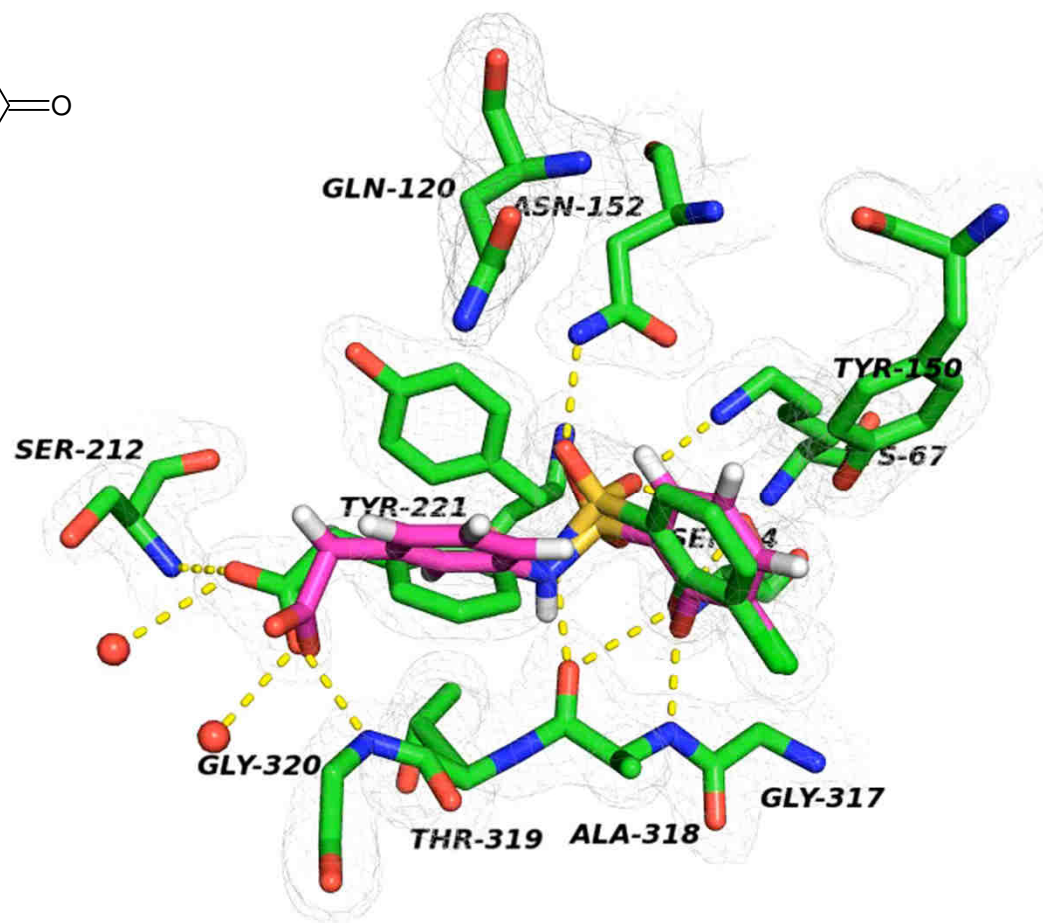


K_i=1.3 μM

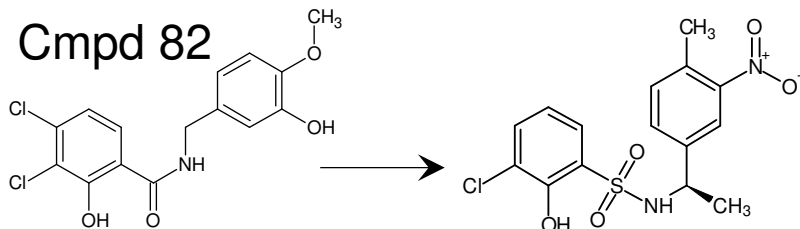
rmsd=0.98Å

Resolution=1.9 Å

pK_a of phenol is 5.8
Determined on a
carboxyl-less analog

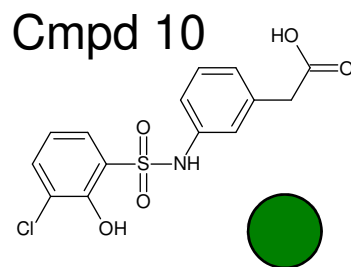


5 hits (out of 45 molecules test) and their analogs

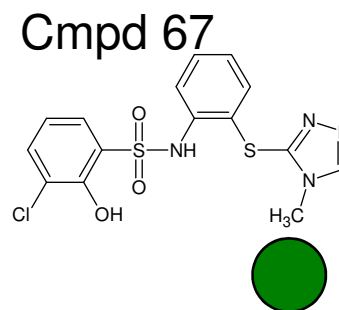


Ki = 210 μ M

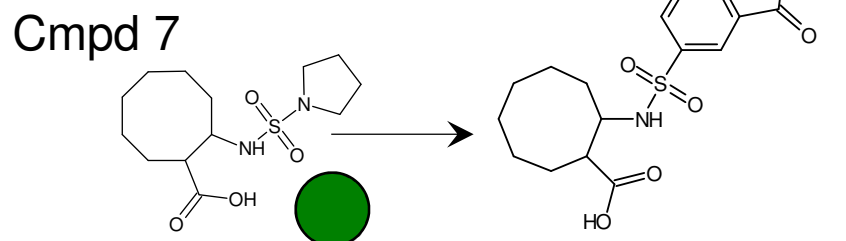
Ki = 100 μ M



Ki = 1.3 μ M

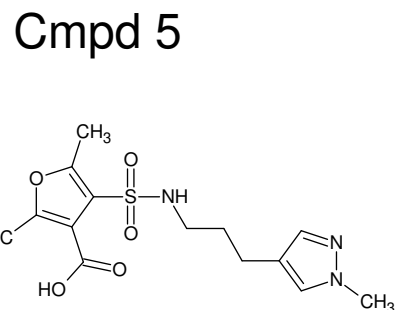


Ki = 77 nM

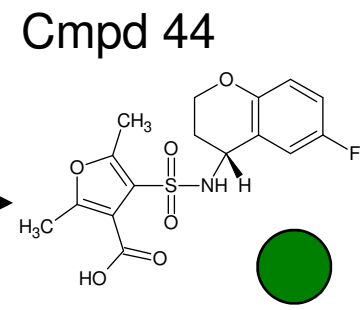


Ki = 260 μ M

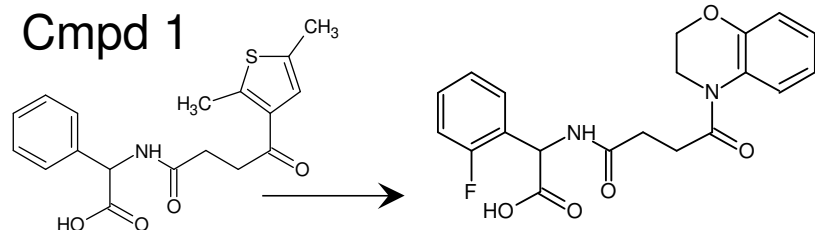
Ki = 80 μ M



Ki = 190 μ M



Ki = 80 μ M



Ki = 400 μ M

Ki = 14 μ M

- 9/31 Cmpd1 Analog
- 8/15 Cmpd 5 Analogs
- 9/19 Cmpd 7 Analog
- 8/11 Cmpd 10 Analogs
- 8/14 Cmpd 82 Analogs

- Determined 4 crystal structures

Isha Singh

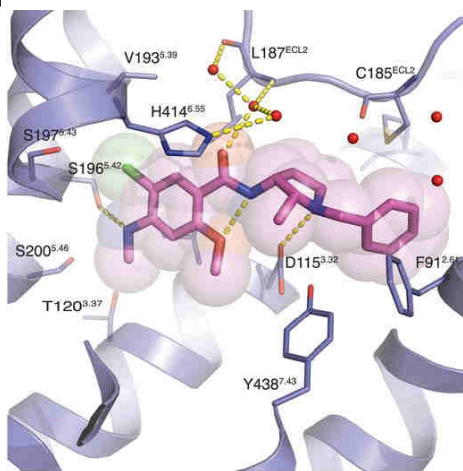
D4 Dopamine receptor

How many active ligands are in the library for **DRD4**?

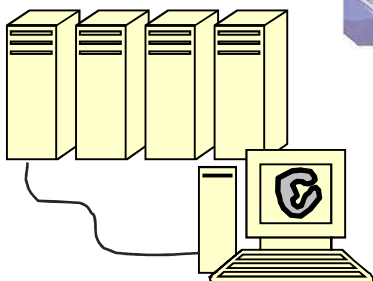
ZINC



138 million cmpds



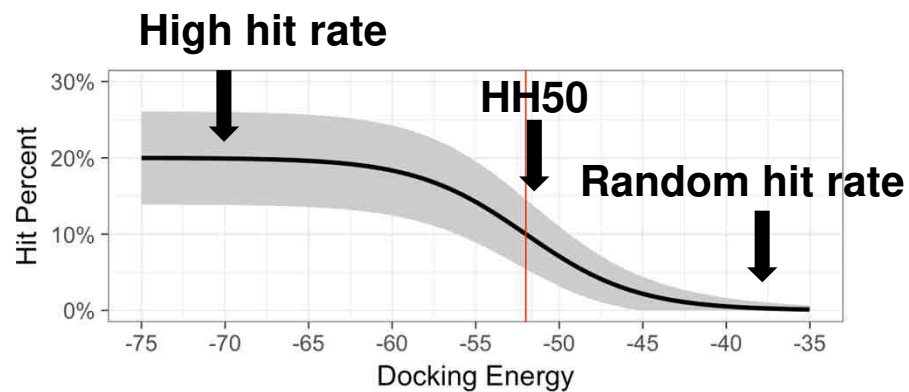
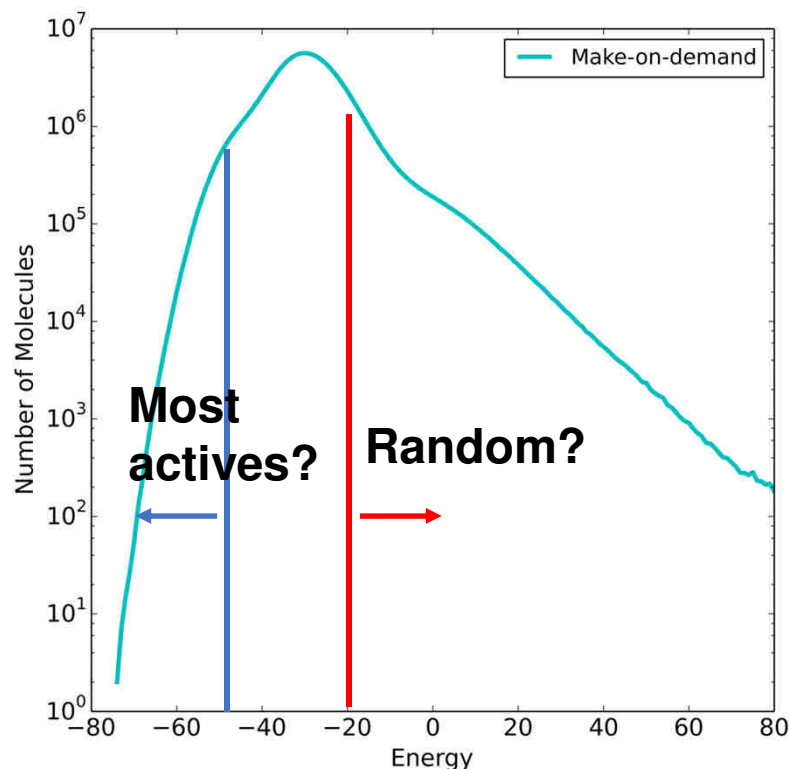
DOCK



~~Test 50~~

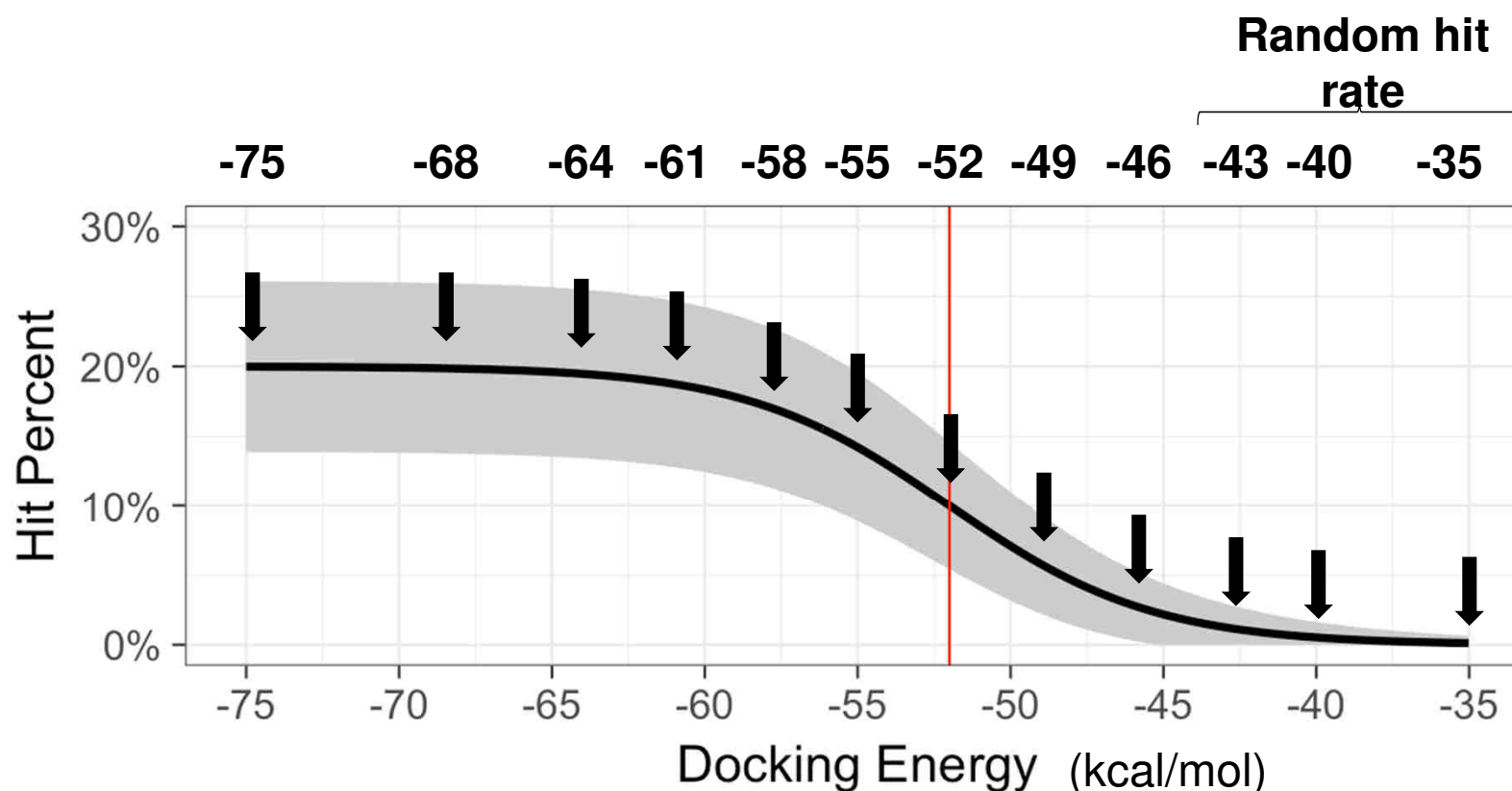
Test 549

6.92 x 10¹³ complexes
4.4x10⁴ core hours
1.8 cluster days



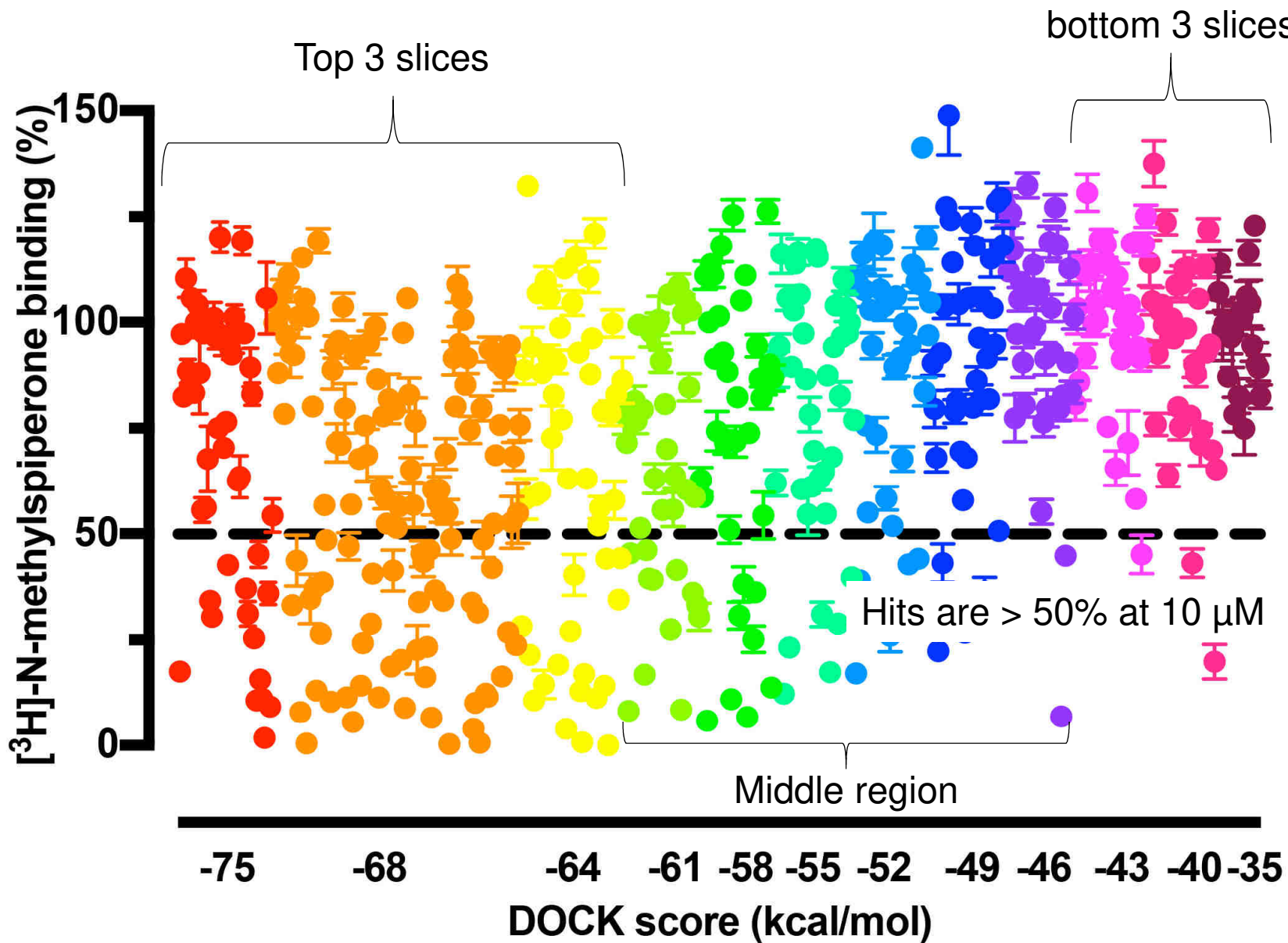
Jiankun Lyu, Matthew O'Meara

Bought 444 molecules to estimate the DOCKing hit rate curve for DRD4

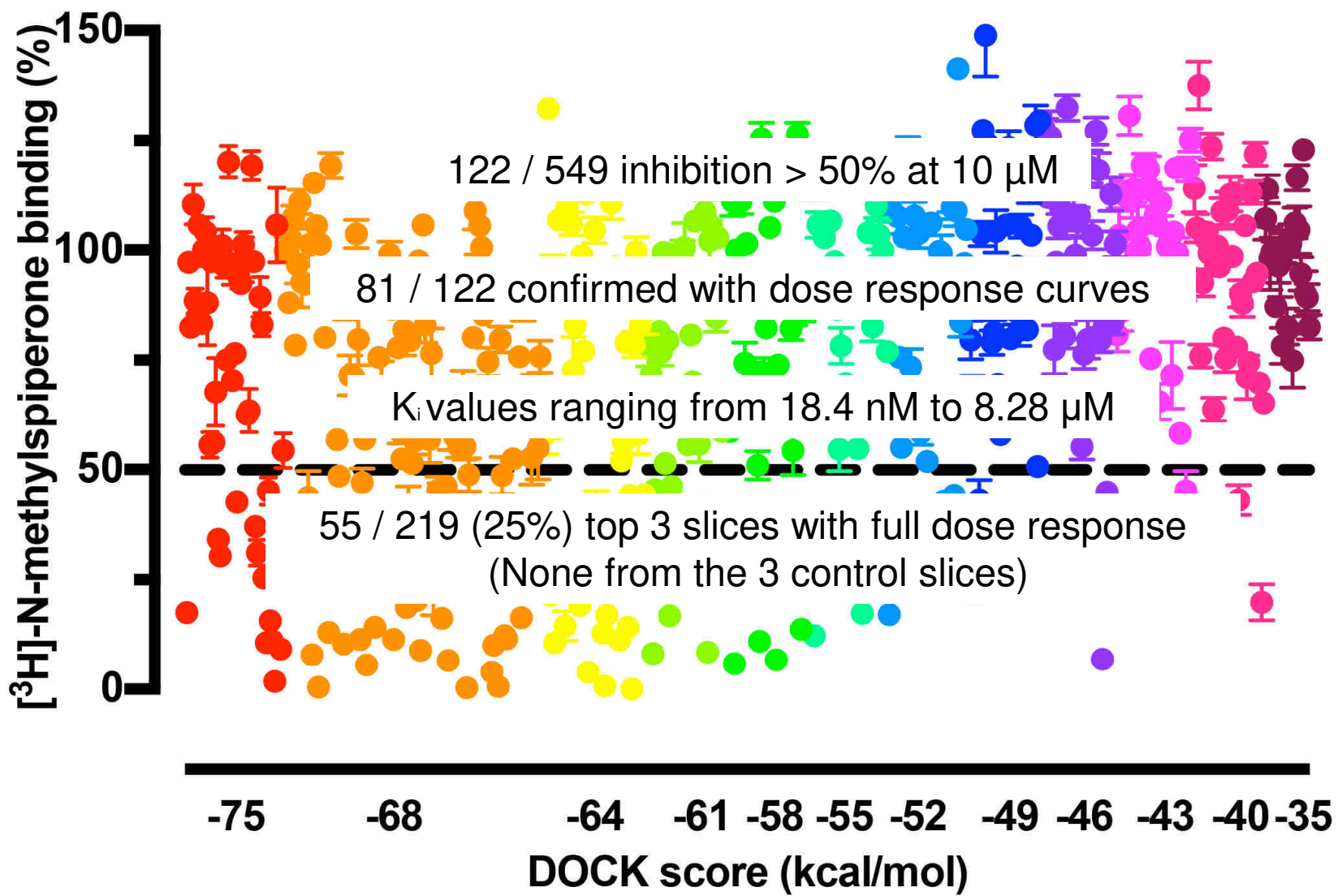


444 molecules were picked automatically, with 35 to 40 molecules sampled at 12 energy windows from docking scores from -75 to -35 kcal/mol.

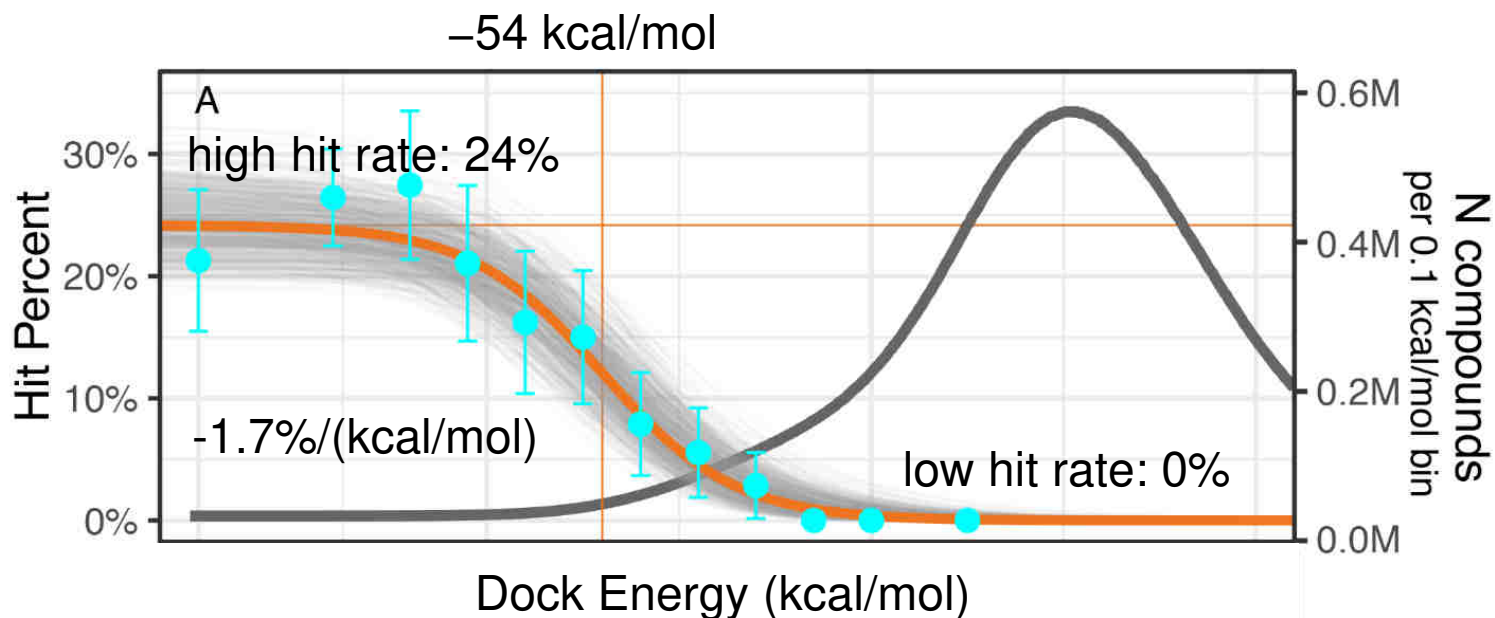
Great hit rate of 25% at top, poor hit rate of 0% on right



Great hit rate of 25% at top, poor hit rate of 0% on right

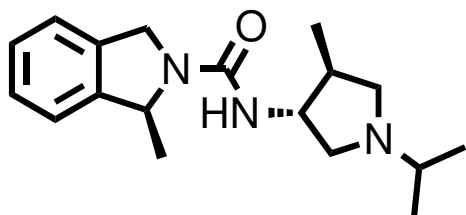


Among the 138 million molecule library there are calculated to be over 481,000 D4 active molecules

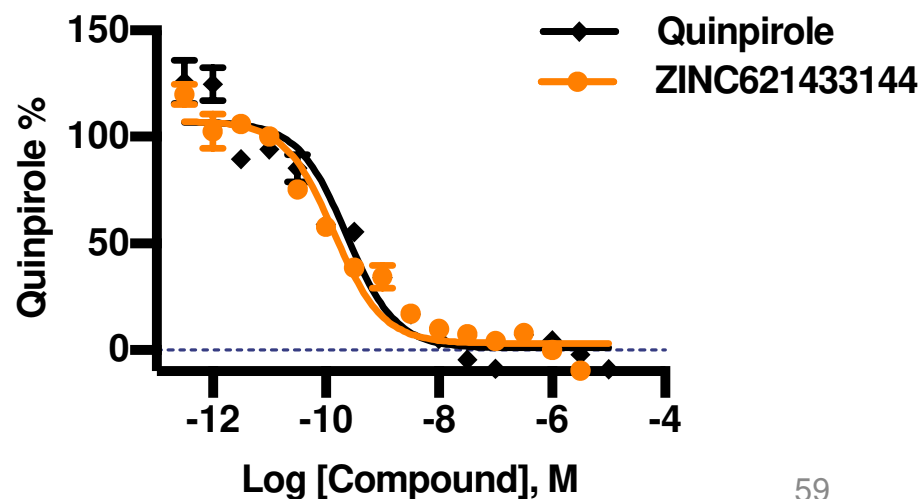
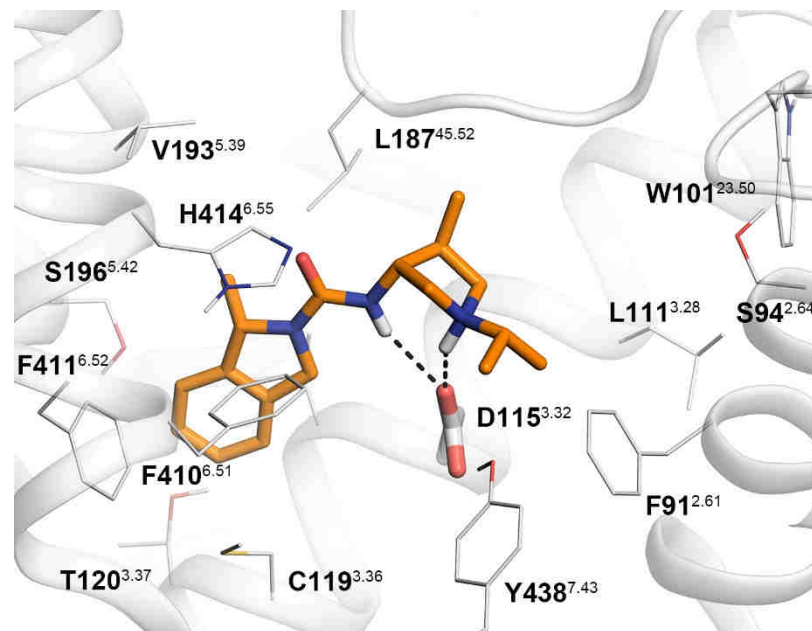


hit-rates fell almost monotonically with score

180 pM Gi-biased, selective, full agonist, among the most potent sub-type selective agonists known for this receptor



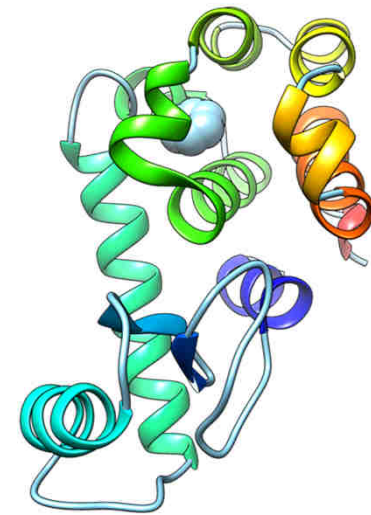
ZINC621433144
 $K_{i,DRD4} = 4.32 \text{ nM}$
 $K_{i,DRD2} > 10,000 \text{ nM}$
 $K_{i,DRD3} > 10,000 \text{ nM}$
cAMP $EC_{50} = 0.18 \text{ nM}$
Tango $EC_{50} = 57.3 \text{ nM}$
 G_i BRET $EC_{50} = 0.56 \text{ nM}$
Arrestin BRET $EC_{50} = 2.3 \text{ nM}$
Bias factor = 17 to G protein



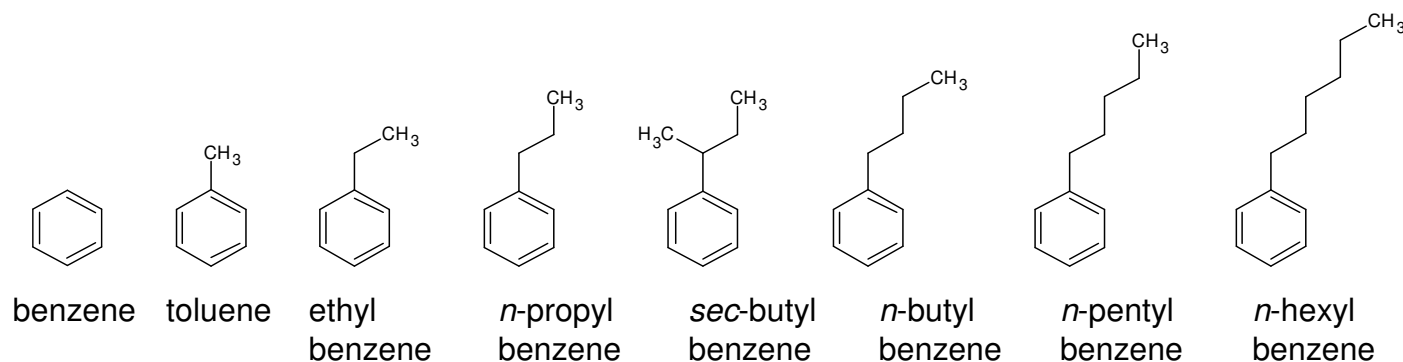
Outline

- The binding event using a thermodynamic cycle
- Introduction to molecular mechanics
- Introduction to molecular dynamics
- Introduction to molecular docking
- **Applications from my work**
 - Large-scale docking
 - **Receptor flexibility**
 - Receptor desolvation

T4 lysozyme L99A

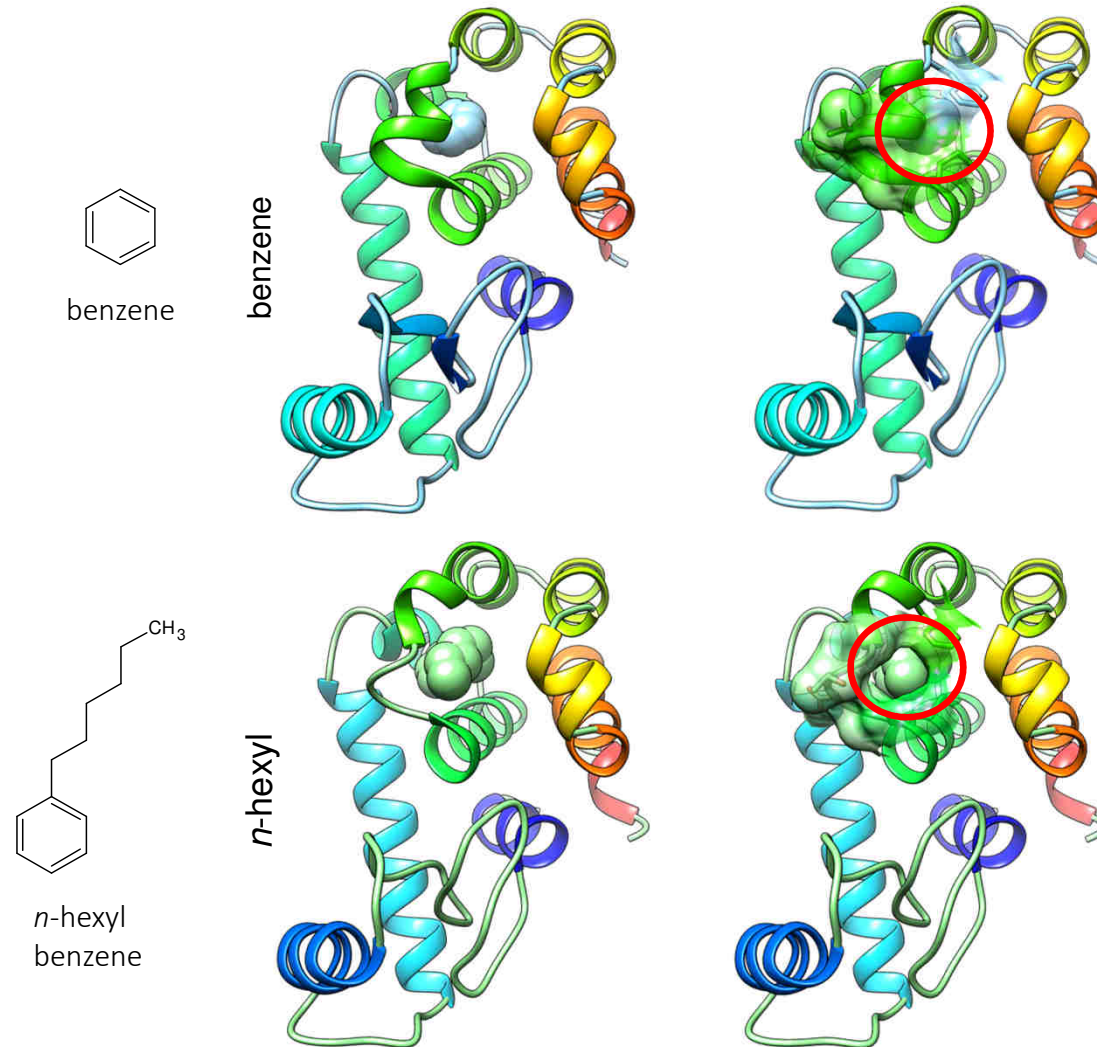


How does T4-lysozyme bind a congeneric ligand series

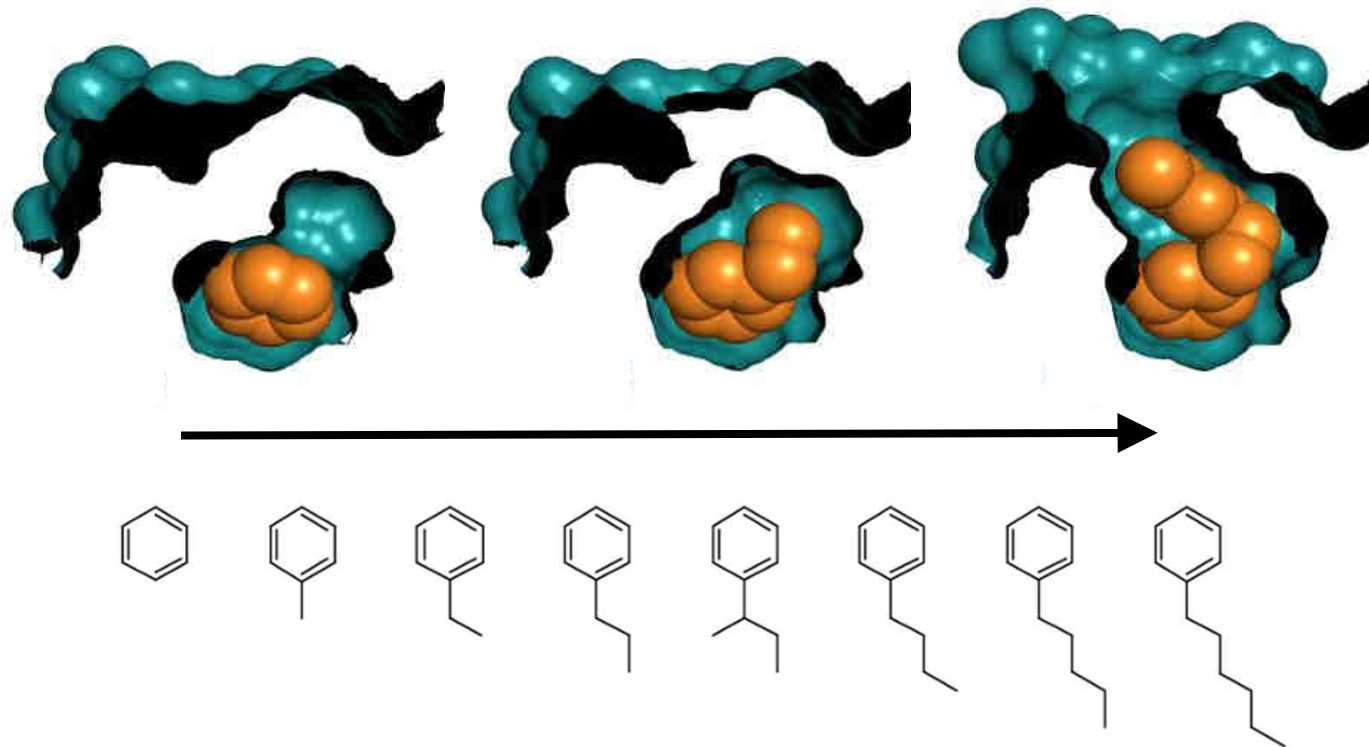


Merski, Fischer, Balias et al 2015 PNAS 112(16):5039-44
Morton, et al. 1995 Biochemistry 34(27):8564–8575
Morton & Matthews 1995 Biochemistry 34(27):8576–8588

T4 lysozyme L99A opens



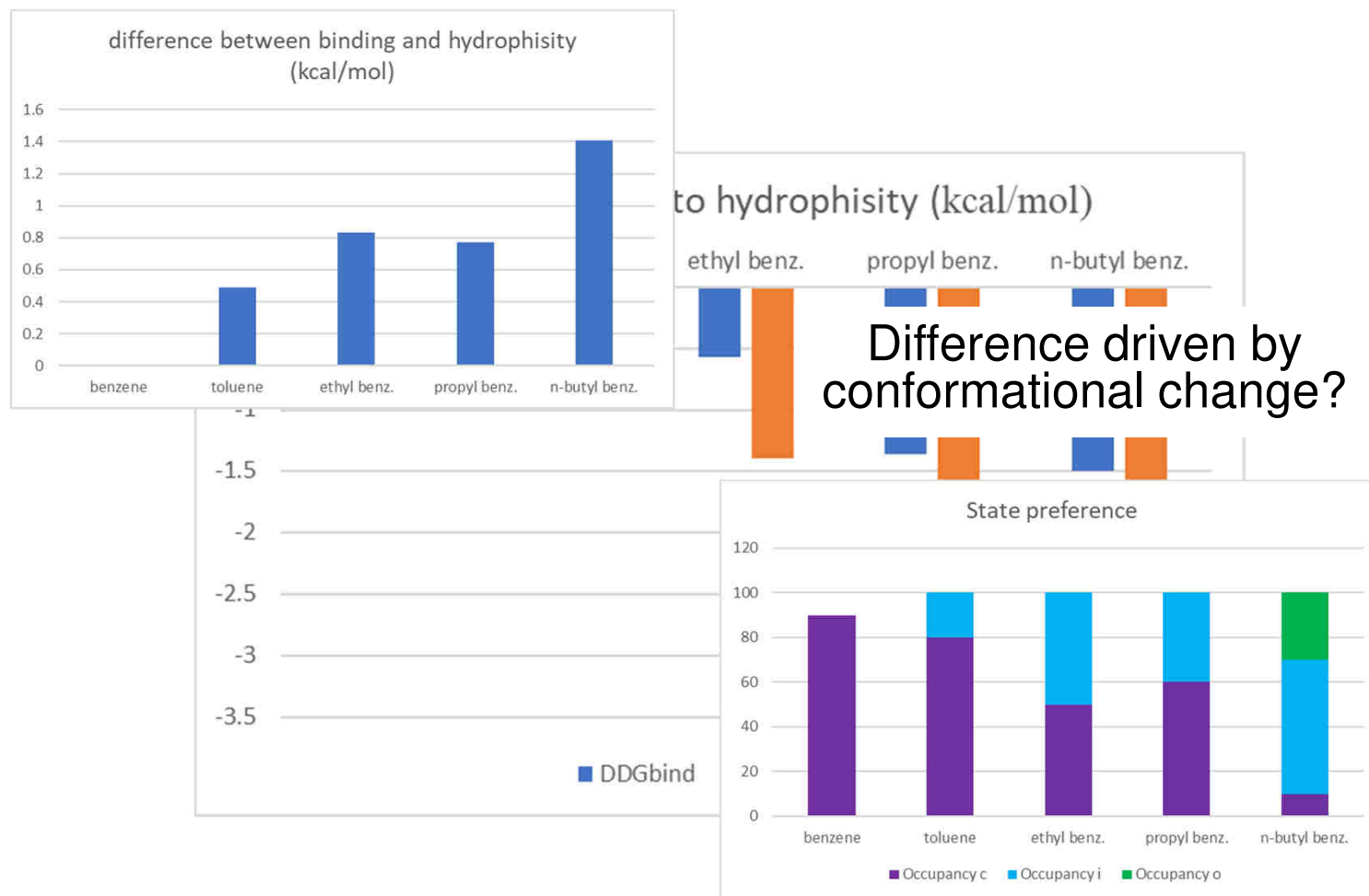
Probing receptor conformational change induce by ligand series



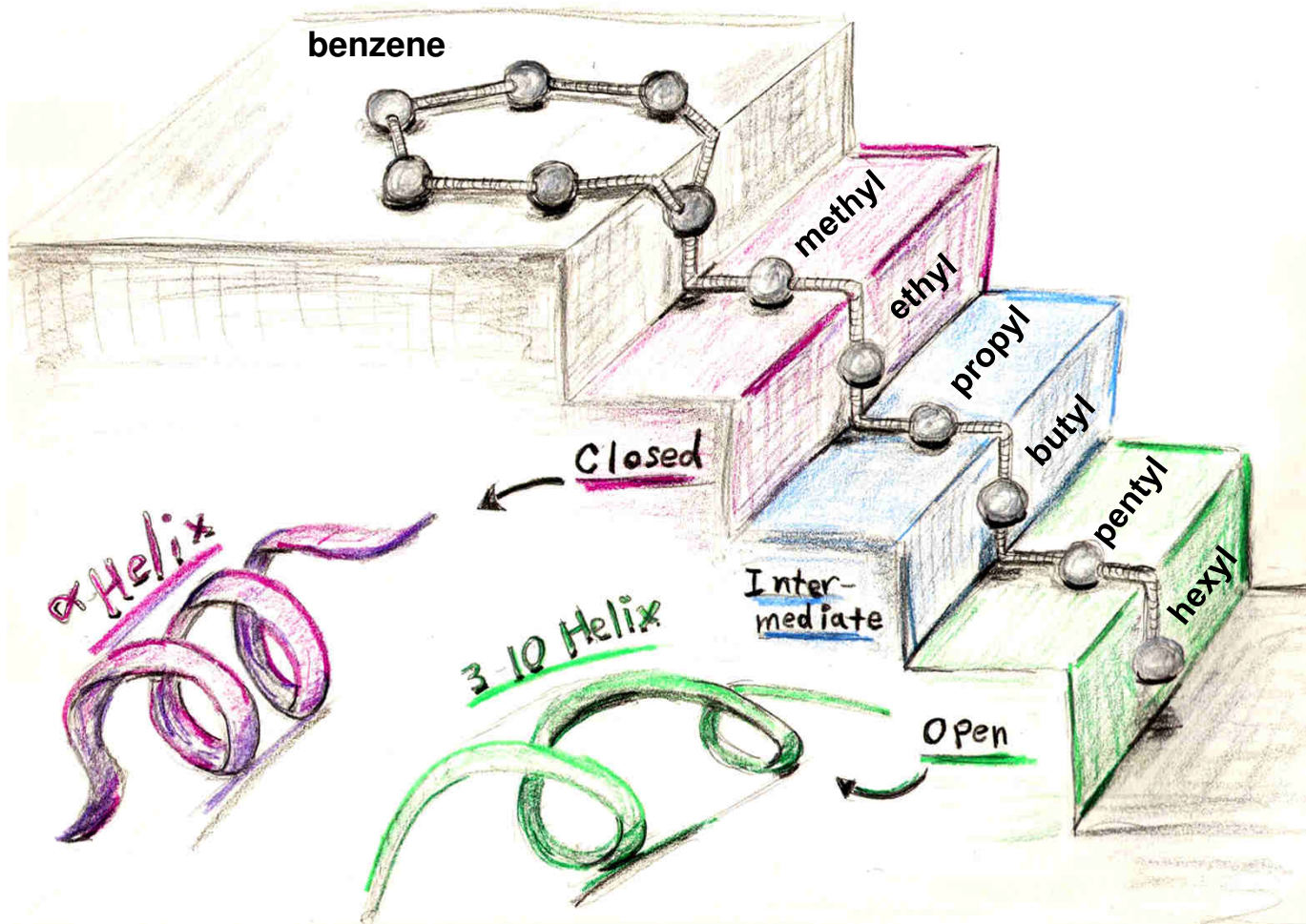
Is this response of the receptor to ligand size discreet or continuous?

Merski, Fischer, Balias, et al 2015 PNAS 112(16):5039-44

Binding affinity is less than hydrophobic burial



Merski, Fischer, Balius, et al 2015 PNAS 112(16):5039-44



Outline

- The binding event using a thermodynamic cycle
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 - Receptor flexibility
 - Receptor desolvation

Adding a Term to the Scoring Function

DOCK 3.7 scoring function:

$$E_{score} = E_{VDW} + E_{ES} + E_{lig,desol}$$

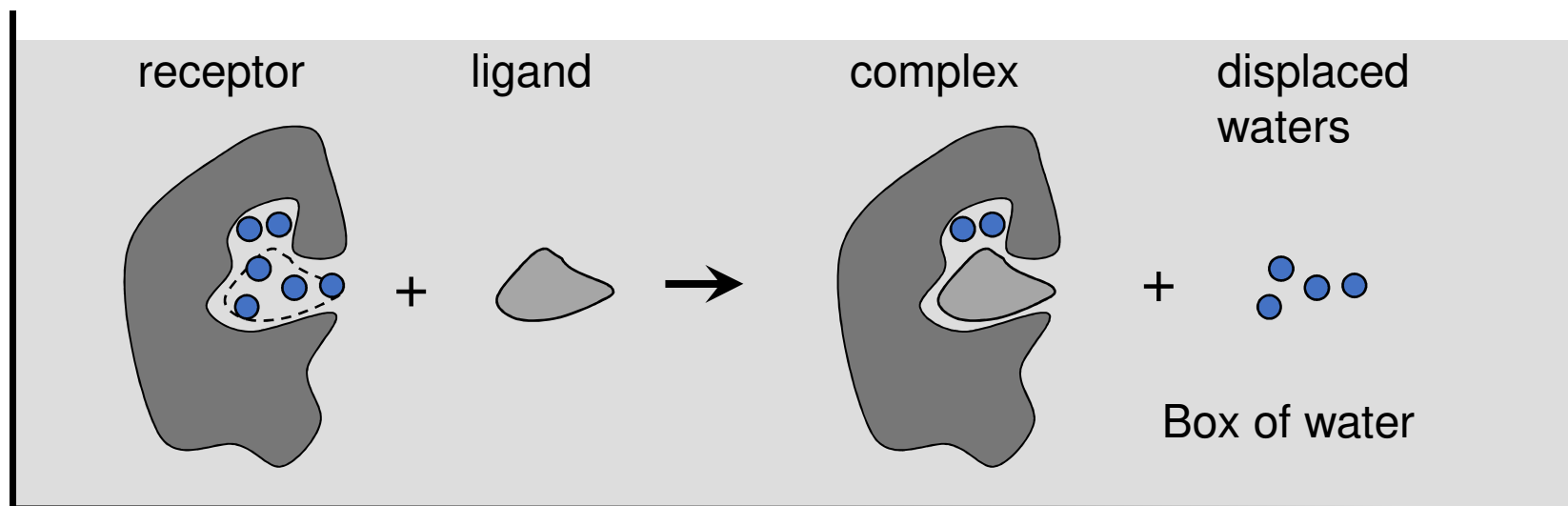
Shape complementarity

Charge complementarity

Cost of removing water from **ligand**

What's missing?

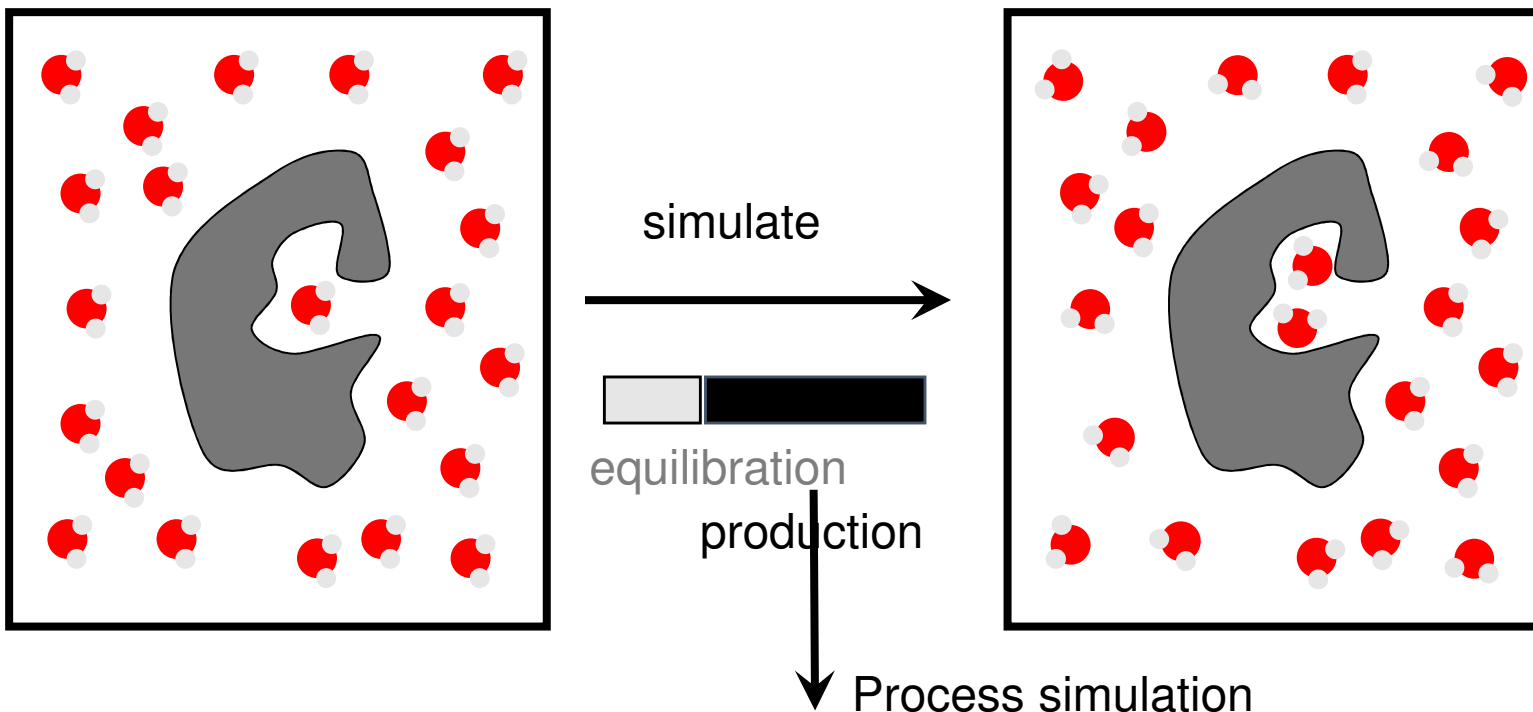
Cost of removing water from **receptor**



Meng, et al J. Comput. Chem. 1992, 13, 505– 524

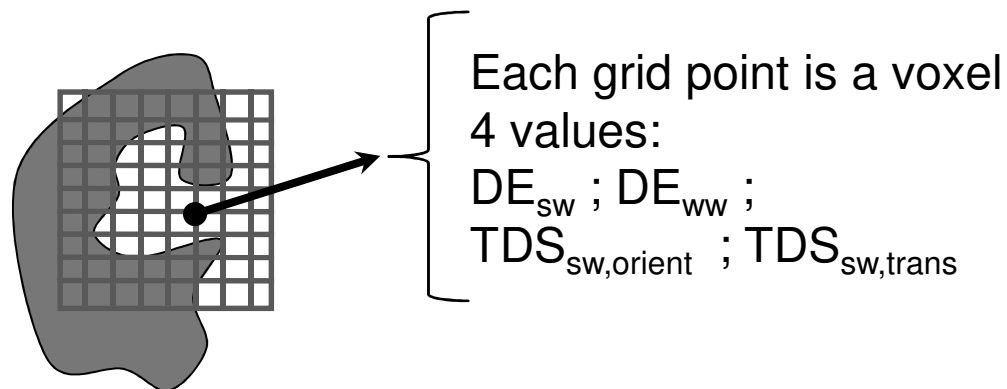
Mysinger and Shoichet J Chem Inf Model. 2010, 50(9):1561-73

Calculating Water Energetics with Receptor



GIST is implemented
in AmberTools14 in CPPtraj

Grid Inhomogeneous
Solvation Theory or GIST

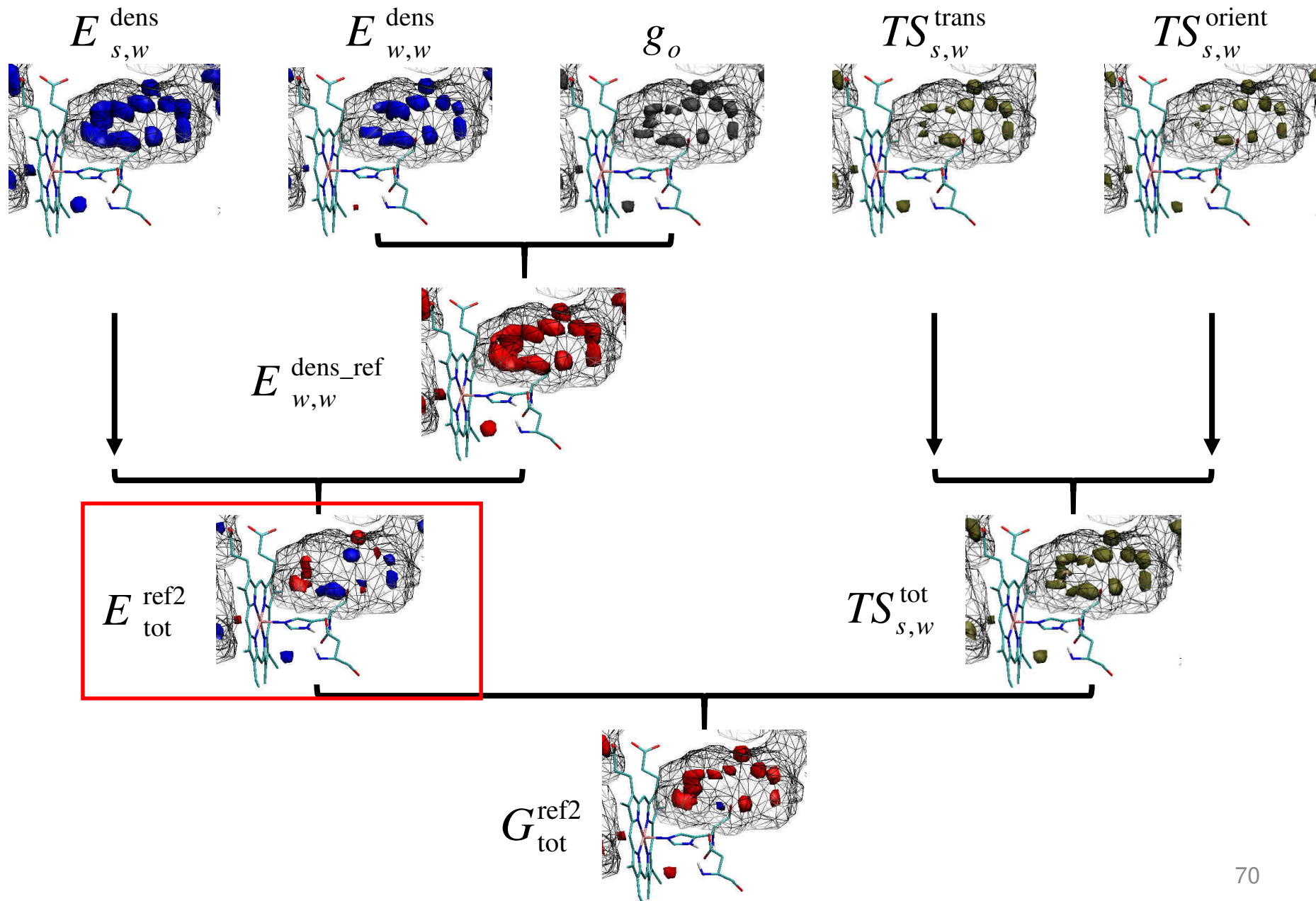


T. Lazaridis, J. Phys. Chem. B, 1998, 102, 3531-3541.

T. Lazaridis, J. Phys. Chem. B, 1998, 102, 3542-3550

C.N. Nguyen, et al, J. Chem. Phys. 2012, 137, 044101

Combining GIST Grids

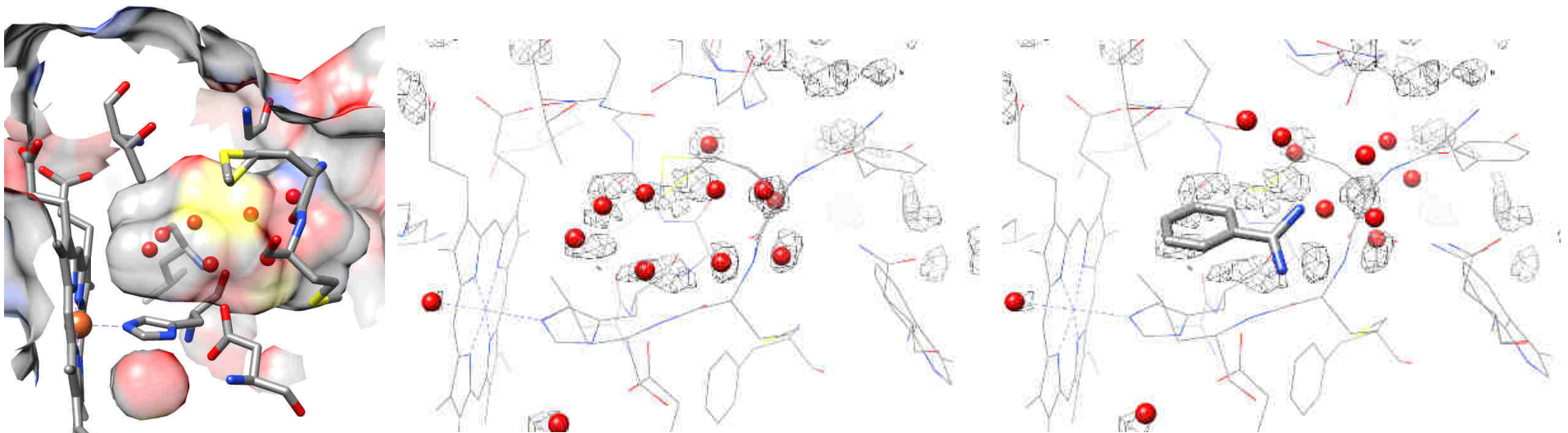


Testing DOCK+GIST Using a Model Cavity

- Model systems are simple engineered cavities
- They are dominated by 1-2 interaction terms, allowing us to disentangle various energetic contributions in docking

Cytochrome *c* peroxidase gateless mutant:

- Mutations/deletions result in solvent-exposed binding site (~8 water molecules)
- Alternative loop conformations (residues 186-194)
- Contains one anionic residue (Asp233)
- Almost exclusively binds small monocations
- Straightforward binding assay and crystallography



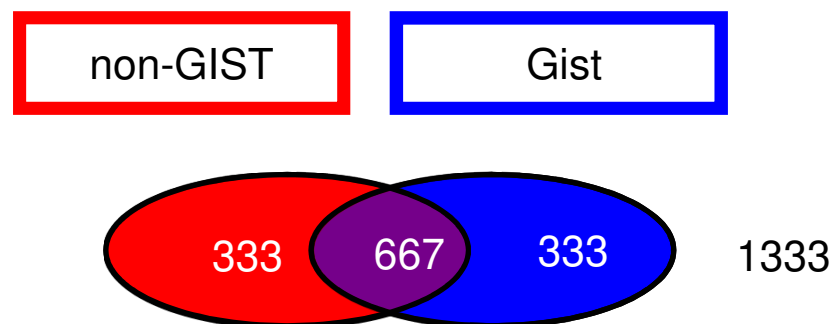
(Fischer et al., 2014; Rosenfeld et al., 2002)

Prospective Screens

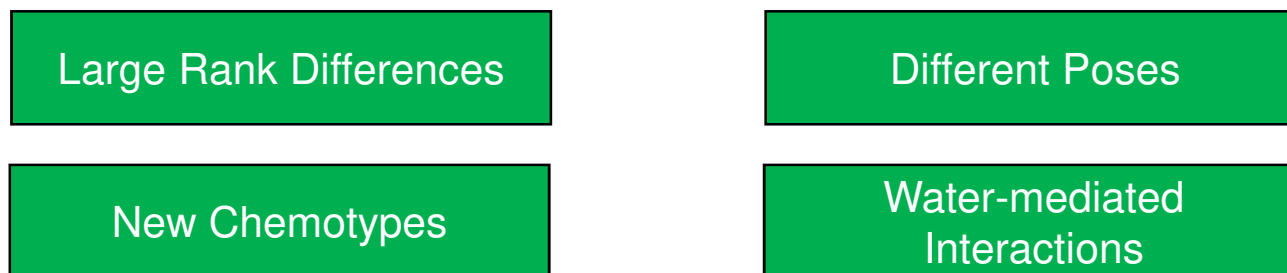
We make a computational prediction and test it experimentally

Selecting Molecules to Test with Differences

- We screened up to 1.8 million fragment molecules to the CcP-gateless mutant
 - Performed 2 screens: Non-GIST and GIST
 - We are interested in differences



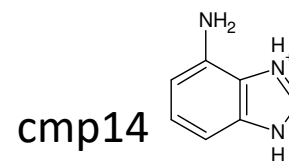
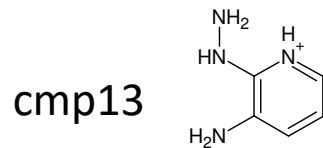
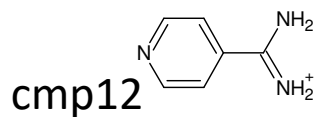
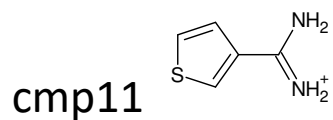
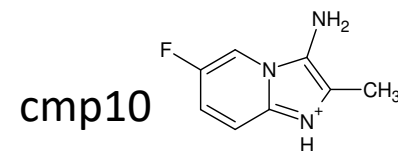
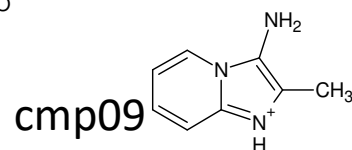
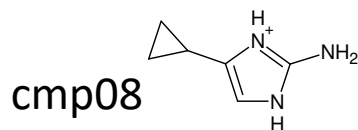
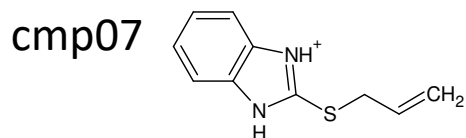
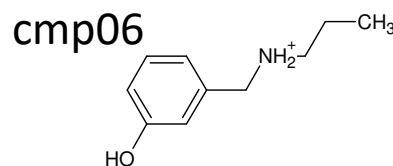
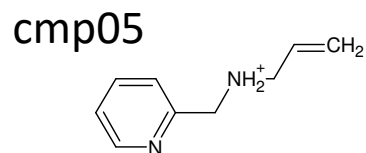
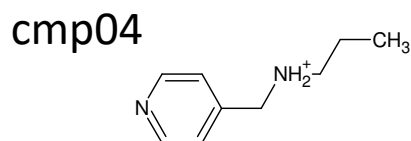
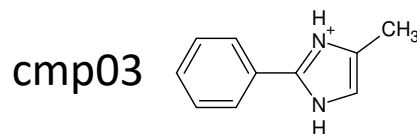
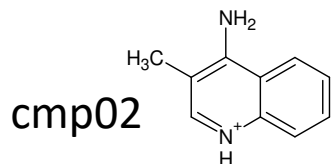
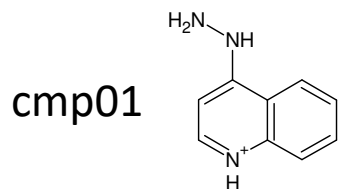
- Comparing GIST to standard screening, molecules were chosen based on:



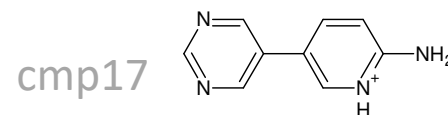
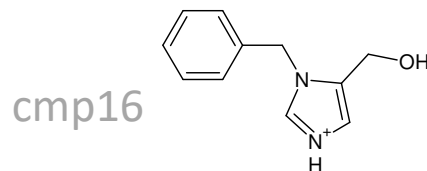
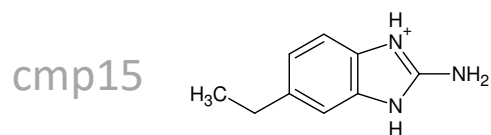
- 17 compounds (**14 pro-GIST** and **3 anti-GIST**) have been bought and tested

We Tested 17 Molecules

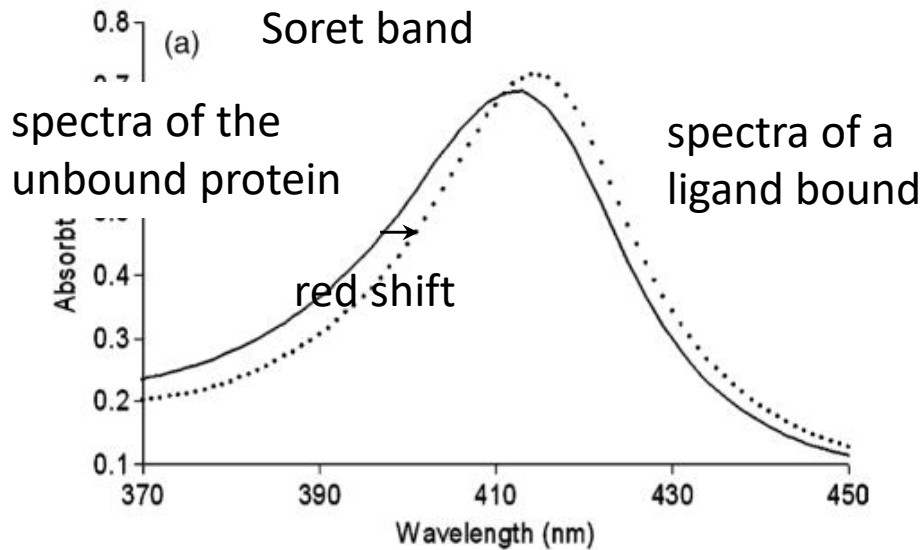
14 pro-GIST, 3 anti-GIST



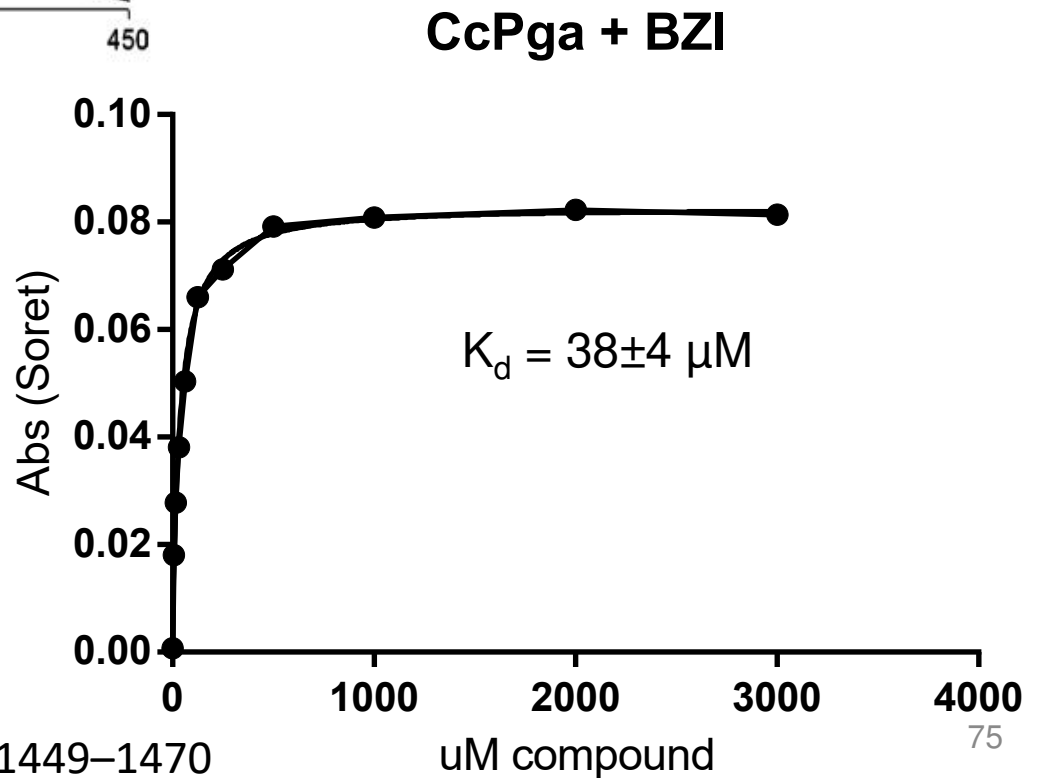
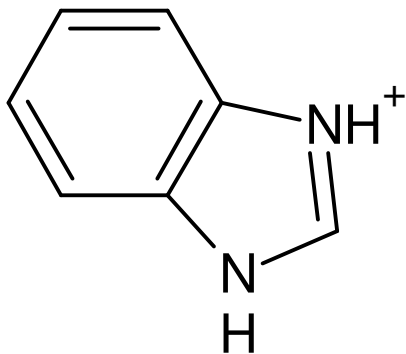
3 Anti-GIST:



Experimental Assay to Detect Binding

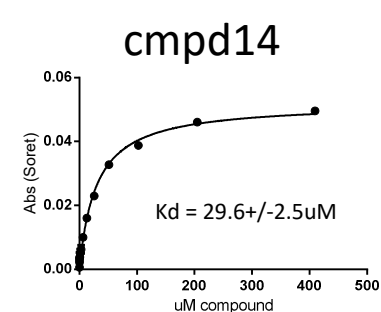
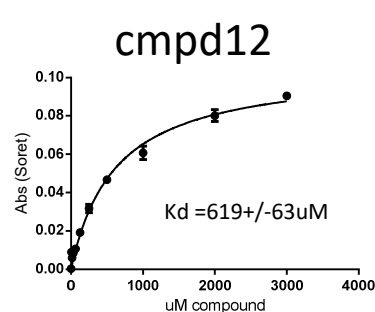
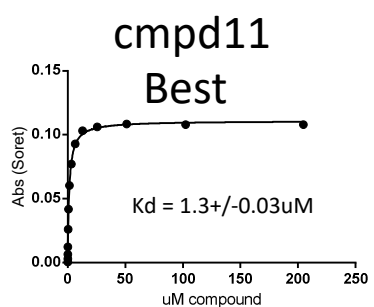
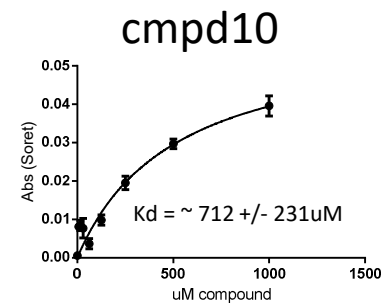
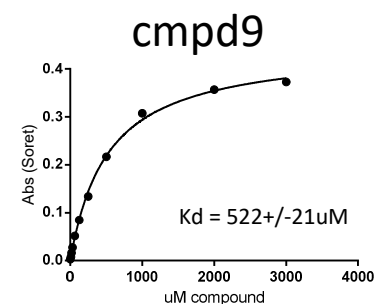
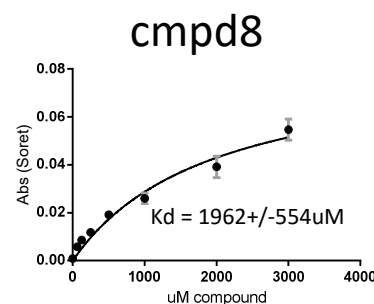
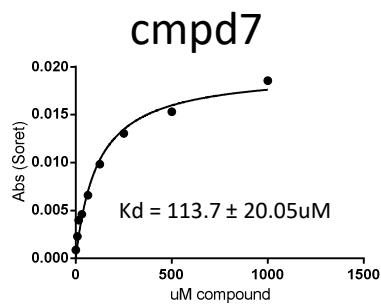
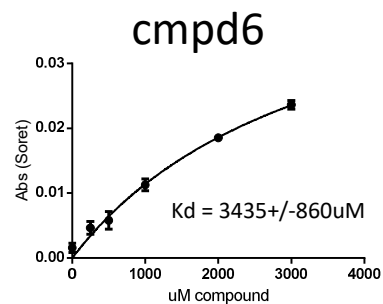
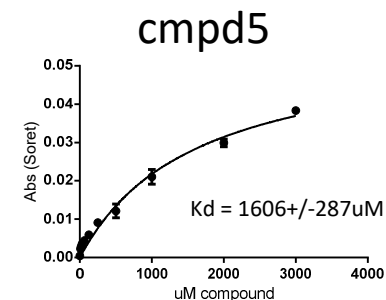
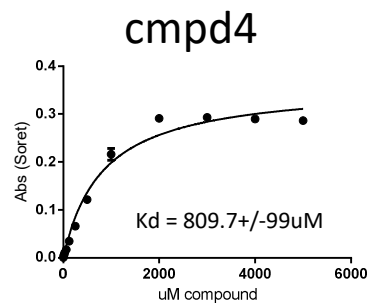
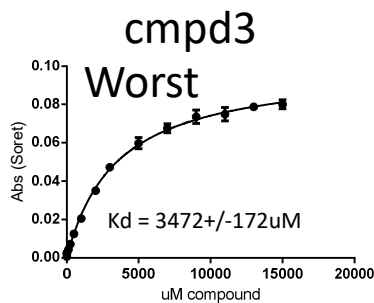
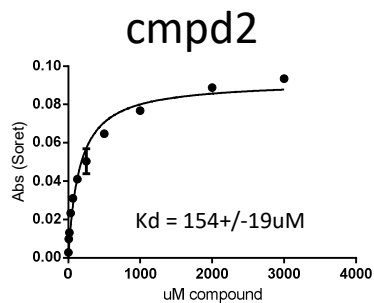


Benzimidazole (control)

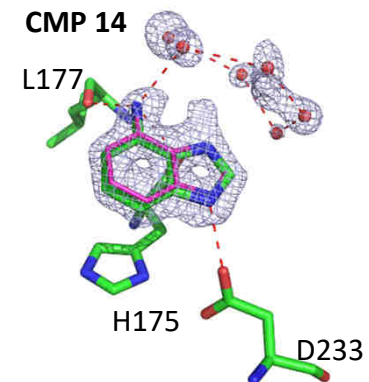
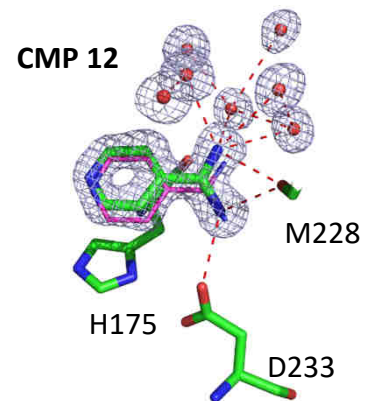
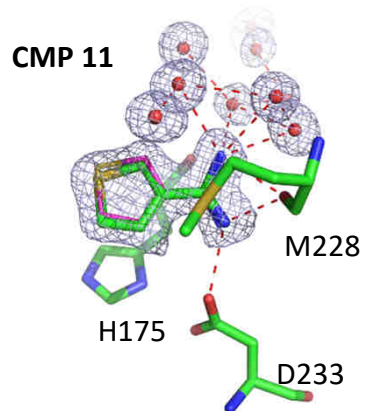
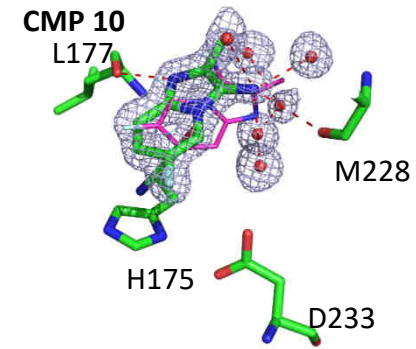
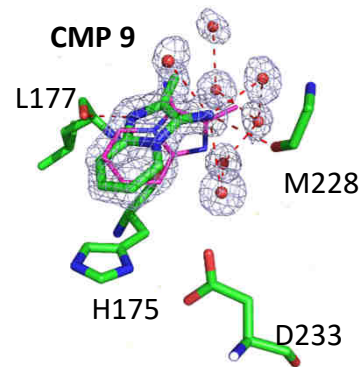
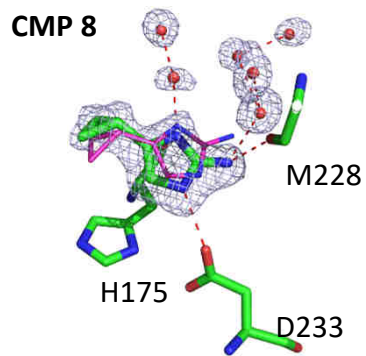
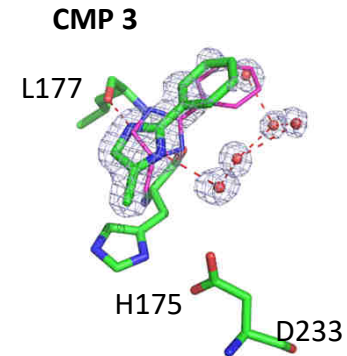
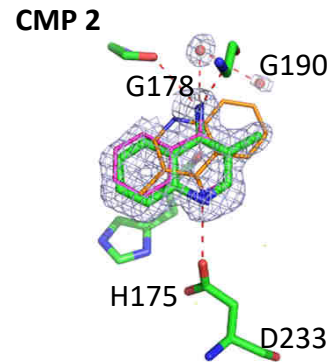
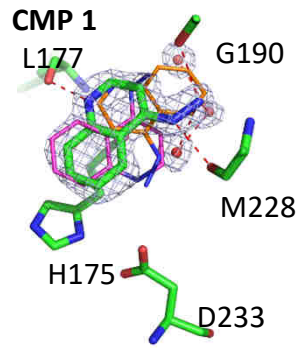


We determined Affinities for 12 molecules

Affinities range from 1 μ M to 3.5mM



9 Crystal Structures for Pose Comparisons



Prospective Summary

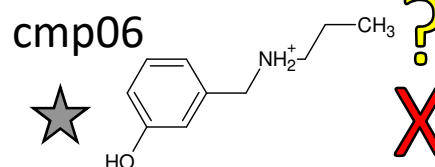
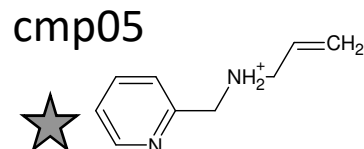
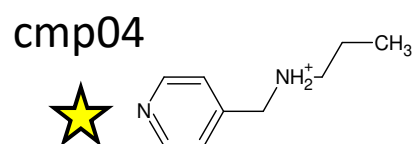
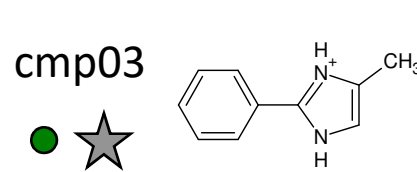
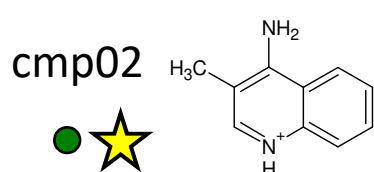
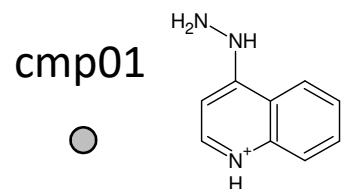
14 pro-GIST, 3 anti-GIST

★ 8 Binder <1mM

☆ 4 Binder [1mM to 4mM]

● 6 Xtal pose, dock is right

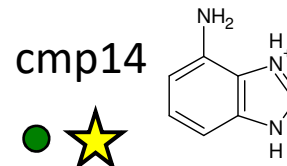
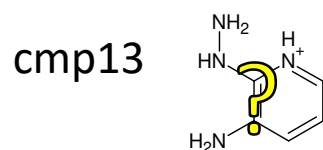
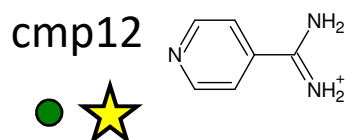
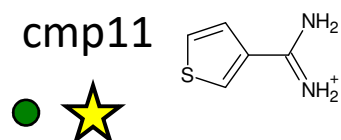
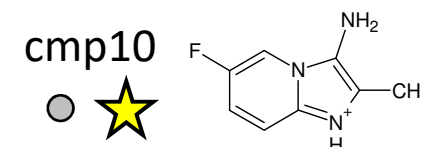
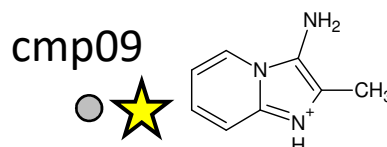
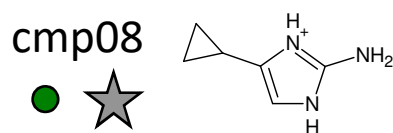
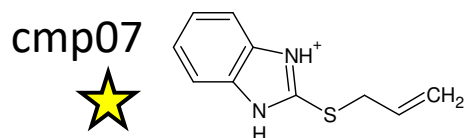
○ 3 Xtal pose, dock is wrong



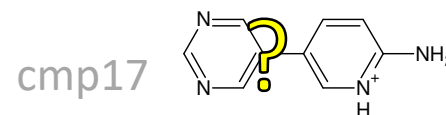
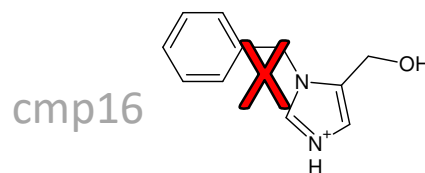
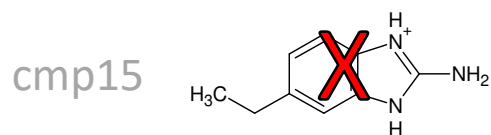
?
X

Non-determined

Non-binder



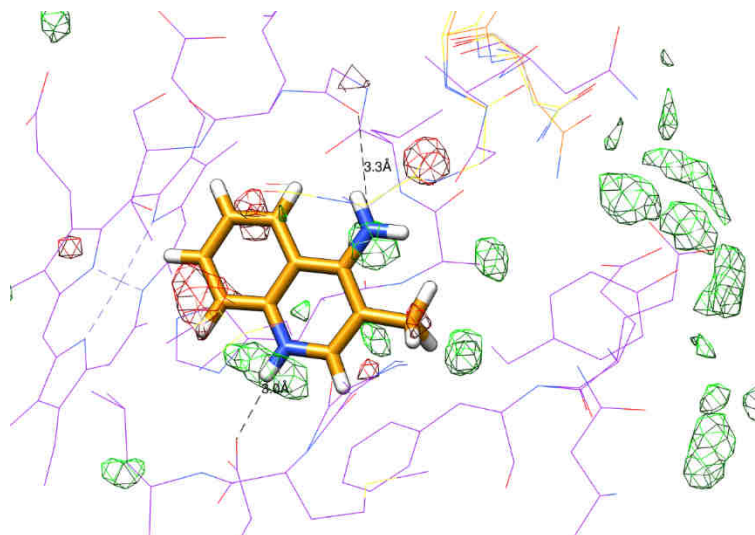
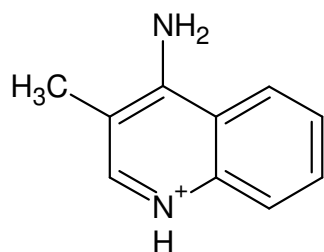
3 Anti-GIST:



Extra slides.

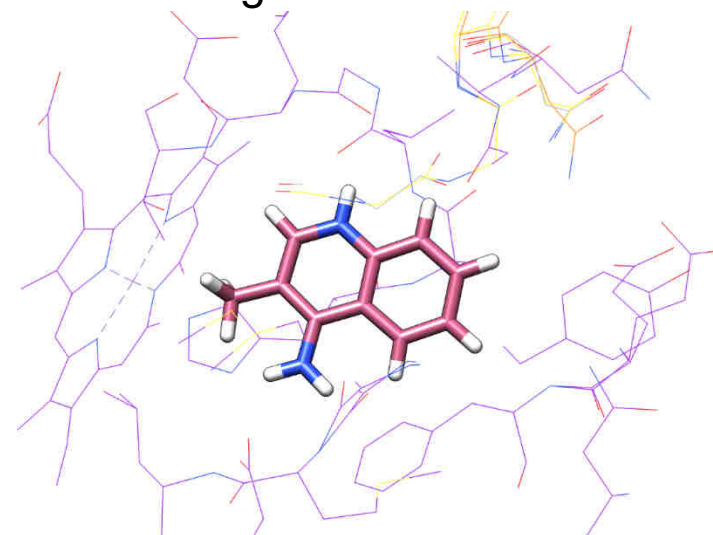
CMP 2: Binder, GIST Pose is Right

name	GIST	nonGIST	logrankdiff	rmsd
ZINC000006557114	664	740	0.047	4.62

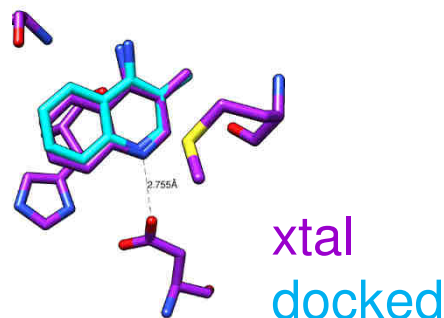
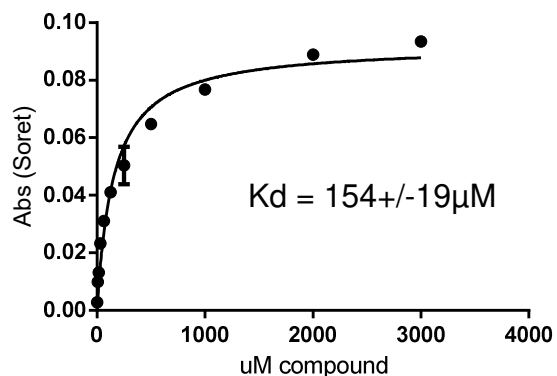


GIST

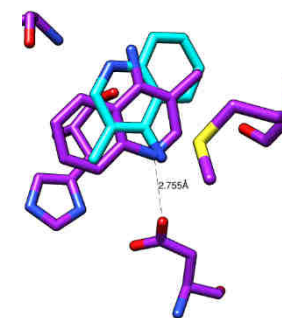
Pose change



Non-GIST



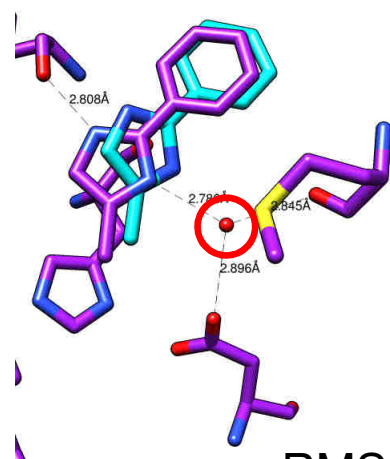
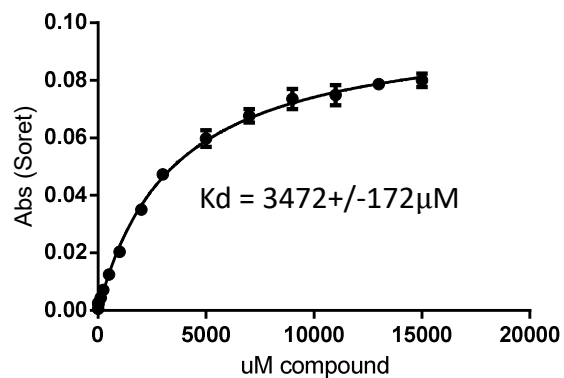
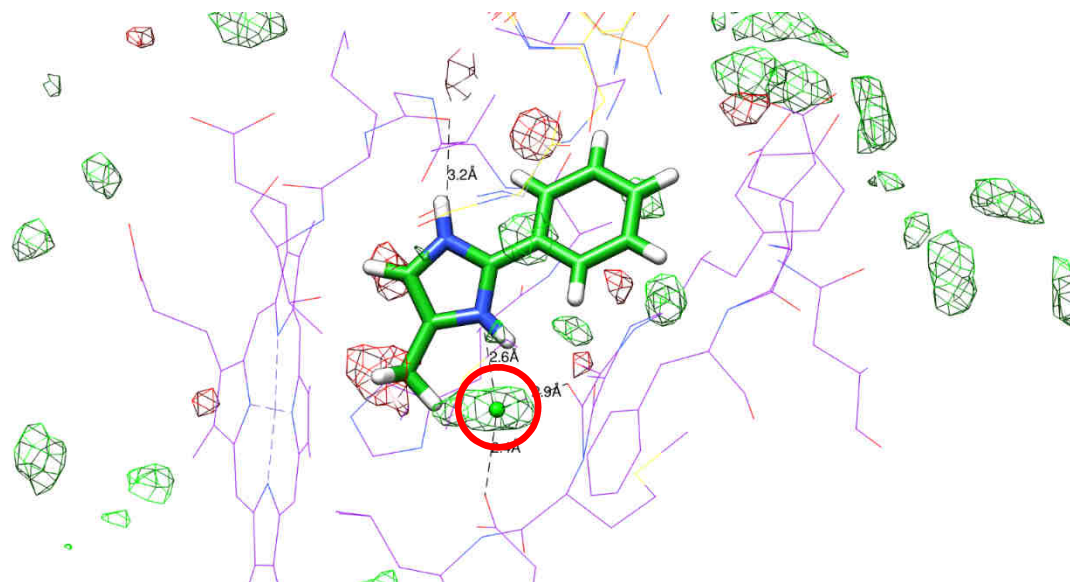
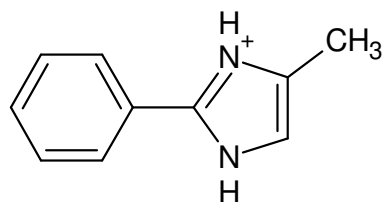
RMSD = 0.34



RMSD = 3.23

CMP 3: Water-Mediated, Weak Binder

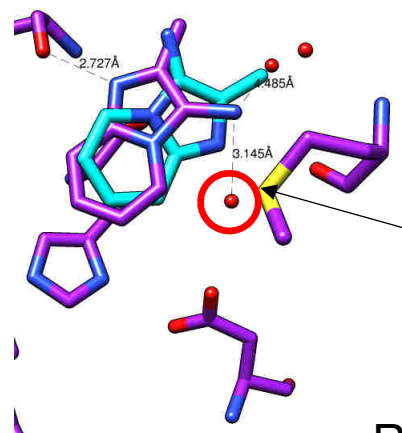
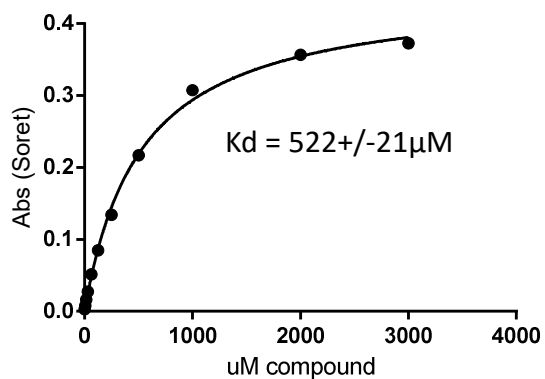
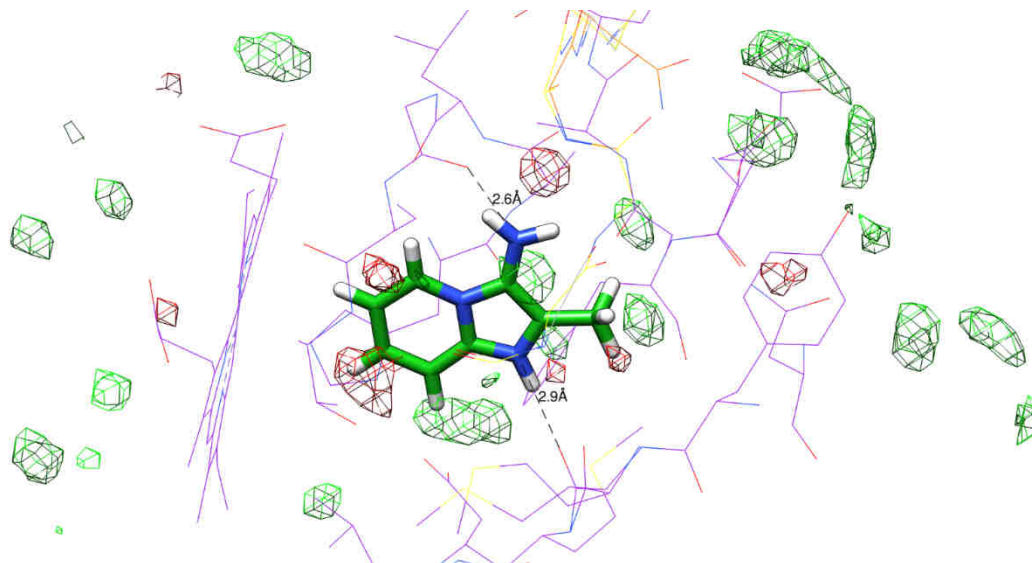
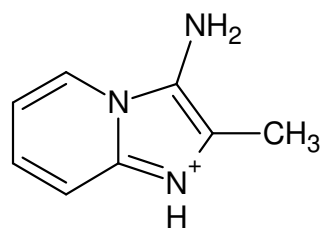
name	GIST	nonGIST	logrankdiff	rmsd
ZINC000004705523	13	249	1.28	0



RMSD = 1.32

CMP 9: binder, wrong pose

name	GIST	nonGIST	logrankdiff	rmsd
ZINC000020357620	98	745	0.88093	0

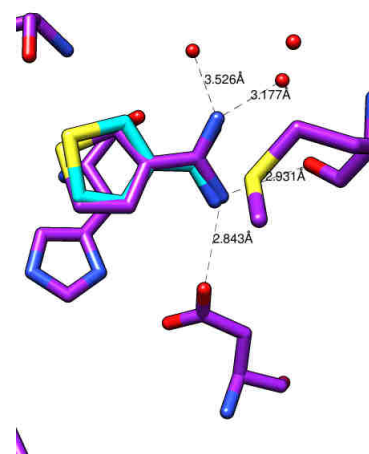
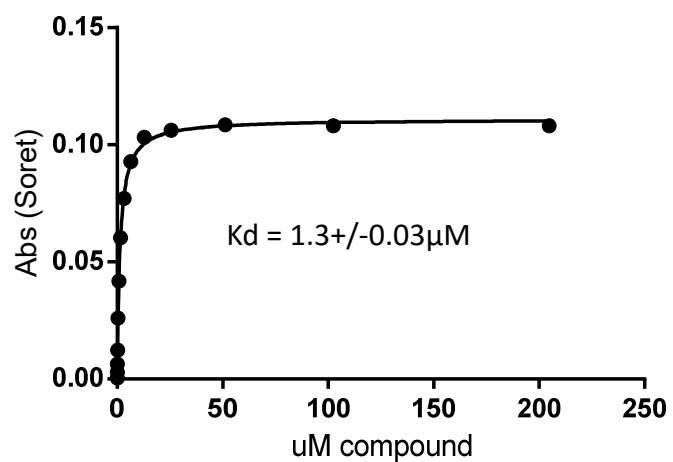
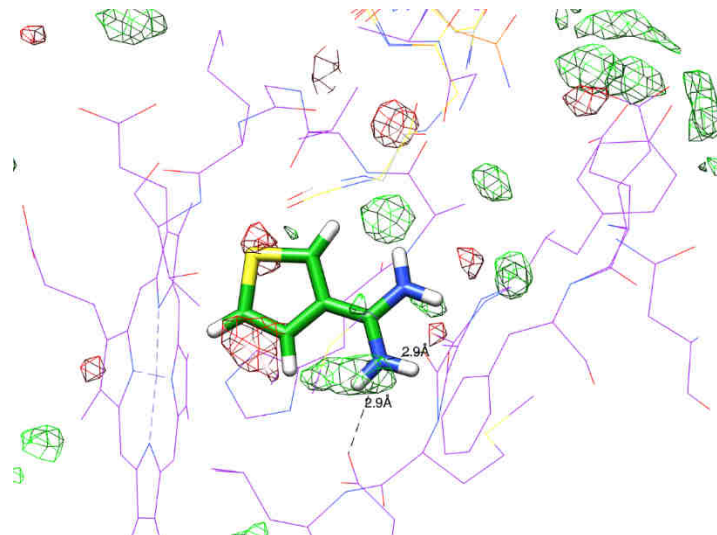
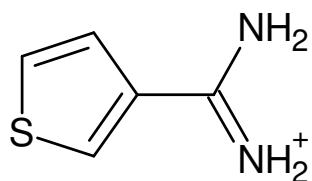


Failed to
predict water-
mediated
interaction

RMSD = 1.73

CMP 11: best binder

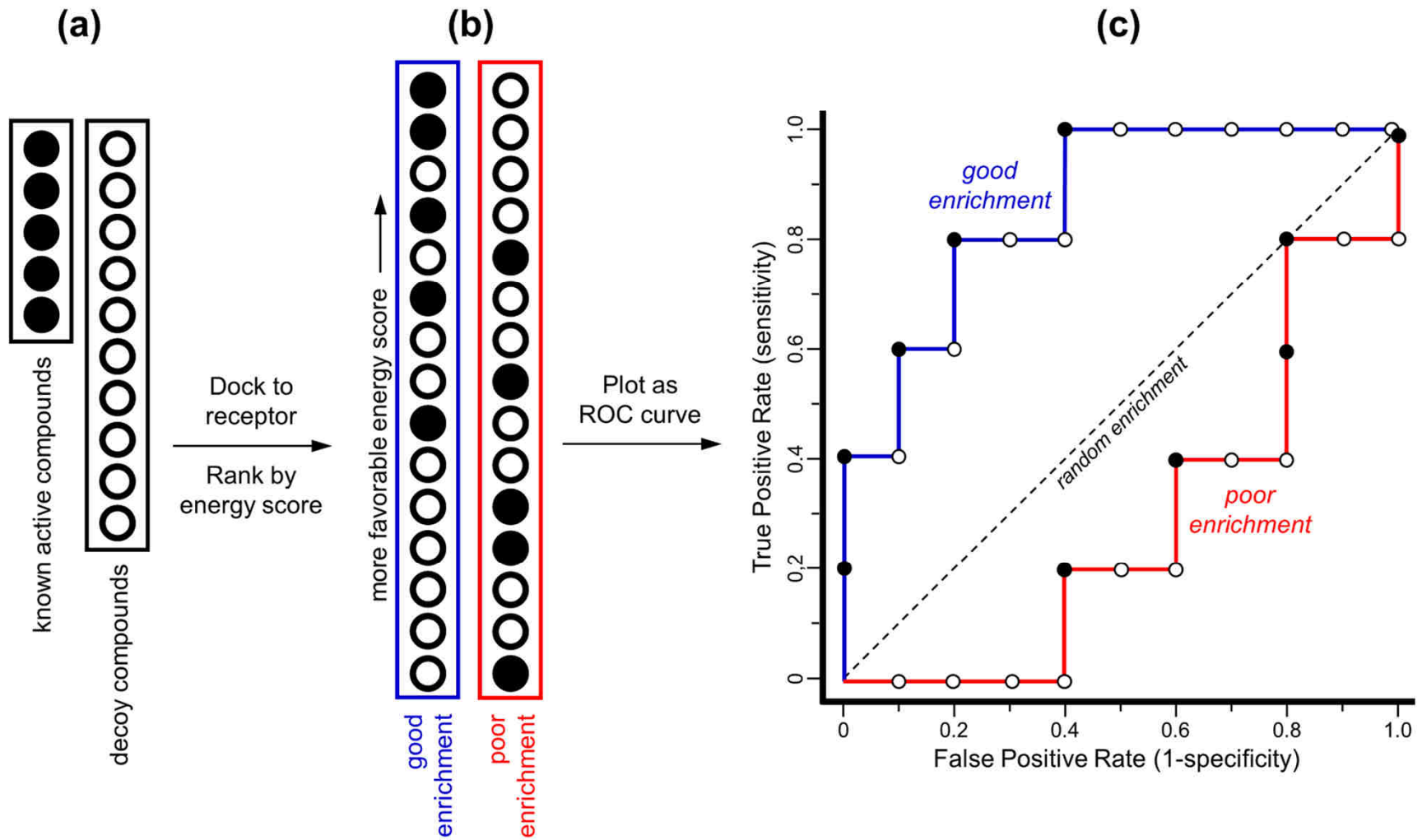
name	GIST	nonGIST	logrankdiff	rmsd
ZINC000000161834	358	1212	0.52962	0



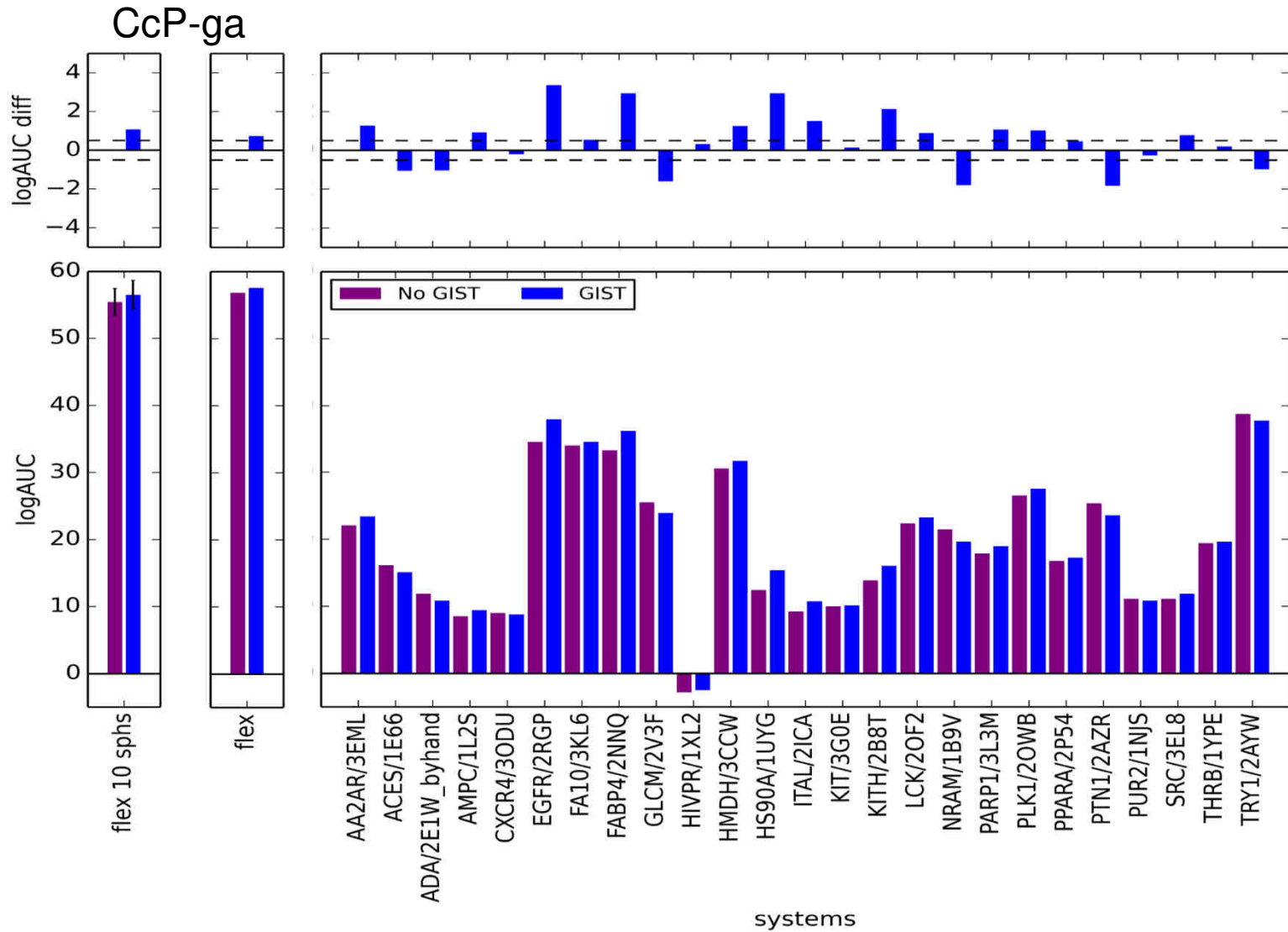
RMSD = 0.4448

Retrospective testing

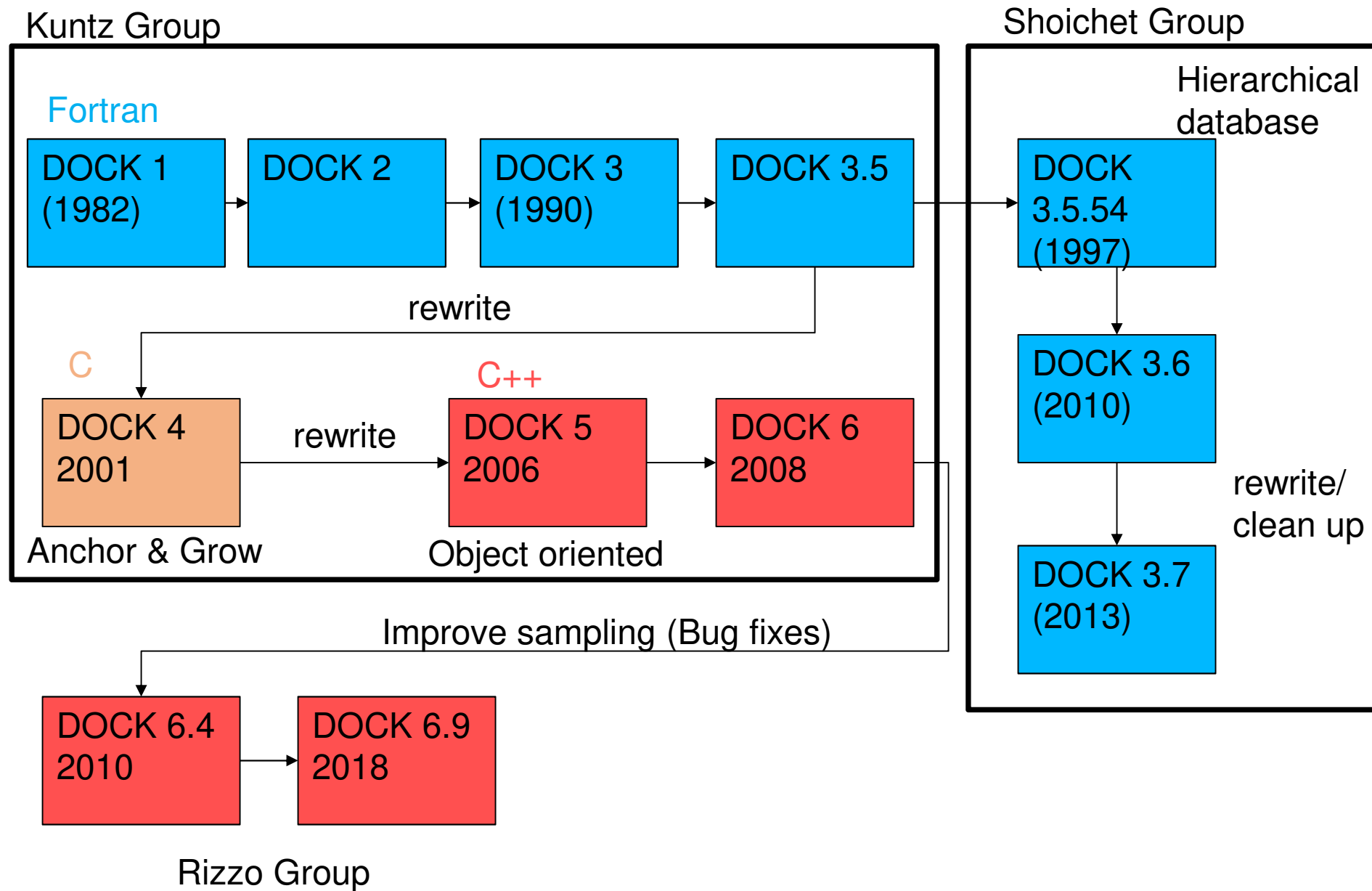
Enrich knowns over Decoys



GIST has little effect on retrospective analysis of 25 DUDE systems



DOCK: A History



Docking is Important in Ligand Discovery

Applications of docking

- Virtual Screening: given a protein and database of molecules find those that bind.
- Pose prediction: given a molecule and a protein predict how that they bind

Docking Tasks

- Sampling
 - generate all the possibilities including finding the correct geometry
- Scoring
 - of all the possibilities, rank the correct pose first
 - also, rank the binders better than decoys
- Balance of speed and accuracy, docking has to be fast.

How to evaluate docking methods

- Pose reproduction, reproduce the crystallographic poses
- Enrichment calculations, make sure ligand found in the top of the rank orders lists.
- Prospective testing on model cavities, make a predication, and test it!

It's a miracle docking ever works

DOCK 3.7 scoring function:

$$p_A(g) = \sum_{b \in rec} \frac{A_b}{r^{12}}$$

$$E_{score} = E_{VDW} + E_{ES} + E_{lig,desol}$$

Charge
complementarity

What's missing?

Shape
complementarity

Cost of removing
water from **ligand**

To make it fast we precompute
potentials, ligand properties,
displacement

$$E_{VDW} = \sum_{a \in lig} A_a * p_A(r_a) - B_a * p_B(r_a)$$

interpolate

$$E_{ES} = \sum_{a \in lig} q_a * \Psi(r_a)$$

$$E_{lig,desol} = \sum_{a \in lig} s_a * D(r_a)$$

Cost of removing
water from **receptor**

$$E_{score} = E_{VDW} + E_{ES} + E_{lig,desol} + E_{rec,desol} \\ + E_{lig,wat} + E_{rec,conf} + E_{lig,strain}$$

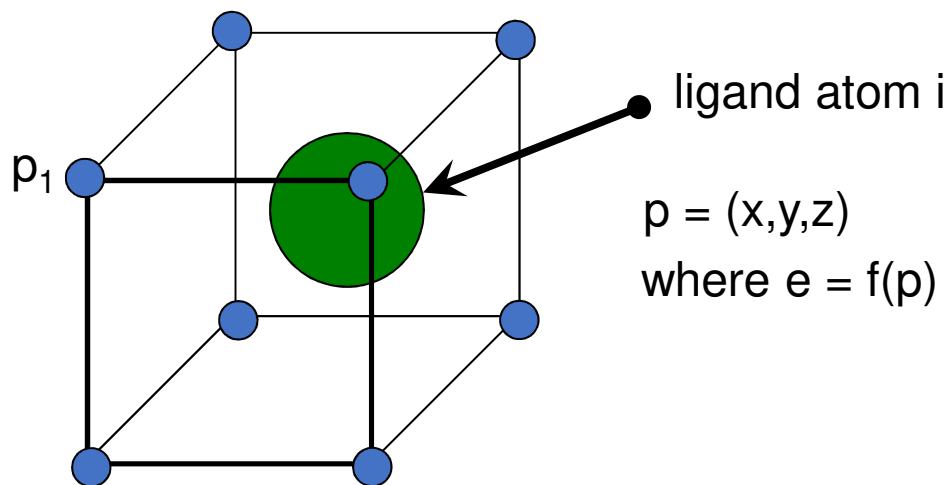
Meng, et al J. Comput. Chem. 1992; Mysinger and Shoichet JCIIM, 2010; Fischer et al, Nature Chem. 2014; Balius et al, PNAS, 2017

$$E = \sum_{i \in L} \left(\sqrt{A_{i,i}} \sum_{j \in R} \frac{\sqrt{A_{j,j}}}{r_{i,j}^a} - \sqrt{B_{i,i}} \sum_{j \in R} \frac{\sqrt{B_{j,j}}}{r_{i,j}^b} + 332q_i \sum_{j \in R} \frac{q_j}{Dr_{i,j}} \right)$$

$$G_{av}(p) = \sum_{l \in R} \frac{\sqrt{A_{l,l}}}{r_{p,l}^a}$$

$$G_{rv}(p) = \sum_{l \in R} \frac{\sqrt{B_{l,l}}}{r_{p,l}^b}$$

$$G_{es}(p) = 332 \sum_{l \in R} \frac{q_l}{Dr_{p,l}}$$



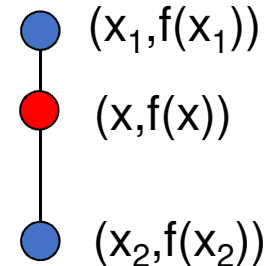
http://dock.compbio.ucsf.edu/DOCK_6/dock6_manual.htm#Grid

$$E \approx \sum_{i \in L} \left(\begin{array}{l} \sqrt{A_{i,i}} \text{interp}[G_{av}(p_1), \dots, G_{av}(p_8)] \\ - \sqrt{B_{i,i}} \text{interp}[G_{rv}(p_1), \dots, G_{rv}(p_8)] \\ + 332q_i \text{interp}[G_{es}(p_1), \dots, G_{es}(p_8)] \end{array} \right)$$

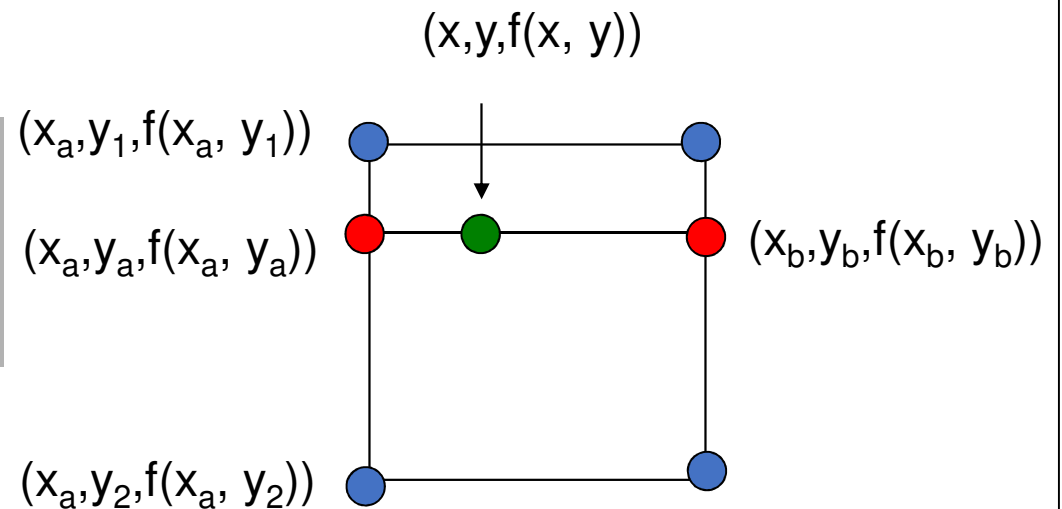
Interpolation

linear

$$f(x) \approx \frac{(x - x_1)f(x_2) + (x_2 - x)f(x_1)}{(x_2 - x_1)}$$



bilinear: Perform 3 linear Interpolations: 2 to calculate red (from cyan); and 1 to calculate green (from red)



Trilinear: for a cube, perform 7 linear interpolations: 4 to calculate red (from the cyan); 2 to calculate green (from red); and 1 to calculate the atomic approximation (from green)

ROC curves

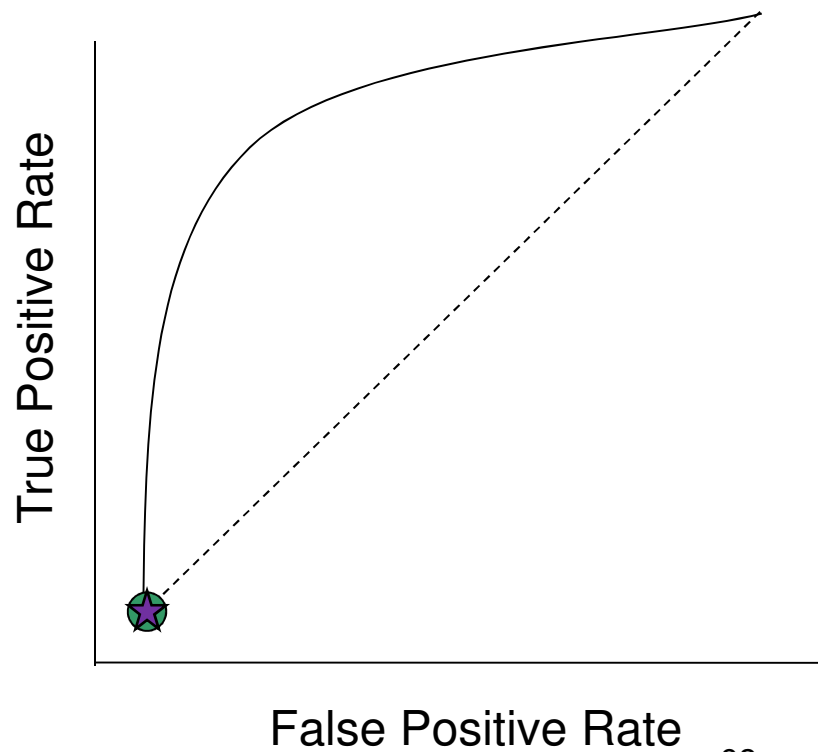
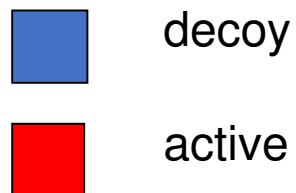
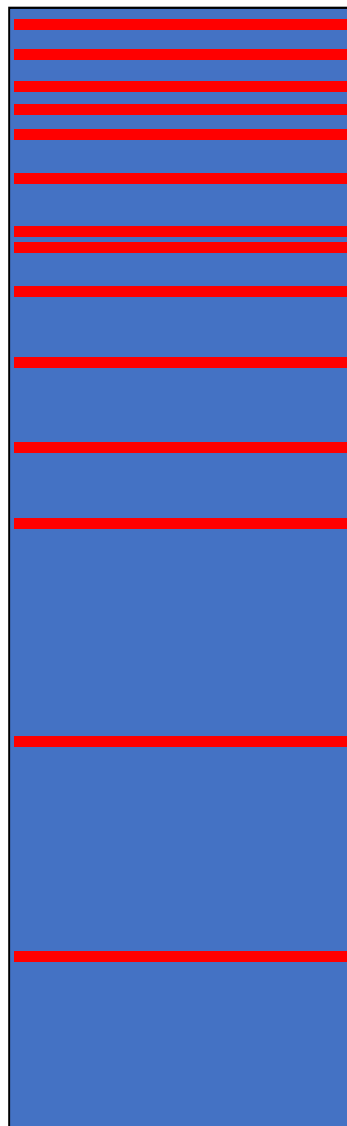
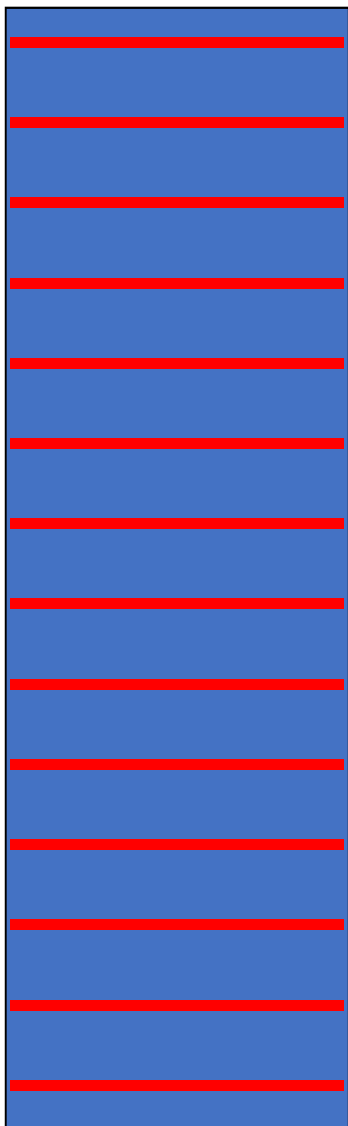
$$TP_{Rate} = Se_{subset} = \frac{ligands_{selected}}{ligands_{total}}$$

$$FP_{Rate} = (1 - Sp)_{subset} = \frac{decoys_{selected}}{decoys_{total}}$$

Se - Sensitivity, Sp - Specificity

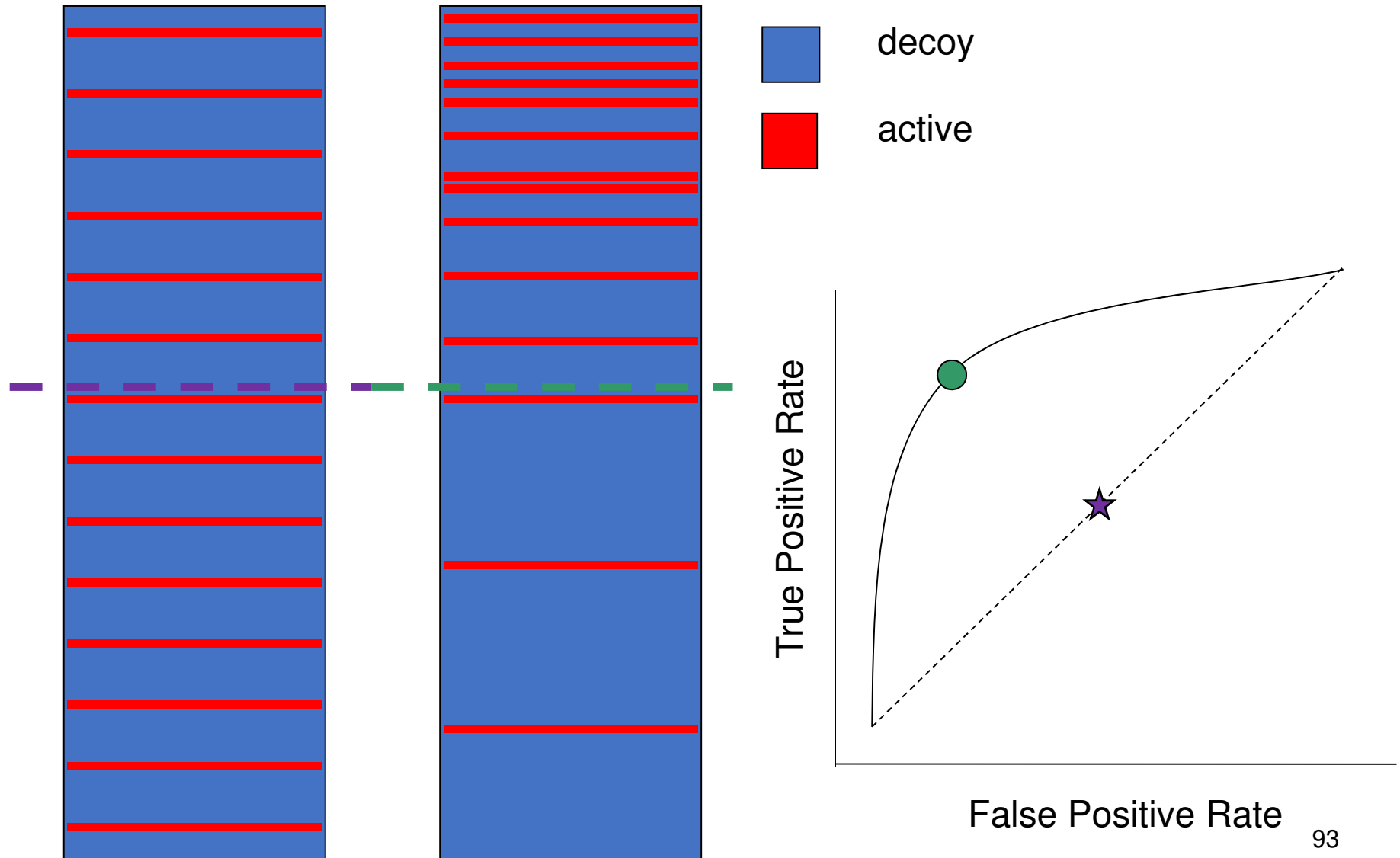
Small molecule
database seed
with actives

ROC Curves



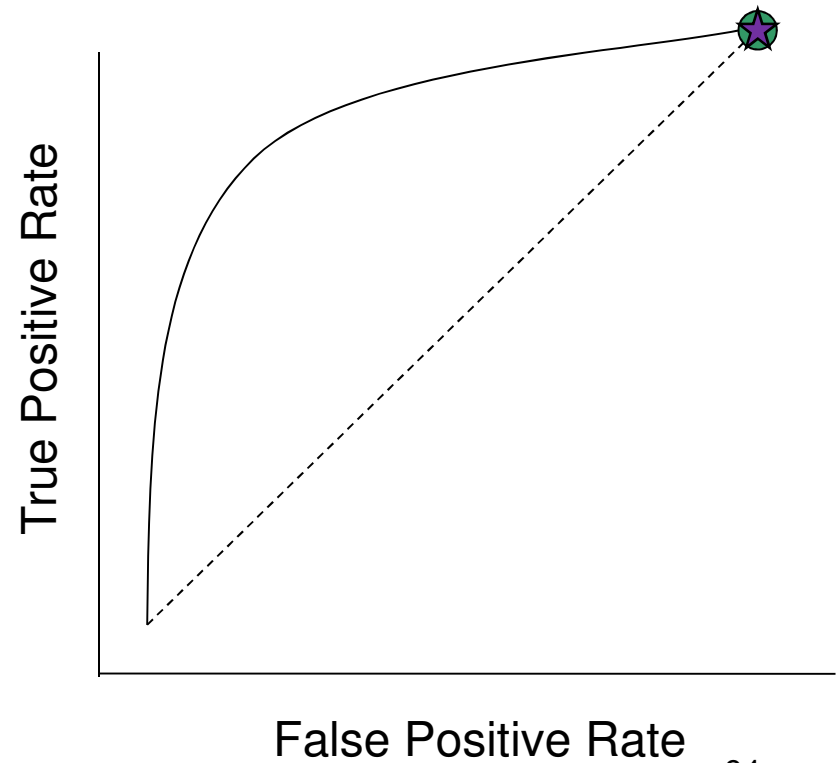
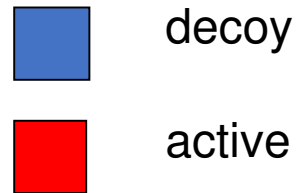
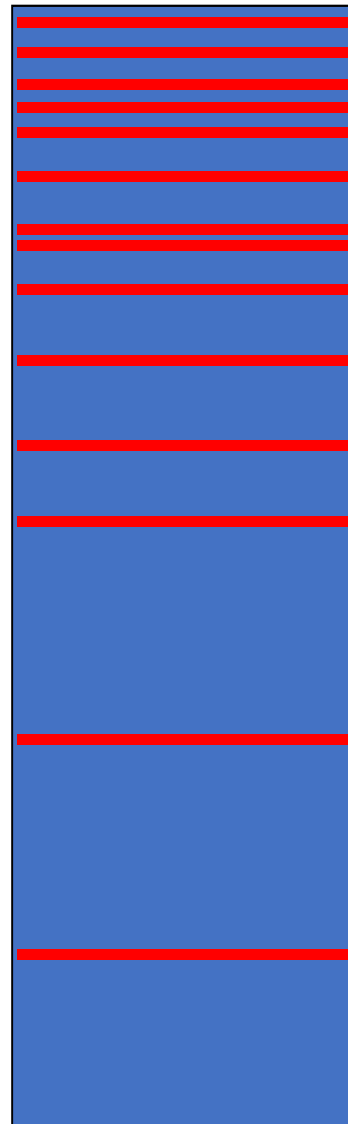
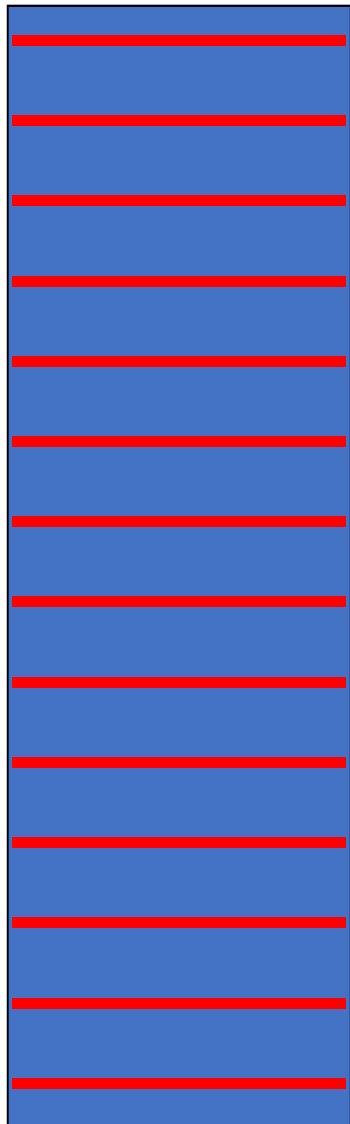
Small molecule
database seed
with actives

ROC Curves



Small molecule
database seed
with actives

ROC Curves



Example ROC Curves

